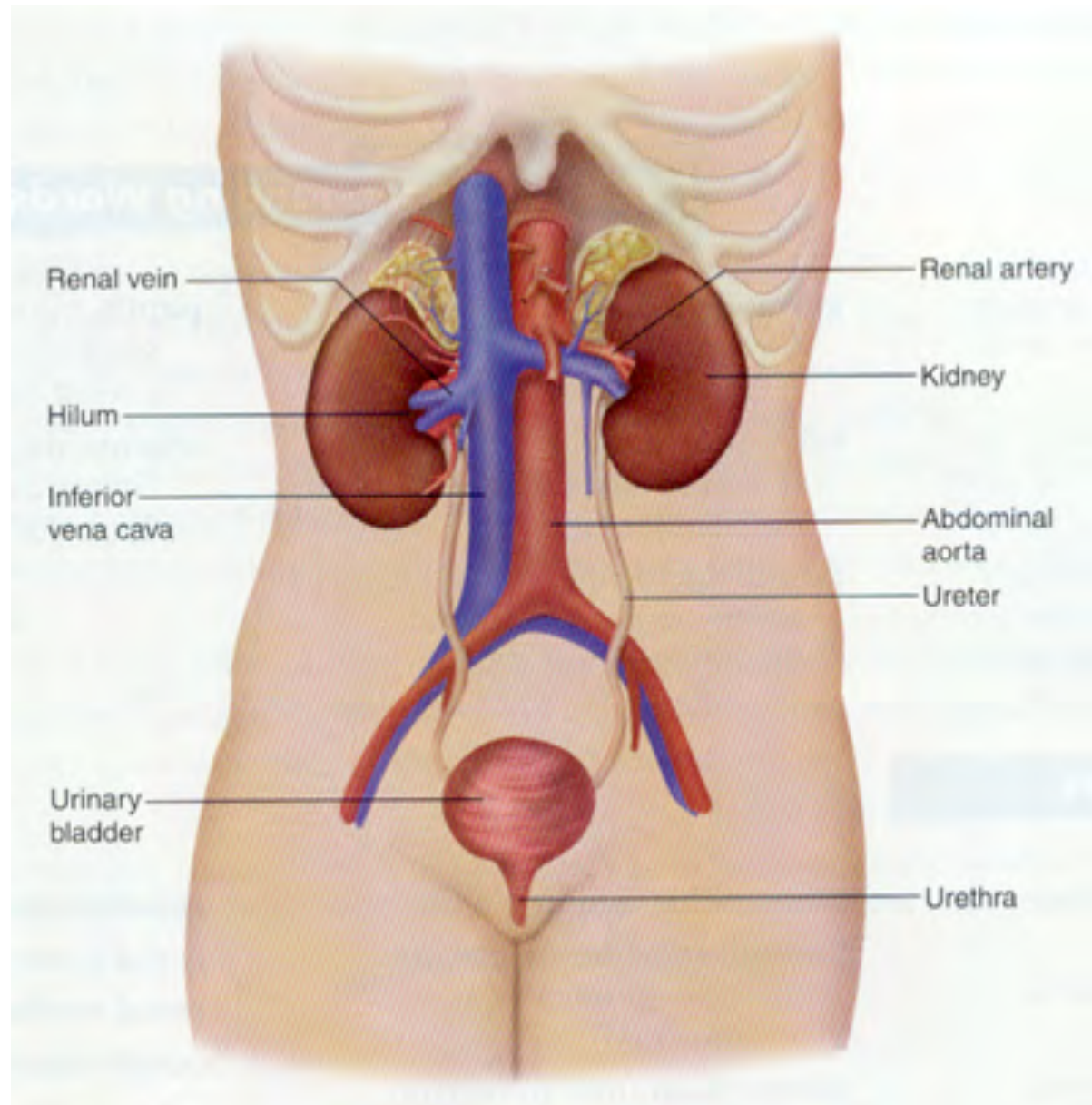


Spring Elective

2018 Spring

Genito-Urinary system



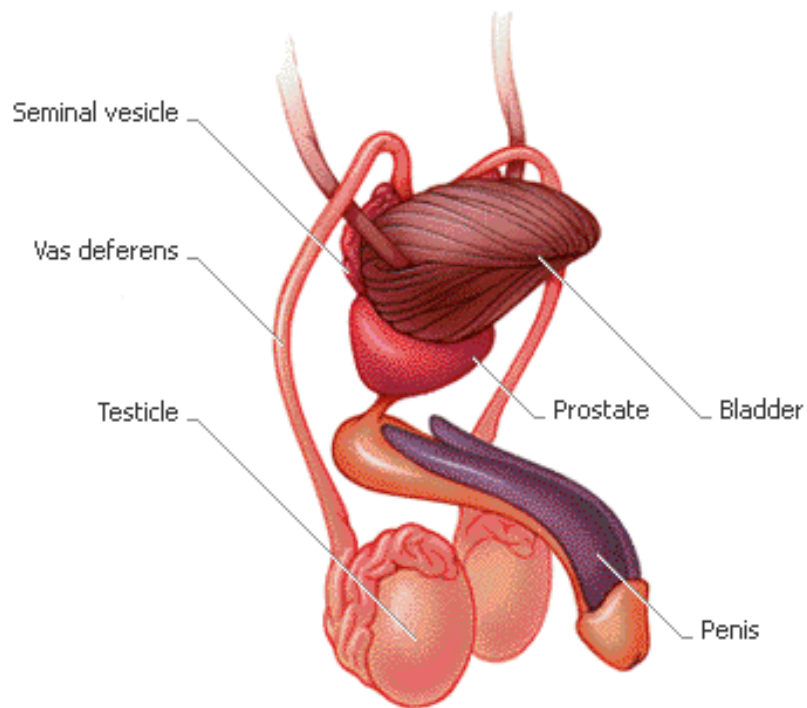
Human Uterus,  
fallopian tubes,  
Ovaries



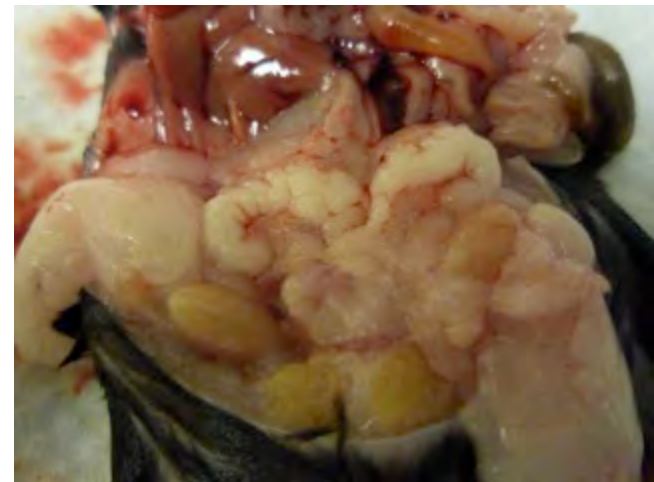
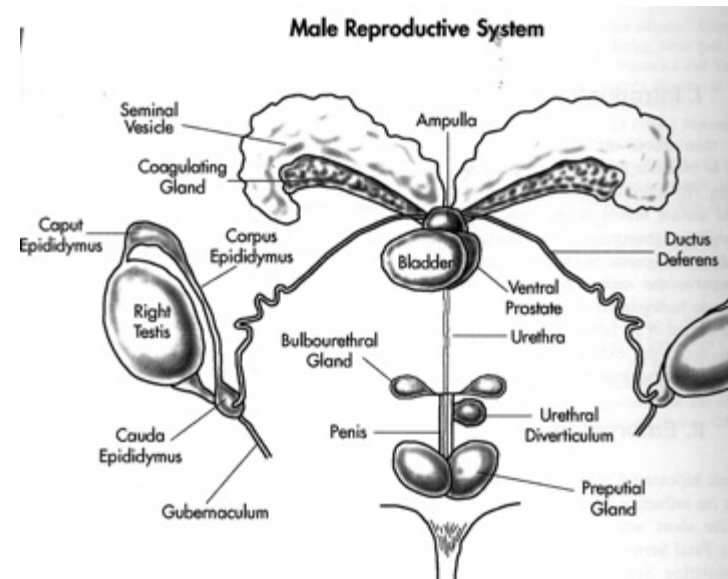
Mouse bi-cornuate uterus,  
tubes and ovaies



# Human Male Reproductive system



# Mouse



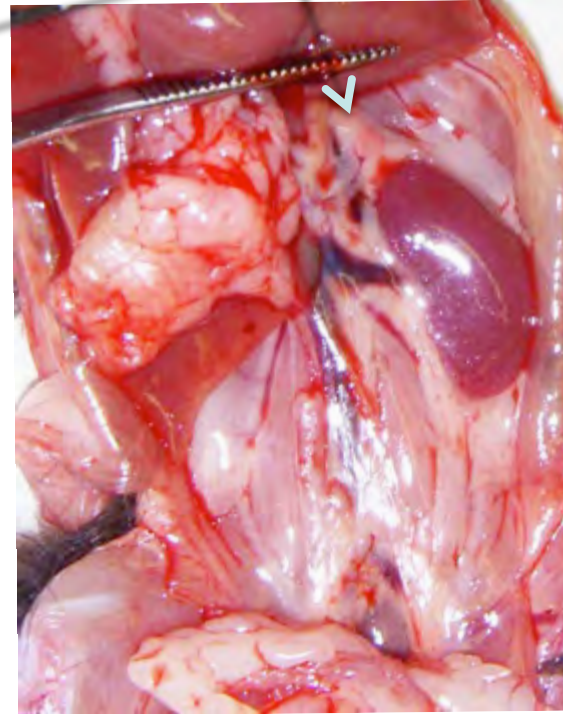
The mouse seminal vesicles are very prominent



# Human kidneys, ureters, bladder, prostate













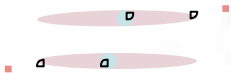
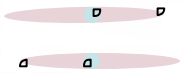
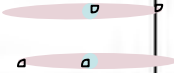
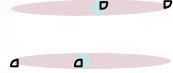


## Mouse kidneys, adrenals



Alan J. Davidson et al., 2008.  
Mouse kidney development

## TYPES OF CAPILLARIES

<b>Continuous</b>	No gaps between endothelial cells	Basal Lamina present	Most common
			
<b>Fenestrated</b>	Endothelial cells separated by gaps of 60-80 nm	Basal Lamina present	Kidney Intestine Endocrine organs
			
<b>Fenestrated2</b>	Gaps	THICK BASAL lamina	Glomeruli of kidney
			
<b>Sinusoidal</b>	Gaps	Discontinuous Basal Lamina	Liver Spleen Bone Marrow
			



# The Nephron

- Filtration:  
Glomerulus

- Secretion & Reabsorption:  
Proximal tubule, loop of Henle, and  
Distal tubule



# Nephron segments

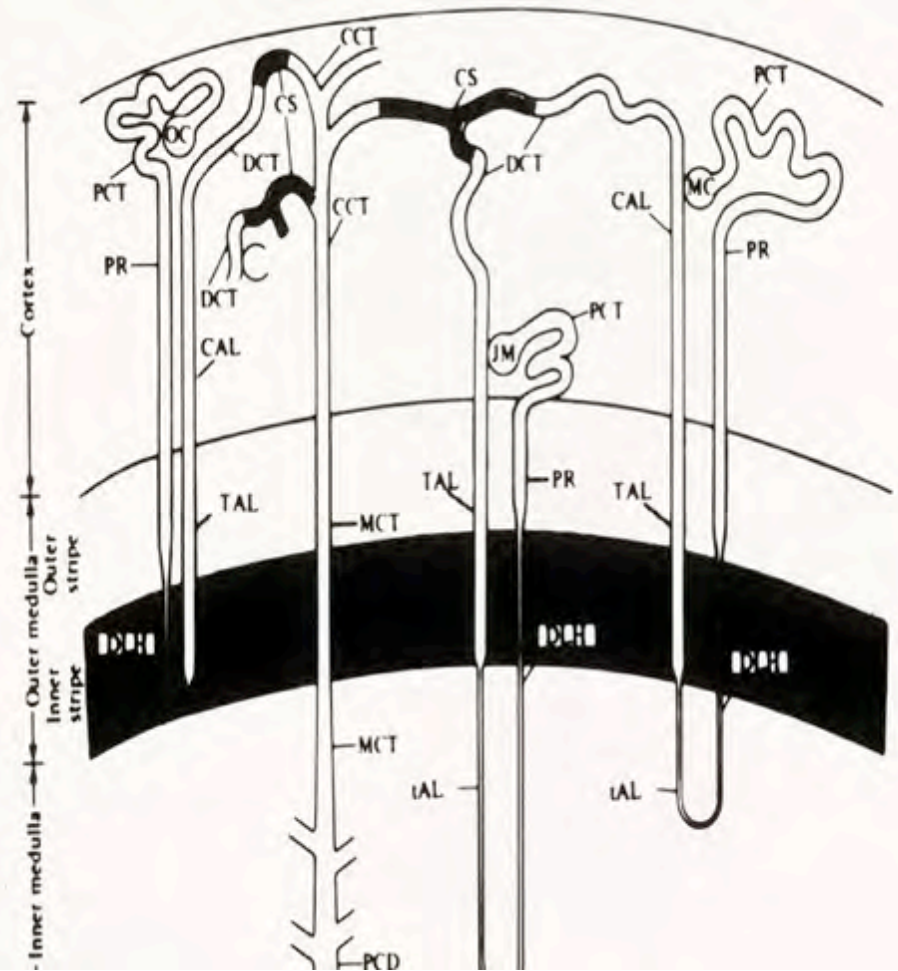
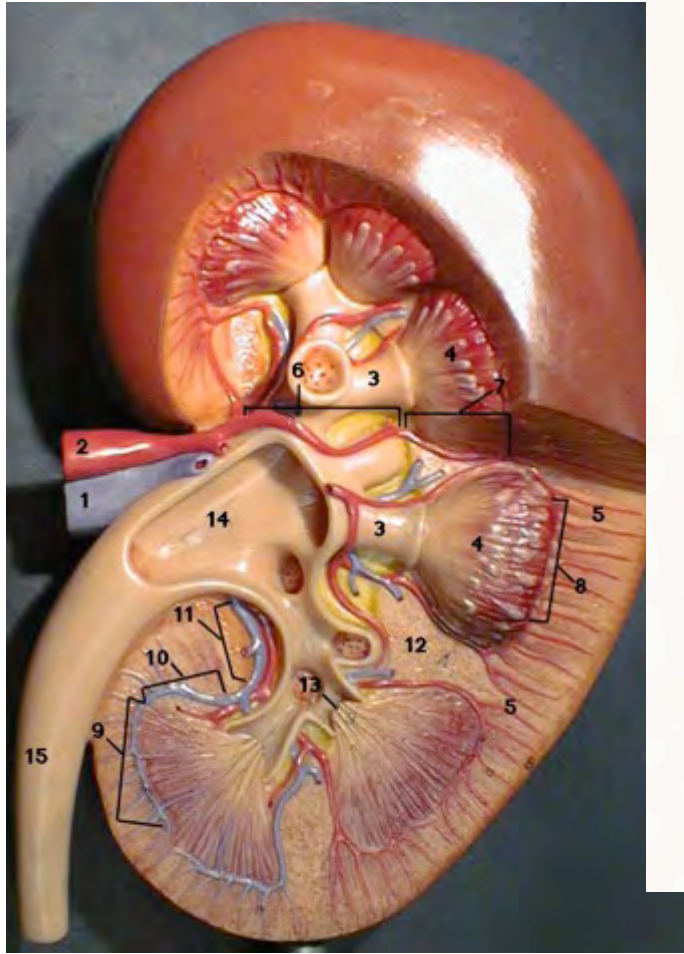
- Bowman's capsule ◆
- Proximal tubule ◆
- Loop of Henle ◆
- Distal tubule ◆
- Collecting duct ◆
- Peritubular capillaries and vasa recta ◆

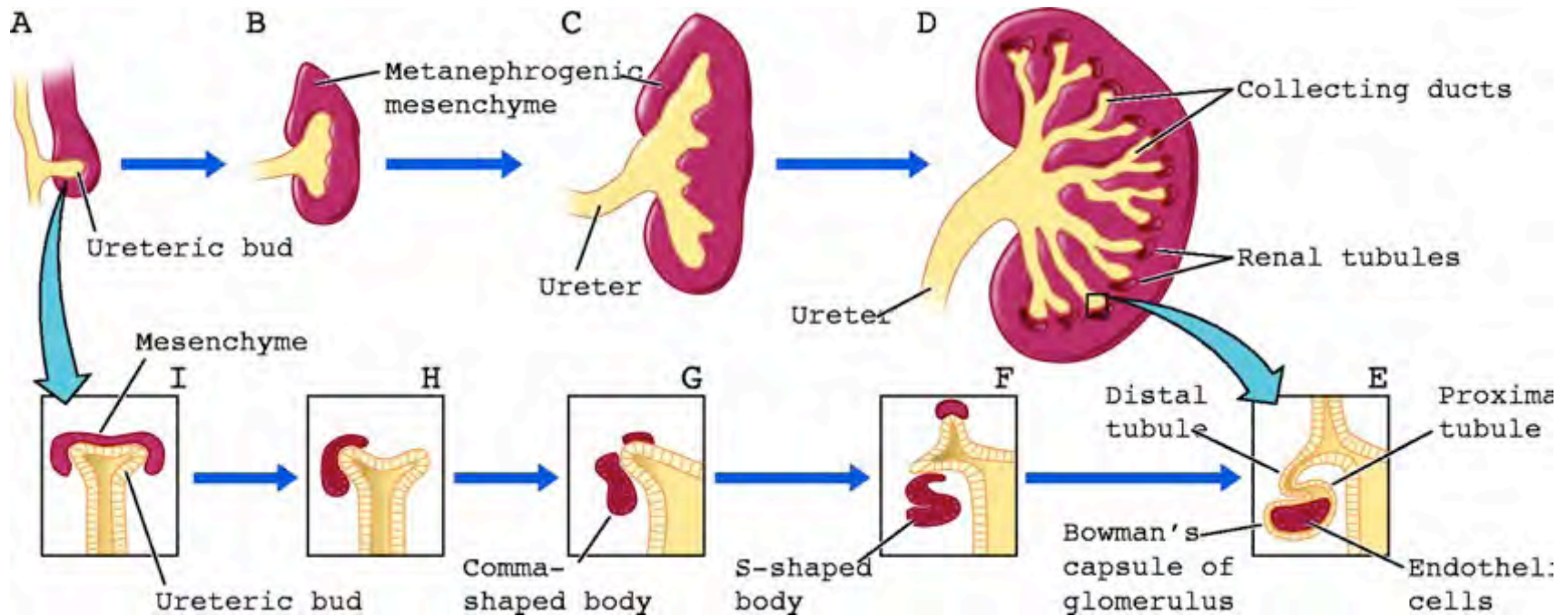


## Glomerulus: Filtration of molecules

- ~40 nM threshold/ 60 kD
  - Inulin 5 kD -----fully filtered  
(also not secreted/reabsorbed or metabolized in the tubule, hence it is used as a tracer to measure Glomerular Filtration Rate)
  - Alb 70 kD---- largely excluded
- Favors cations because it is negatively charged due to heparan sulfation of basement membrane;
  - thus negatively charged proteins such as albumin are preferentially excluded
  - albuminuria can be largely a result of charge alteration



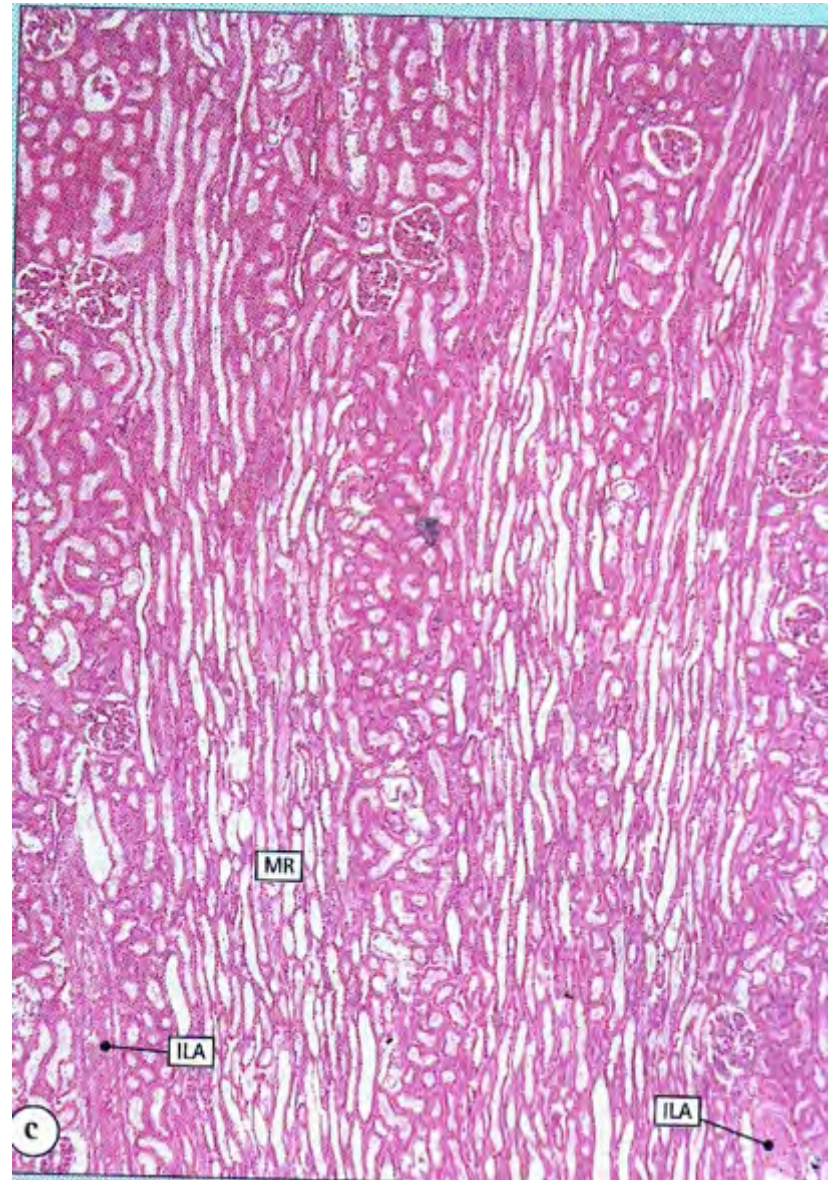
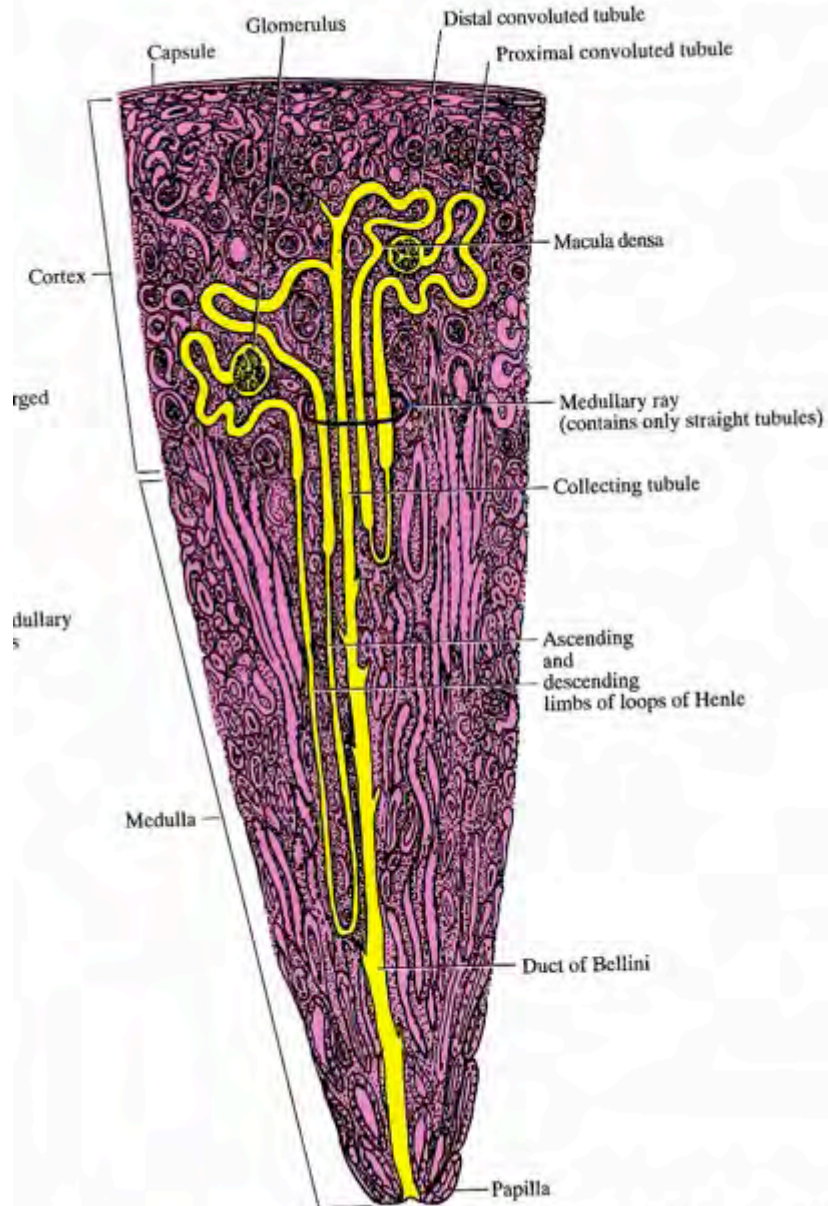




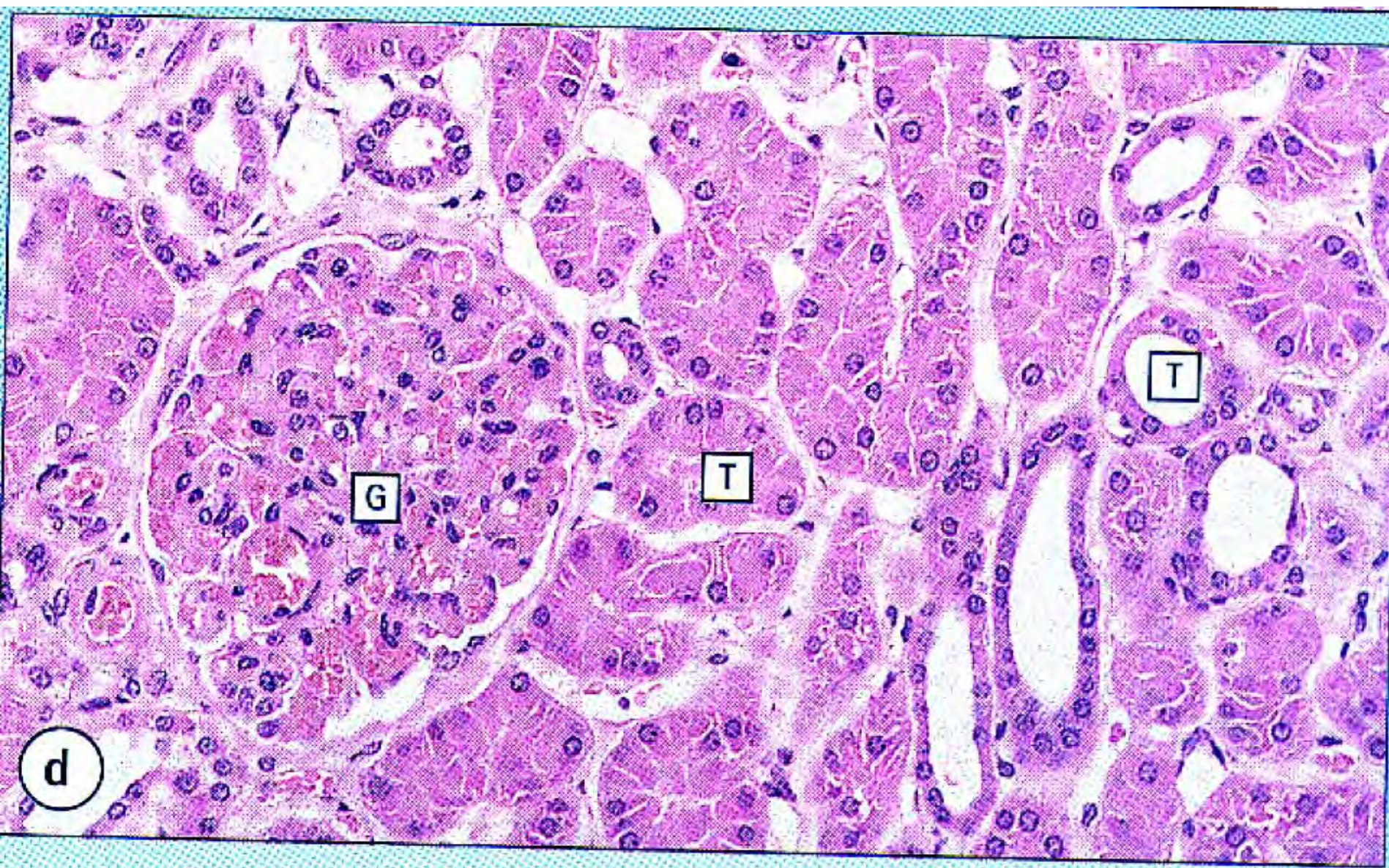
Continued growth and transformation of mesenchymal cell clusters results in epithelialization and tubulogenesis, ultimately forming the portion of the nephron comprising the glomerulus, proximal tubule, loop of Henle, and distal tubule.



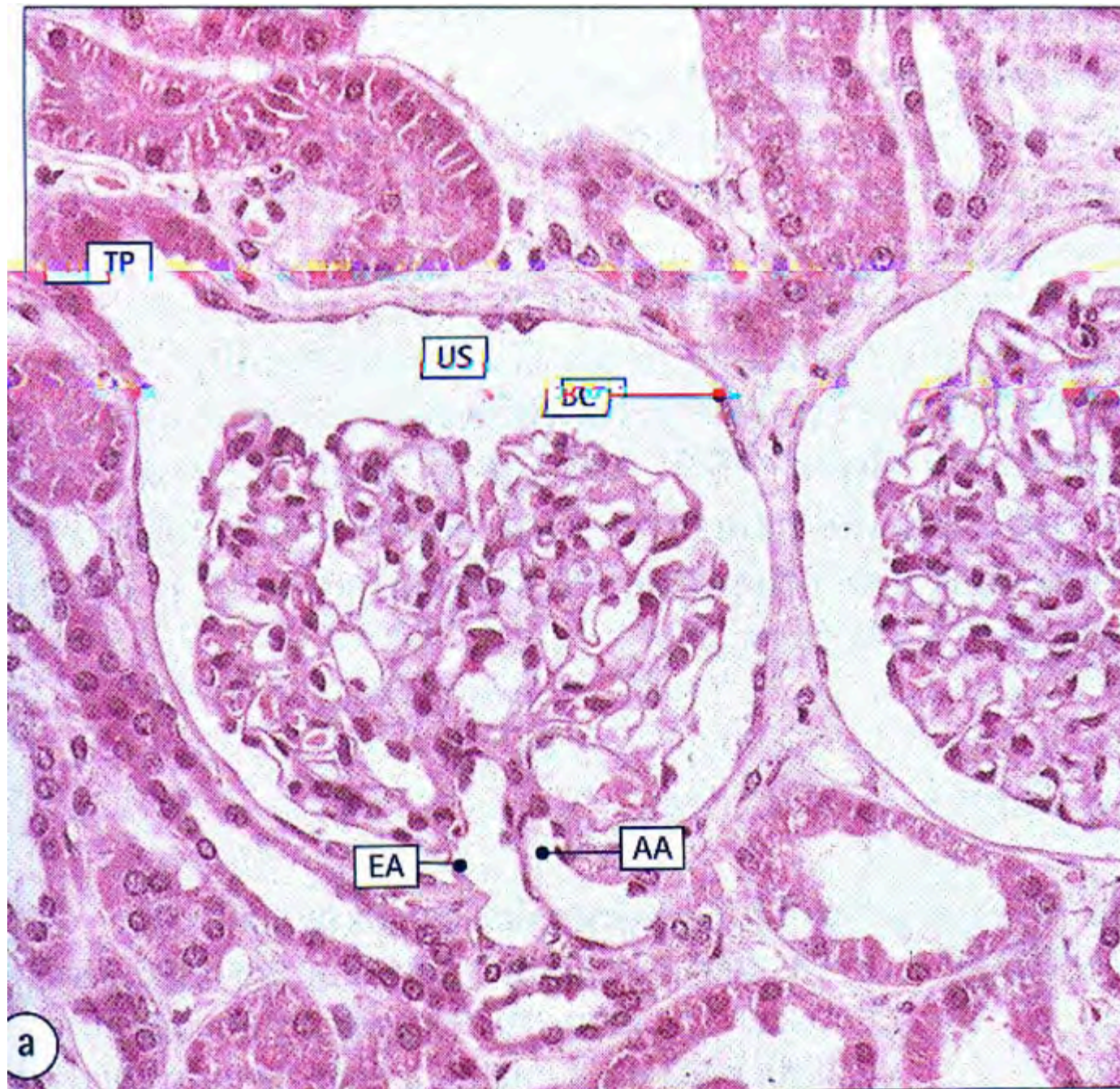
# Kidney Structure



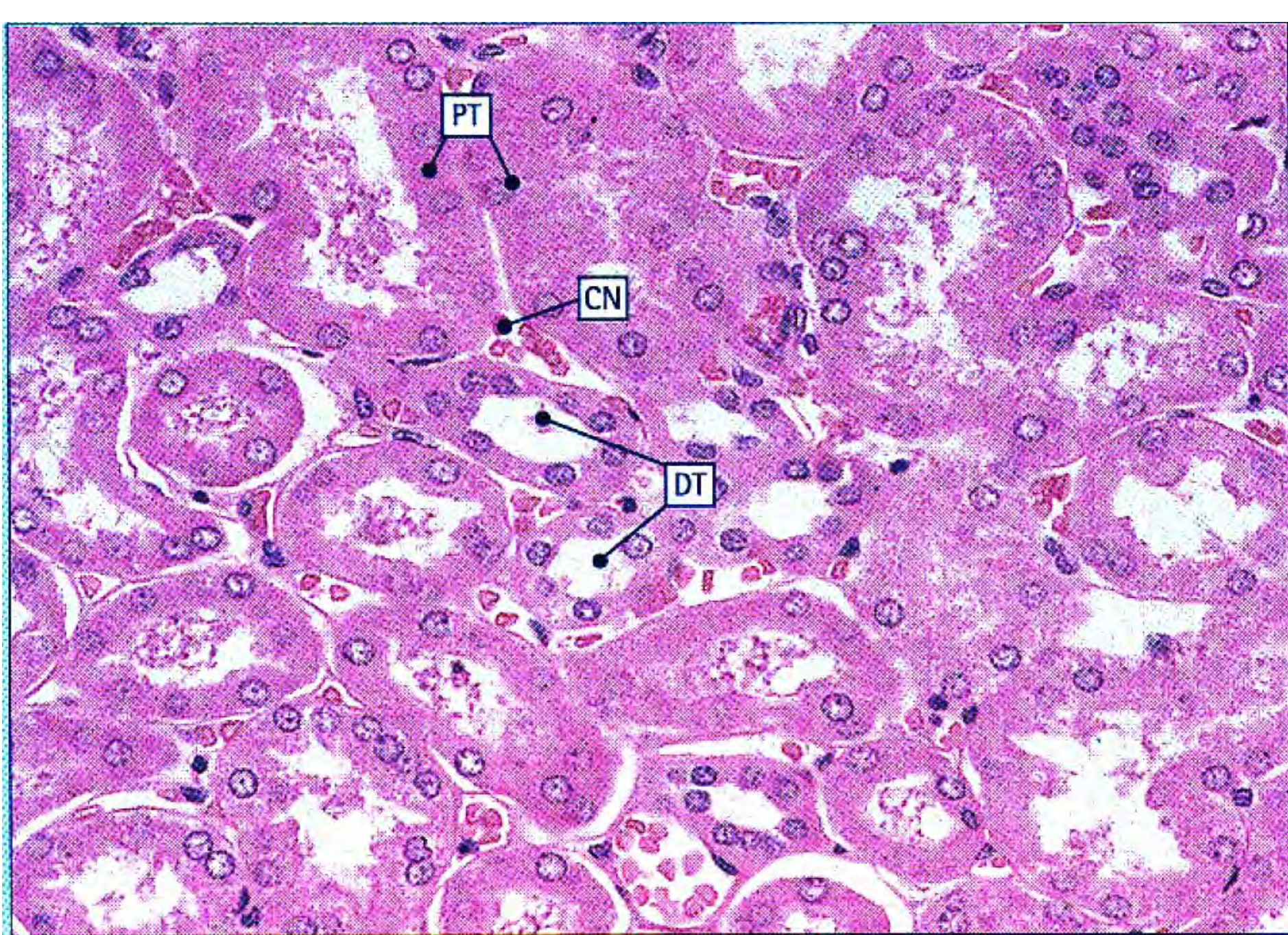




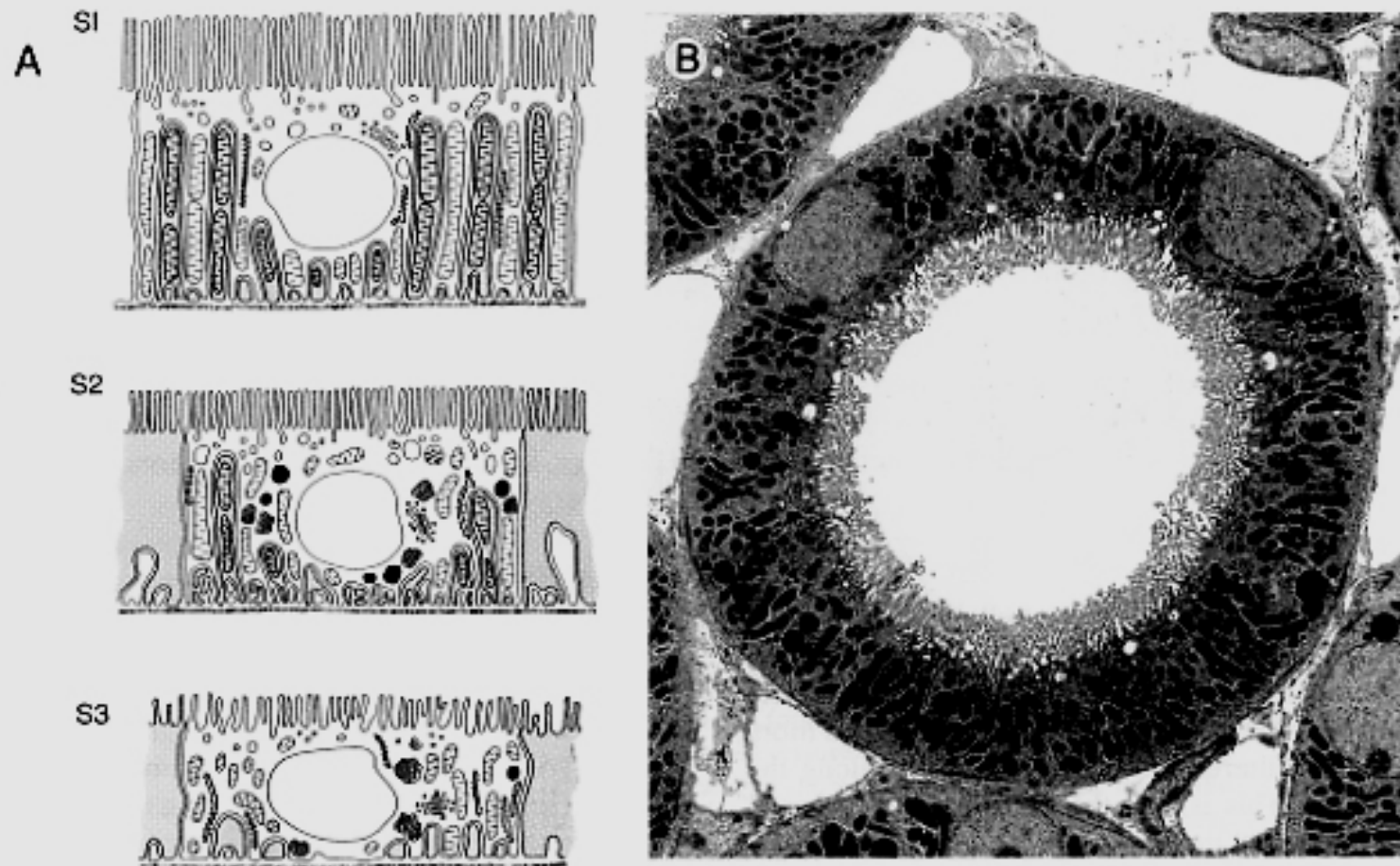








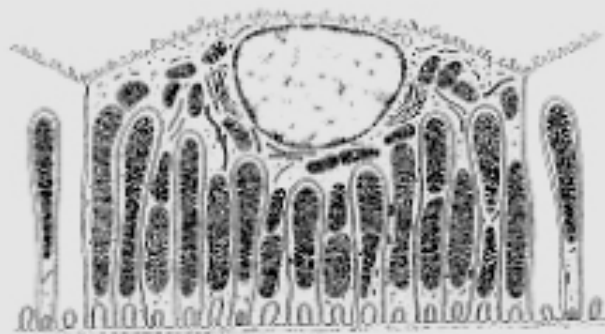




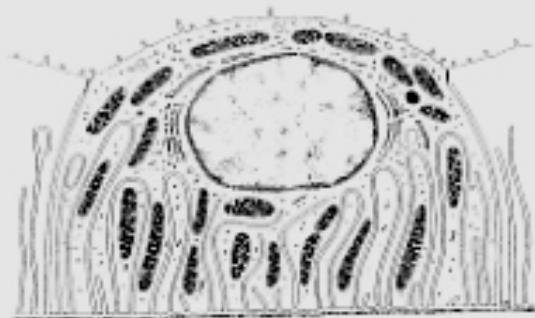
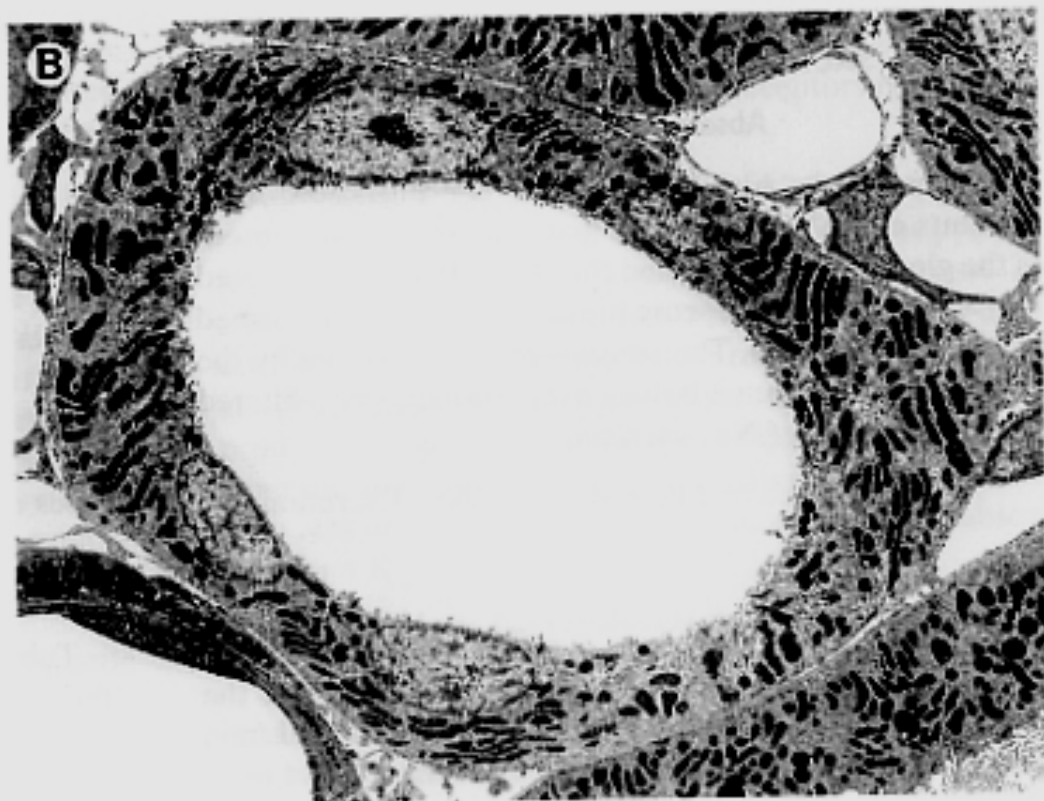
**FIGURE 7** Proximal tubule. The proximal tubule consists of three segments: S1, S2, and S3. The left panel shows schematic diagrams of the typical cells from these three segments, the right panel shows a cross section of the S1 segment. The S1 begins at the glomerulus, and extends several millimeters, before the transition to the S2 segment. The S3 segment, which is also called the proximal straight tubule, descends into the renal medulla to the inner medulla. The proximal tubule is characterized by a prominent brush border, which increases the membrane surface area by a factor of about 40-fold. The basolateral infoldings, which are lined with mitochondria, are interdigitated with the basolateral infoldings of adjacent cells (in these diagrams, processes that come from adjacent cells are shaded). These adaptations are most prominent in the first parts of the proximal tubule, and are less well developed later along the proximal tubule.

**A**

DCT

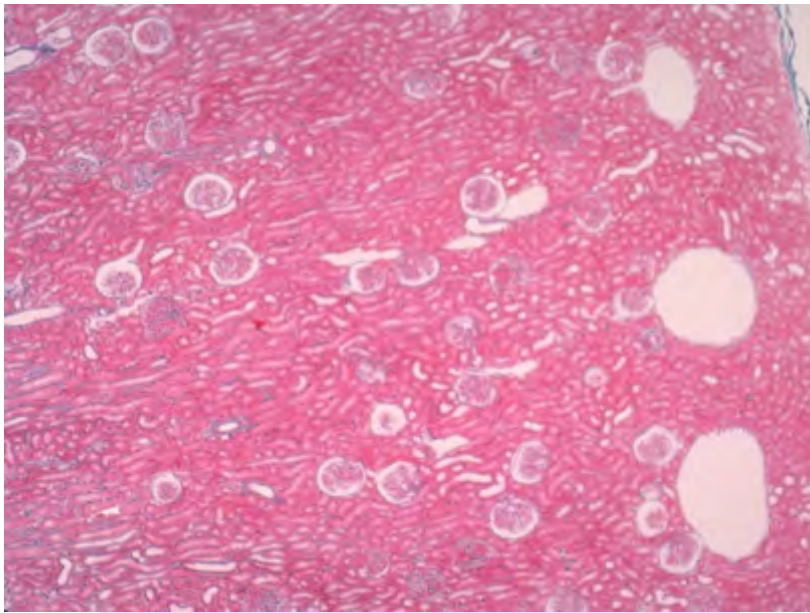


CT

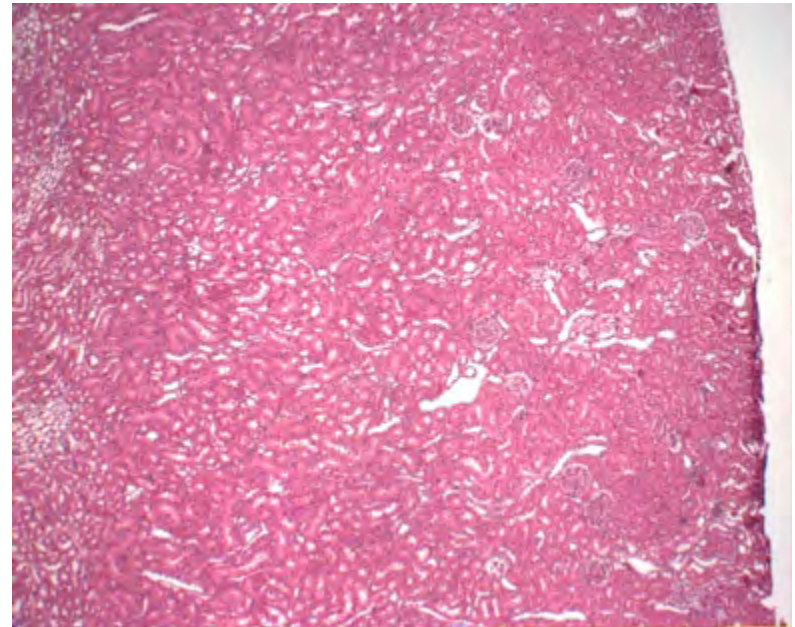
**B**

**FIGURE 9** Distal convoluted tubule. The distal convoluted tubule is customarily divided into two parts: the true distal convoluted tubule (DCT, shown schematically on the left, and in cross-section on the right) and the connecting tubule (CT), where cell morphology is somewhat more similar to collecting duct.

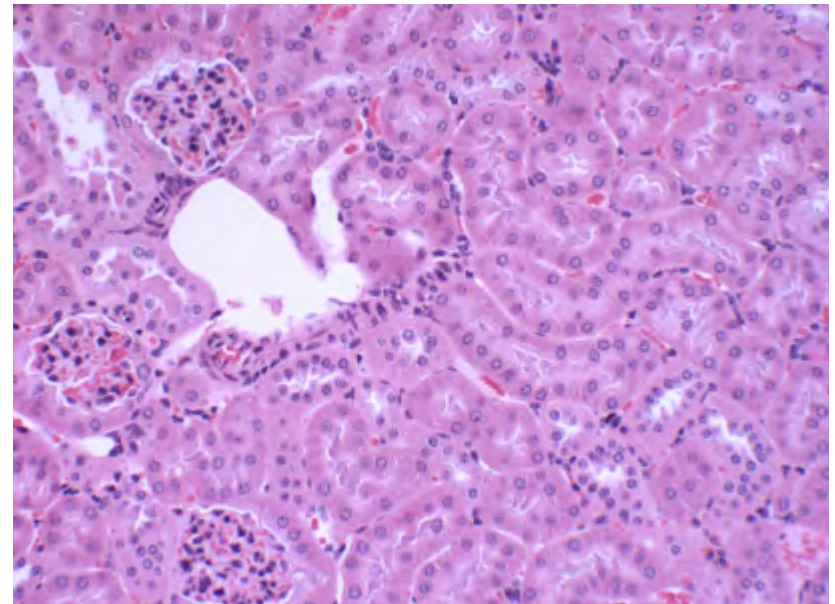




**H&E of human Kidney  
x40 magnification**

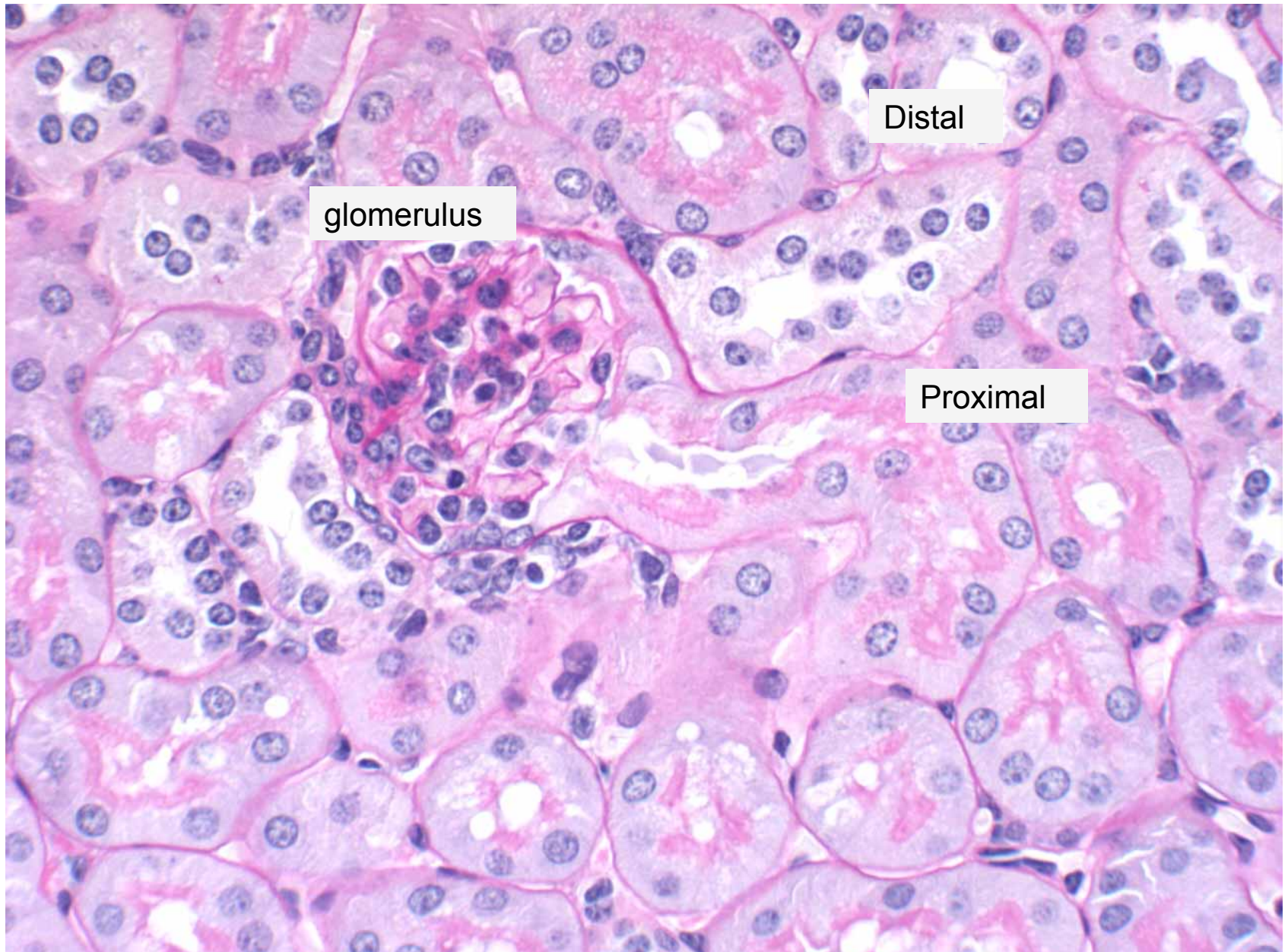


**H&E of mouse Kidney x40 magnification**



**H&E of mouse Kidney x200 magnification**





PAS stain (Periodic Acid Schiff) highlights basement membranes (carbohydrate structures)

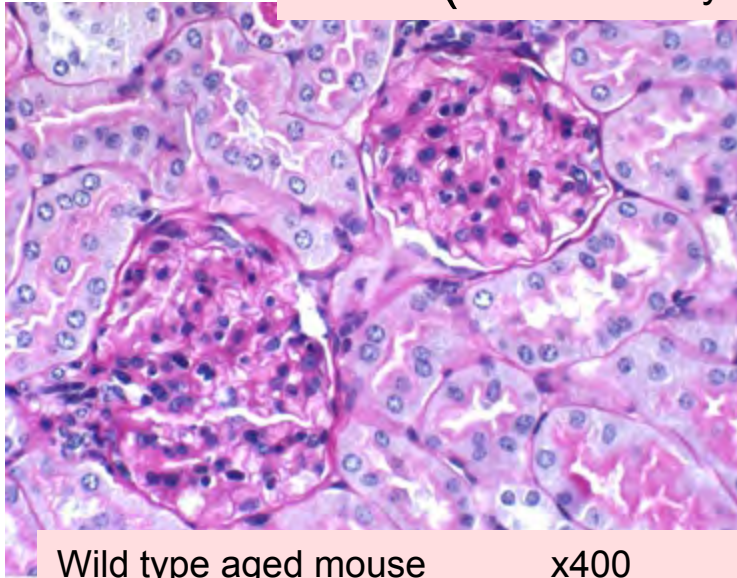


**The PAS stain  
identifies the  
mucus  
producing  
goblet cells in  
human colon**



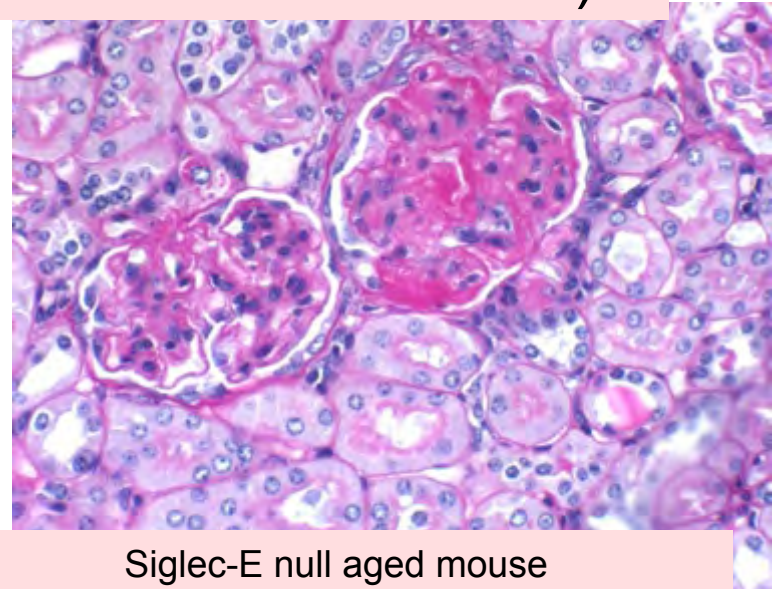


## PAS (mouse kidneys sectioned at 3 micron thickness)

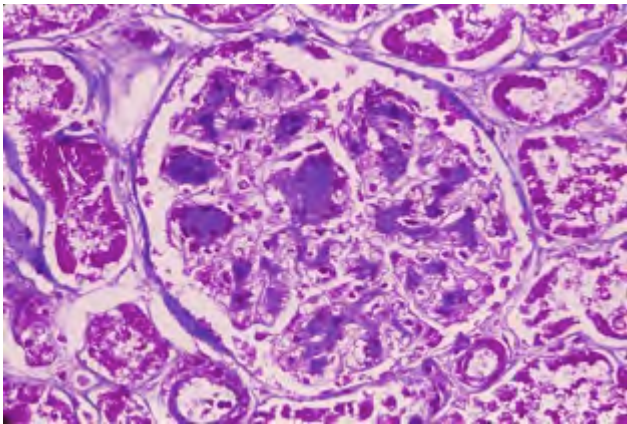


Wild type aged mouse

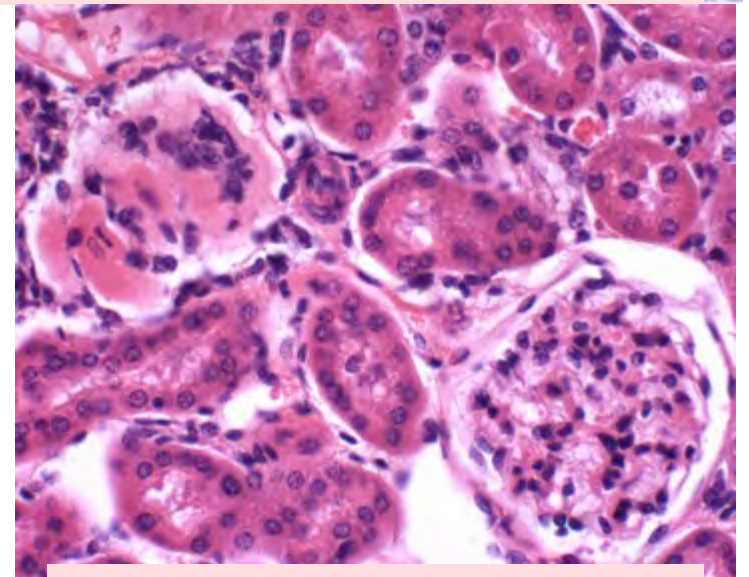
x400



Siglec-E null aged mouse



Human Kidney with Expanded nodular formations in glomerular capillaries -- positive staining with PAS-- in diabetic glomerulosclerosis

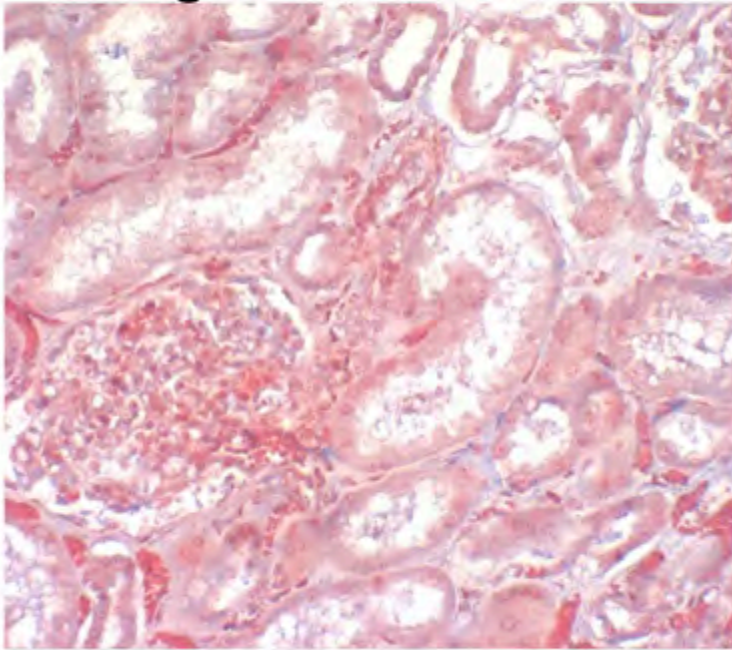


H&E of same mouse kidney x400

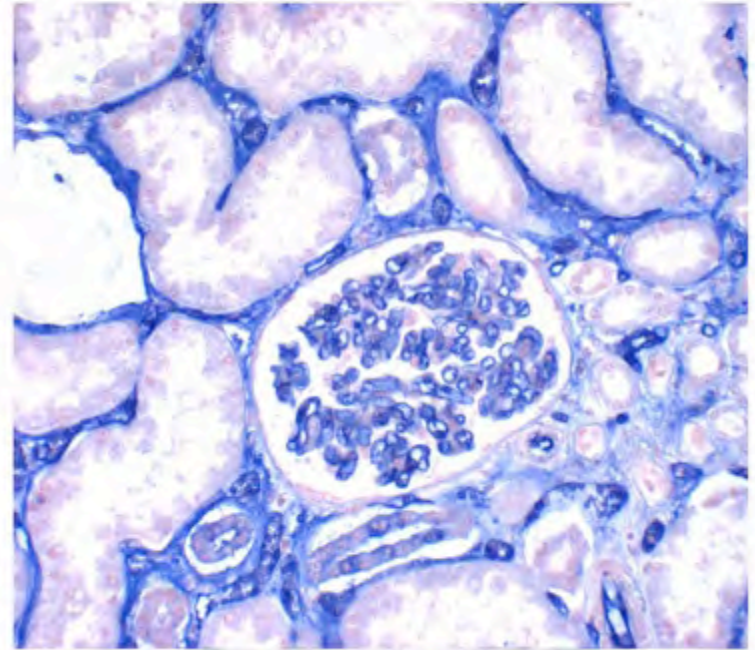


## Human Kidney x400

Negative control



UEA lectin for blood vessels



Capillaries of the glomerulus highlighted with the UEA lectin

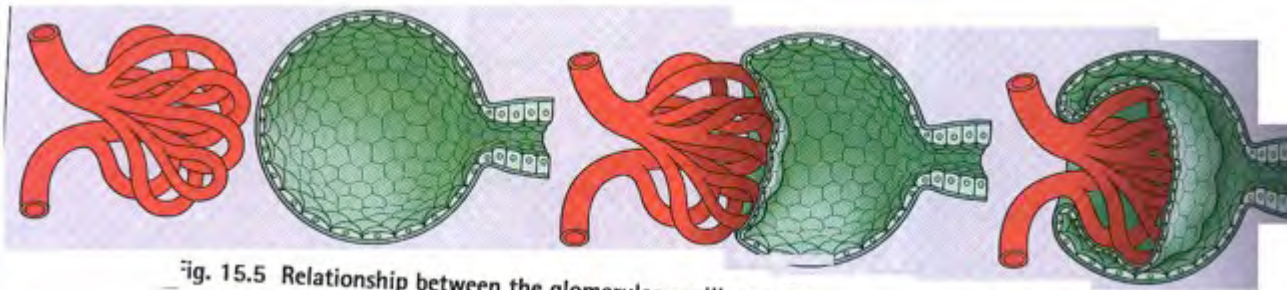
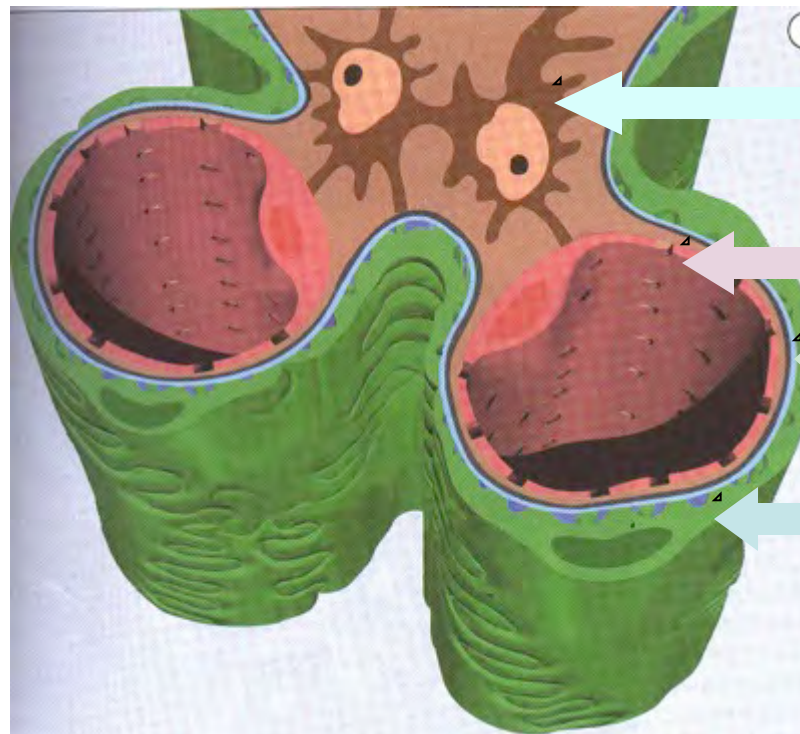


Fig. 15.5 Relationship between the glomerular capillary tuft and Bowman's capsule.



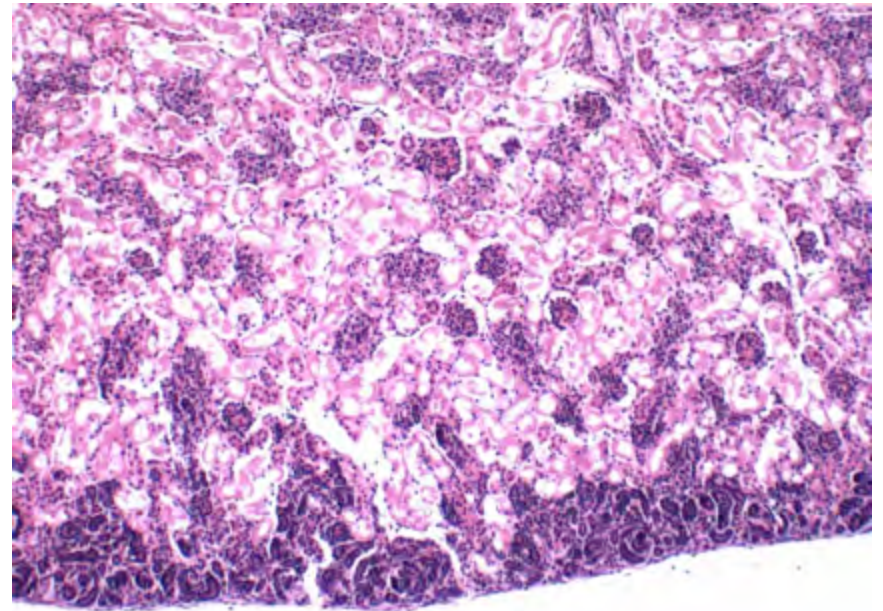
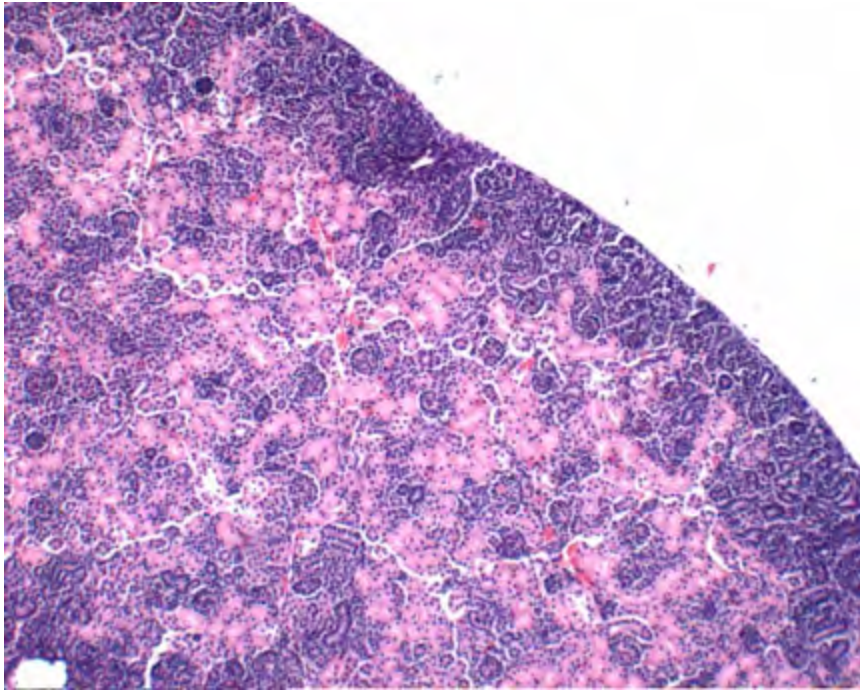
**Mesangial cells**

**Endothelial cells**

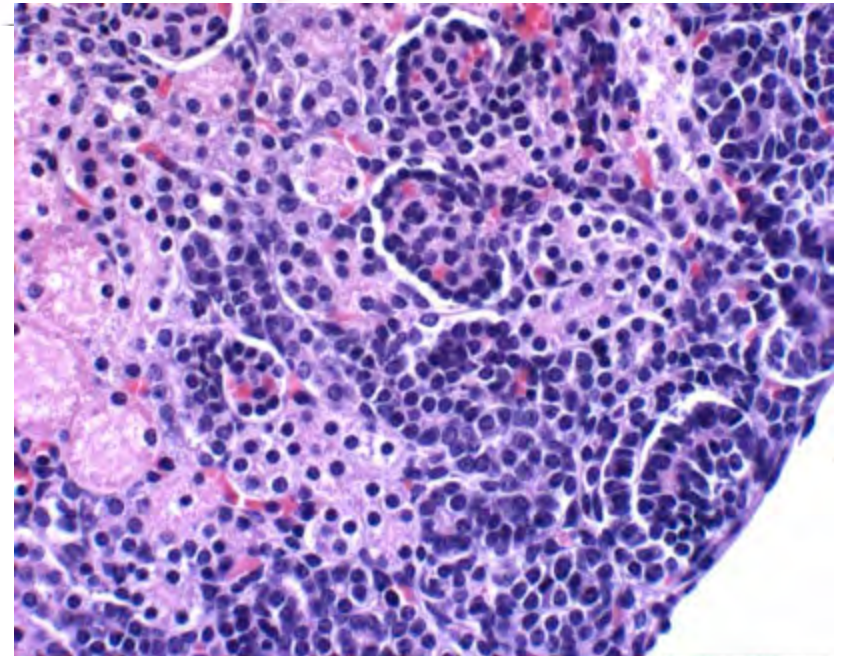
**Basement  
membrane**

**Epithelial cells**

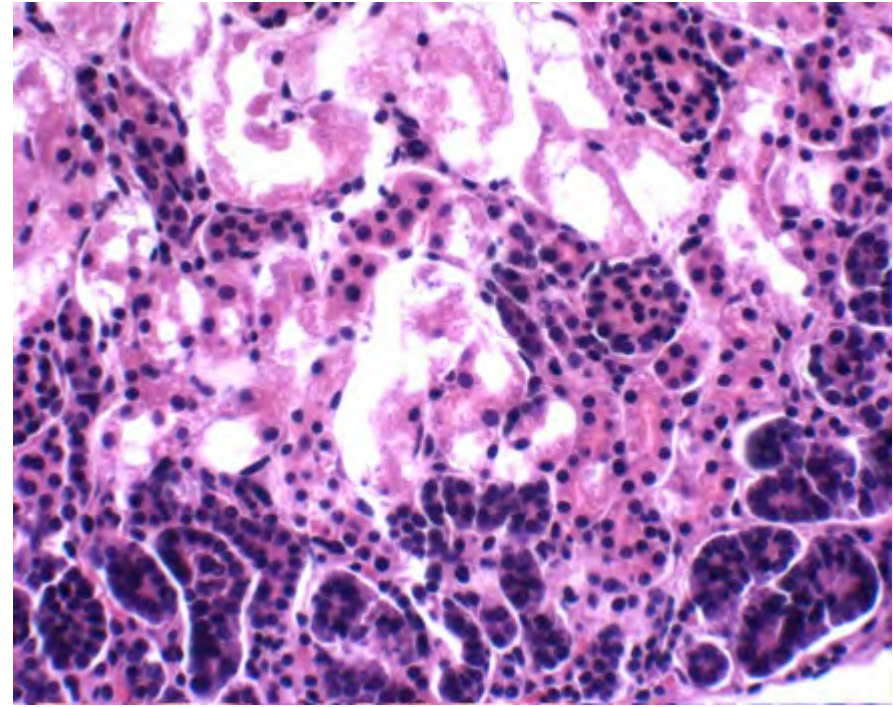
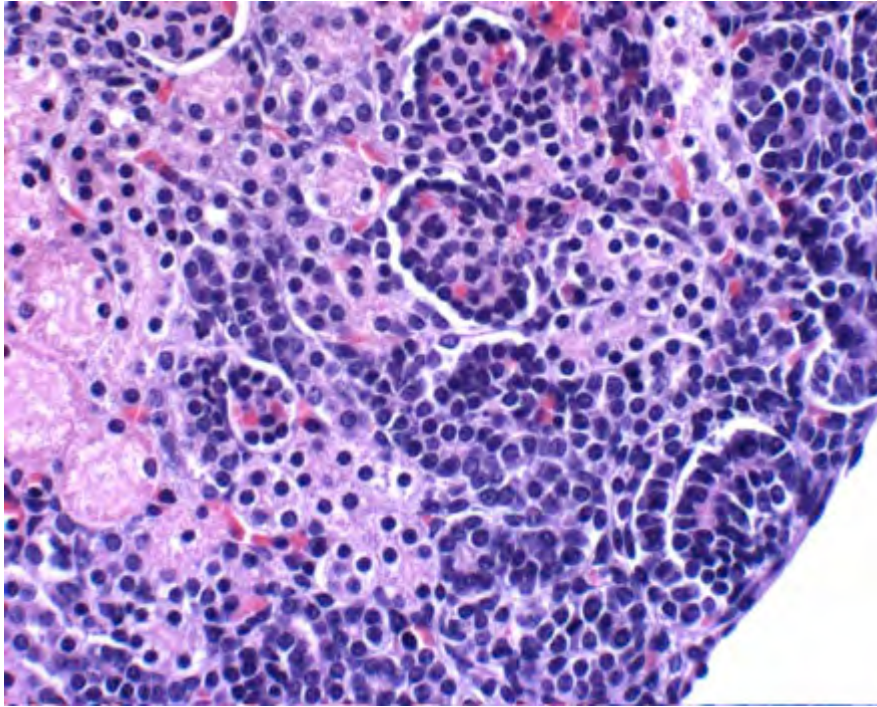




**H&E of Embryo d16.5  
Mouse Kidney cortex with  
developing glomeruli**



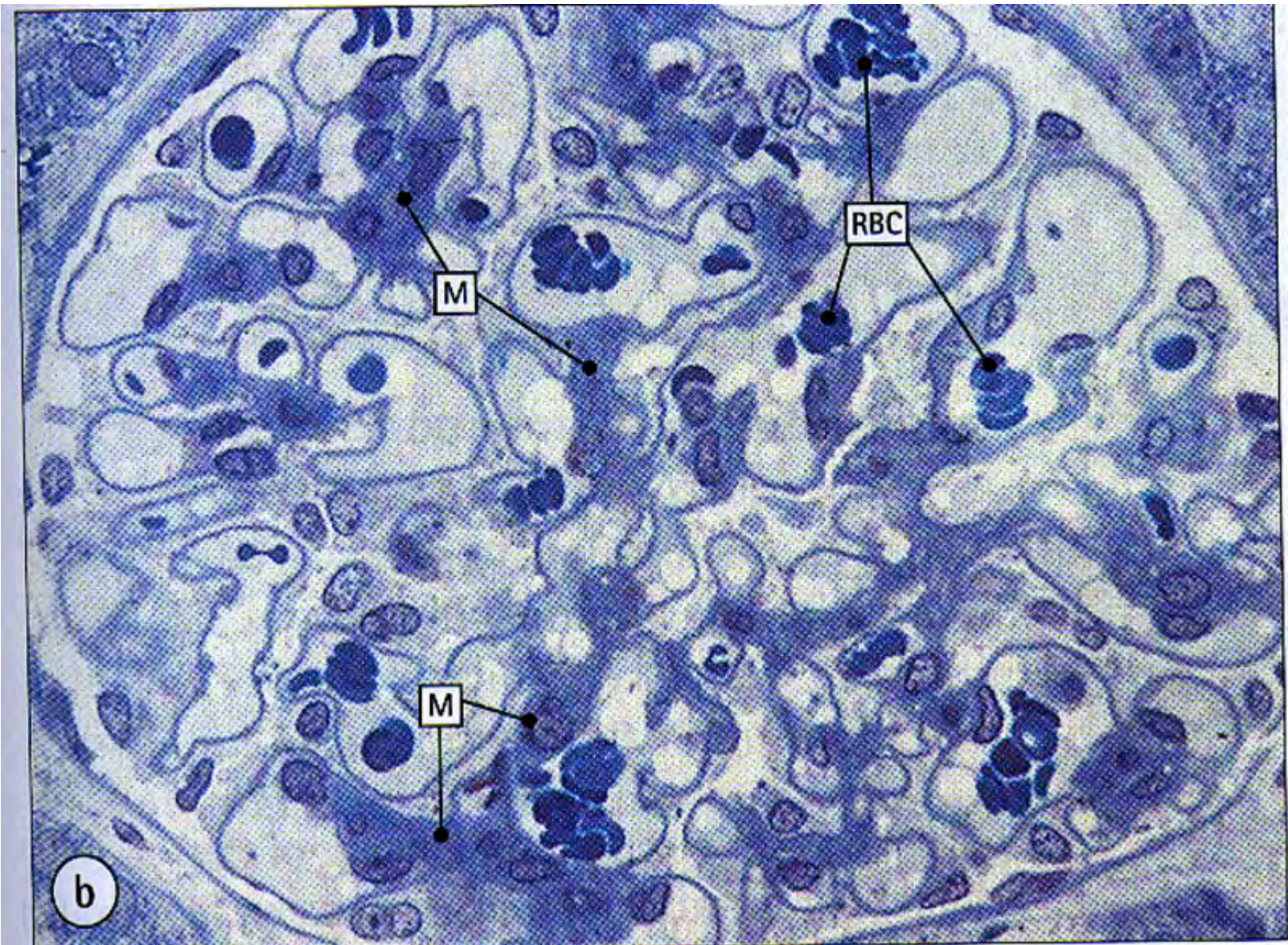




**H&E of Embryo d 16.5 Mouse Kidney cortex**

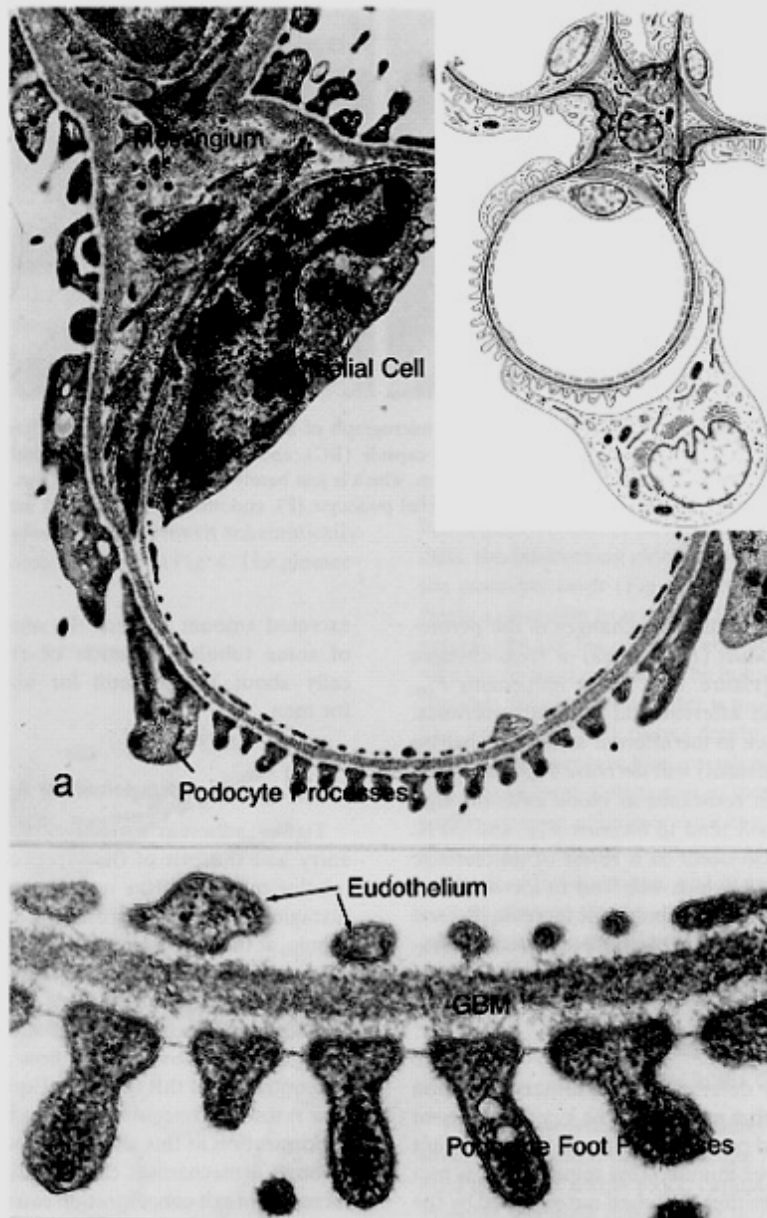
**x400 magnification**



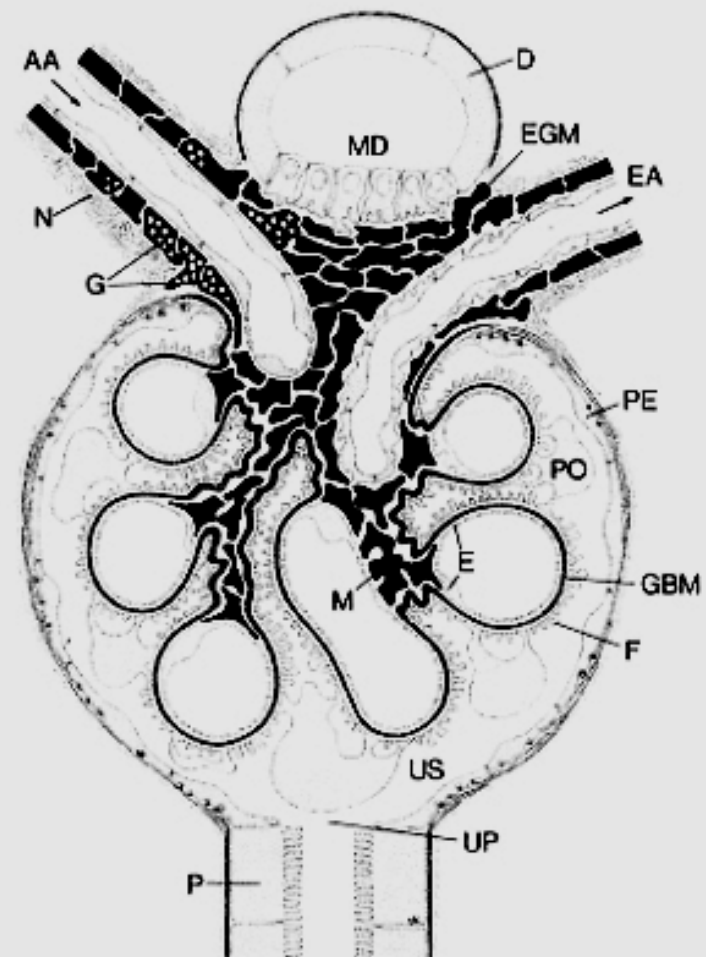


Thin section of glomerulus



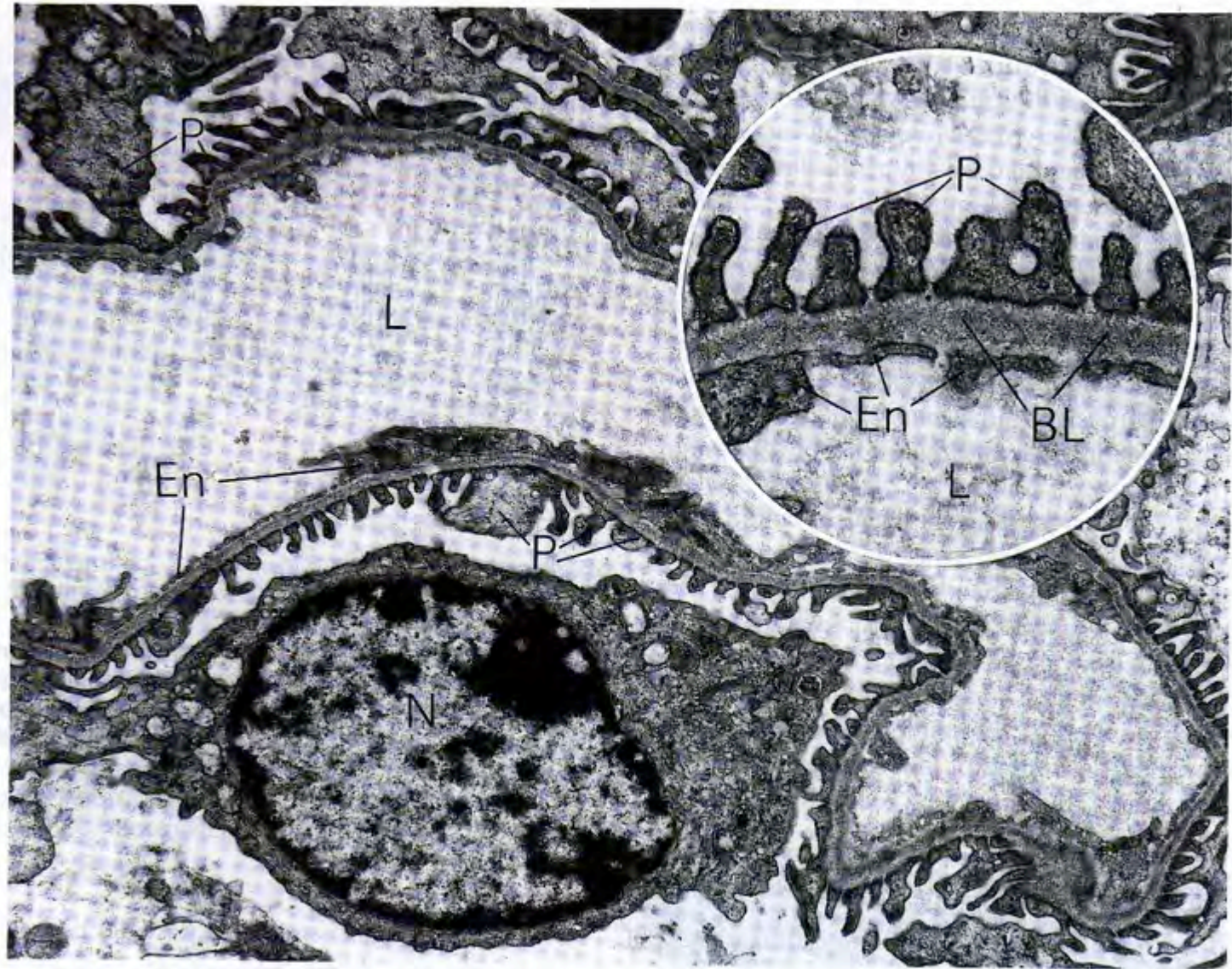


**FIGURE 5** Structure of the glomerular capillary loop and the filtration barrier. (a) A single capillary loop showing the endothelial and foot process layers and the attachments of the basement membrane to the mesangium. Pressure in the glomerular capillary bed is substantially higher than in other capillaries. As shown in the diagrammatic insert, the mesangium provides the structural supports which permit these cells to withstand these high pressures. (b) The glomerular filtration barrier.



**FIGURE 3** Anatomy of the glomerulus and juxtaglomerular apparatus. Schematic diagram of a section of a glomerulus and its juxtaglomerular apparatus. Structures shown are as follows: afferent arteriole (AA), efferent arteriole (EA), macula densa (MD), distal tubule (D), juxtaglomerular granular cell (G), sympathetic nerve endings (N), mesangial cell (M), extraglomerular mesangial cell (EGM), endothelial cell (E), epithelial podocyte (PO), with foot process (F), parietal epithelial cell (PE), glomerular basement membrane (GBM), urinary space (US), urinary pole (UP), and proximal tubule (P).



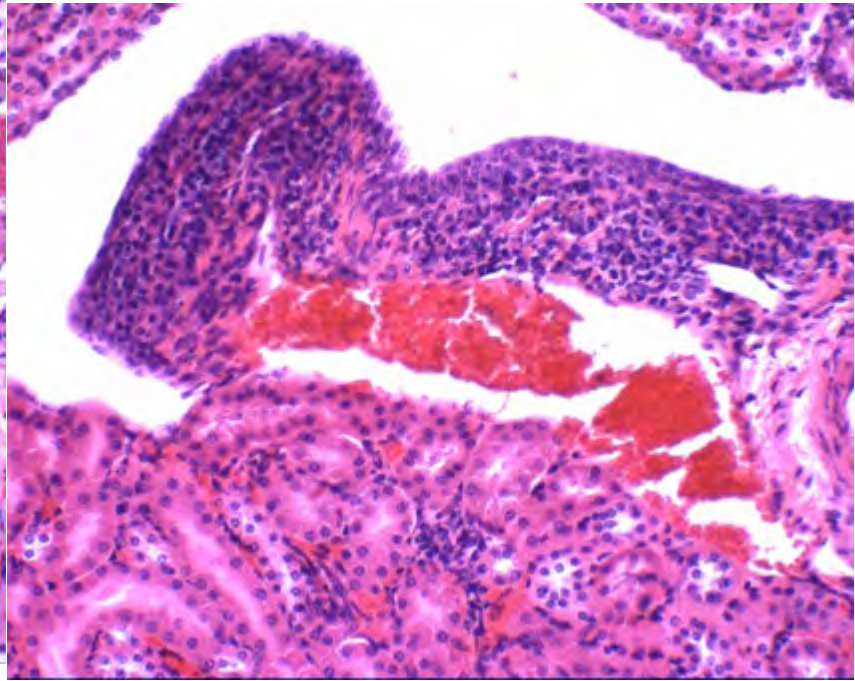
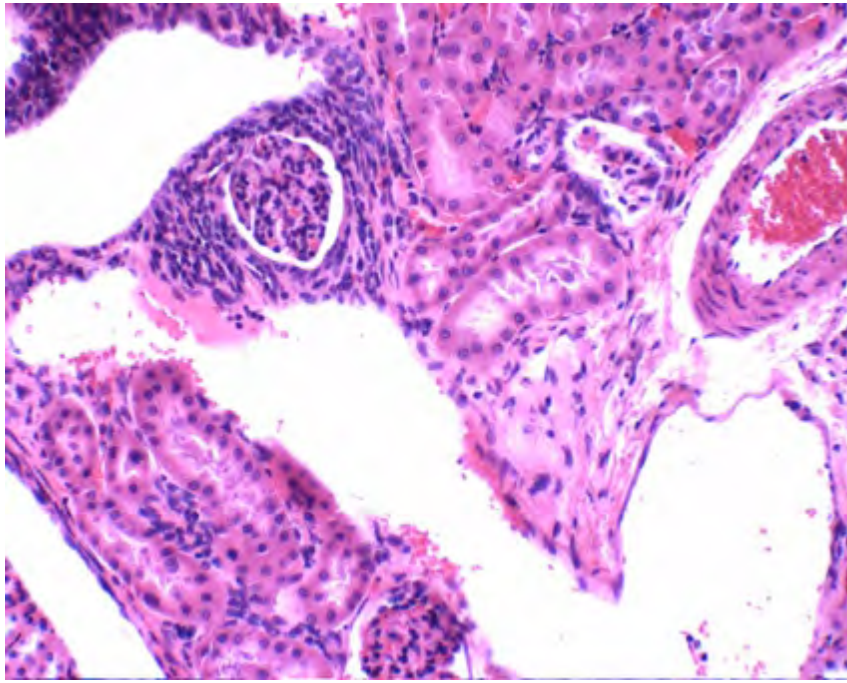


Electron micrograph of a kidney element.

Figure 10-10



	A	B	C	D
1	C57BL/6 blood chemistry normal ranges		n ~= 400	
2	age ~ 2-5 months			
3		mean	sd	
4				
5	Glucose (mg/dL)	196.7	91.2	
6	Urea nitrogen (mg/dL)	21.4	4.4	
7	Creatinine (mg/dL)	0.2	0.2	
8	Bicarbonate (mEq/L)	15.9	3.2	
9	Chloride (mEq/L)	107.5	4.2	
10	Sodium (mEq/L)	150.5	3.9	
11	Potassium (mEq/L)	5.0	1.1	
12	Calcium (mg/dL)	9.0	0.4	
13	Direct bilirubin (mg/dL)	0.1	0.1	
14	Total bilirubin (mg/dL)	0.4	0.2	
15	Albumin (g/dL)	1.4	0.2	
16	Total protein (g/dL)	4.2	0.3	
17	Phosphorus (mg/dL)	8.0	1.6	
18	AST (SGOT) (IU/L)	72.7	36.3	
19	ALT (SGPT) (IU/L)	38.7	25.9	
20	Alkaline phos, total (IU/L)	101.5	32.2	
21				
22	LIPID PANEL (n~=50)			
23	Lipase (U/L)	53.0	12.9	
24	Cholesterol (mg/dL)	98.6	17.9	
25	HDL-chol. (mg/dL)	78.6	21.9	
26	LDL-chol. (mg/dL)	12.2	8.6	
27	Triglycerides (mg/dL)	67.8	26.1	
28				



## **Pathology of kidney:**

### ***Inflammation:***

---glomerular = glomerulonephritis,-autoimmune

---tubular=pyelonephritis

### ***hyperplasia, accumulations:***

***fibrosis/scarring, cysts***

### ***Infarct***

***Cancer:*** primary adenocarcinomas or metastatic



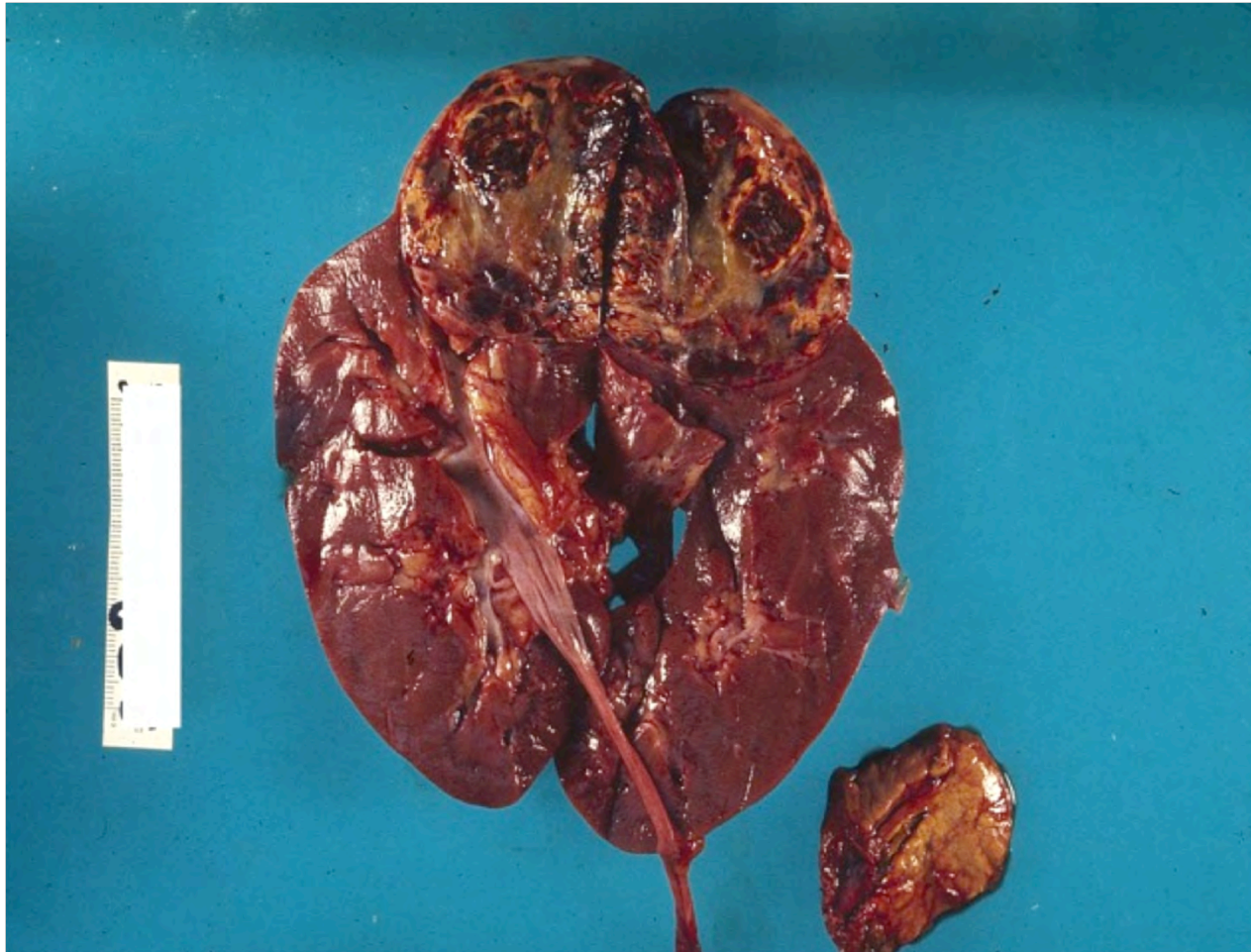


As mice age, the kidneys develop similar cysts due to chronic pyelonephritis (Tubules inflammation, scarring and repair) with consequent stasis

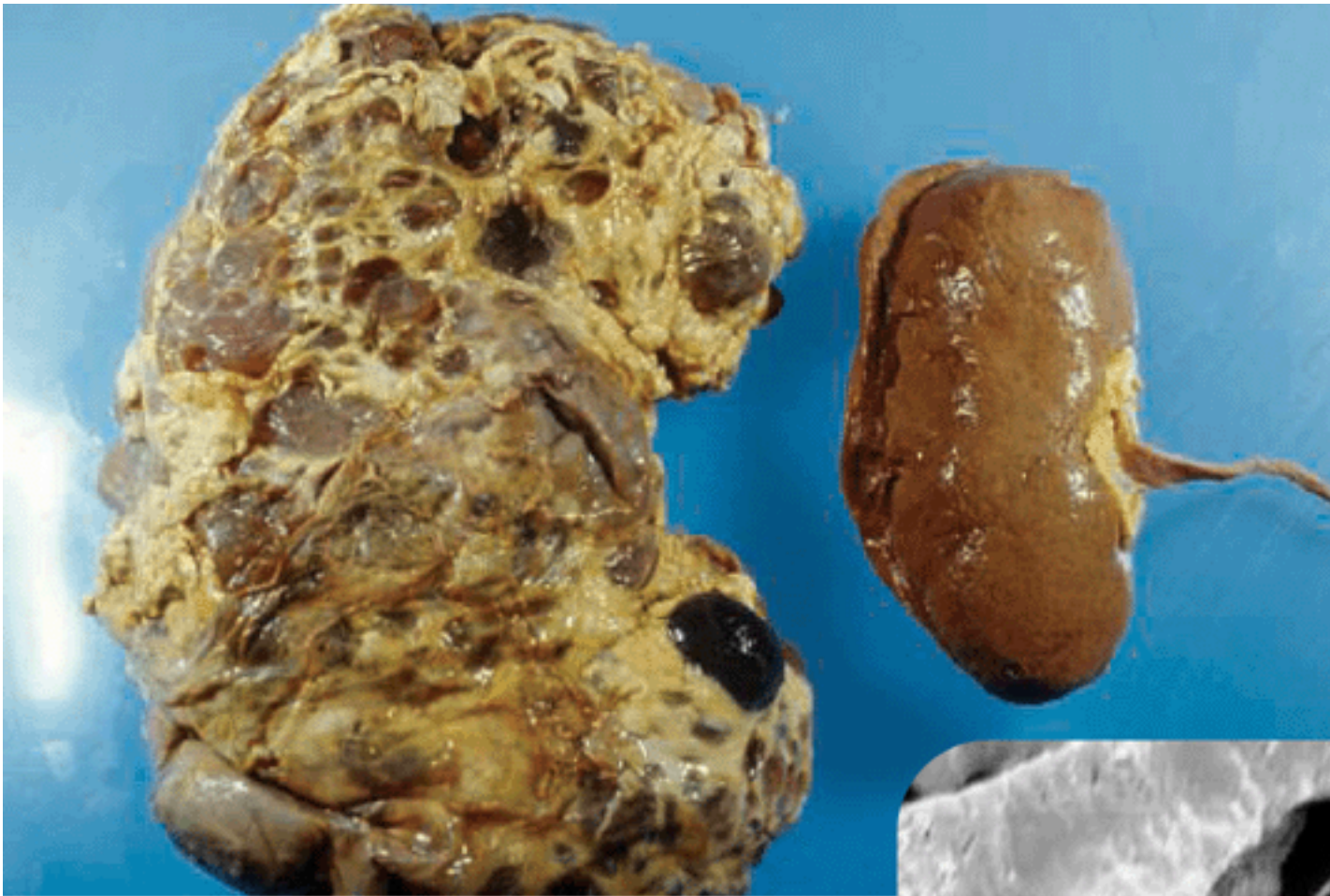
## Simple cysts in human kidney

<http://www-medlib.med.utah.edu>

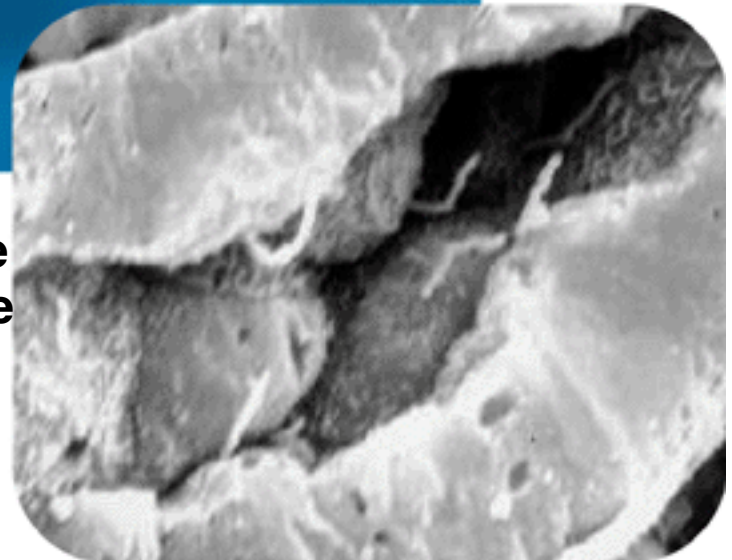




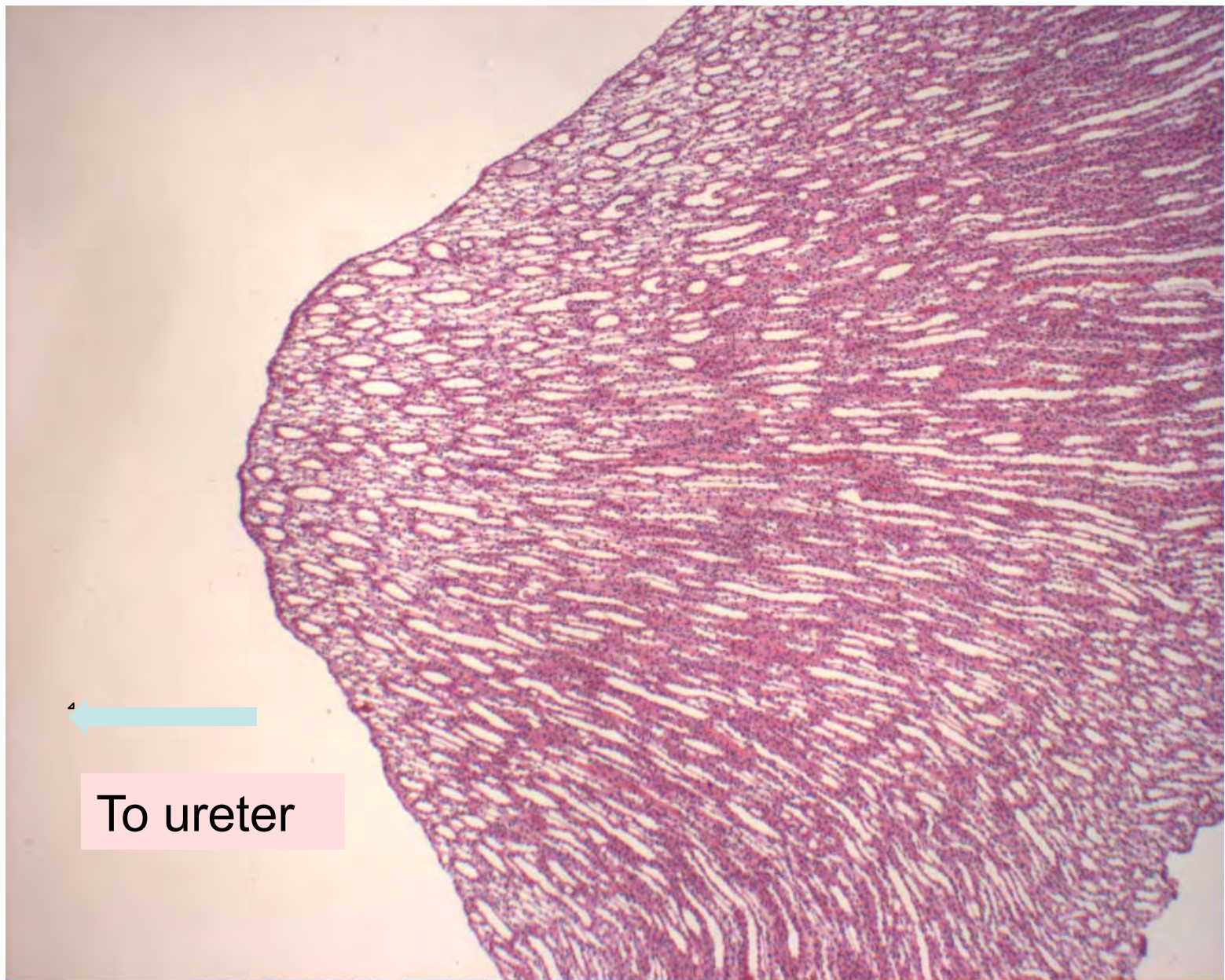
Resected renal cell carcinoma, at upper pole of the kidney



Compared to a normal kidney (*right*), those from people with polycystic kidney disease (*left*) are enlarged and ravaged with cysts. Defects in the primary cilia on kidney tubule epithelial cells (*inset*) lead to the devastation. Science Oct 14, 2005







To ureter

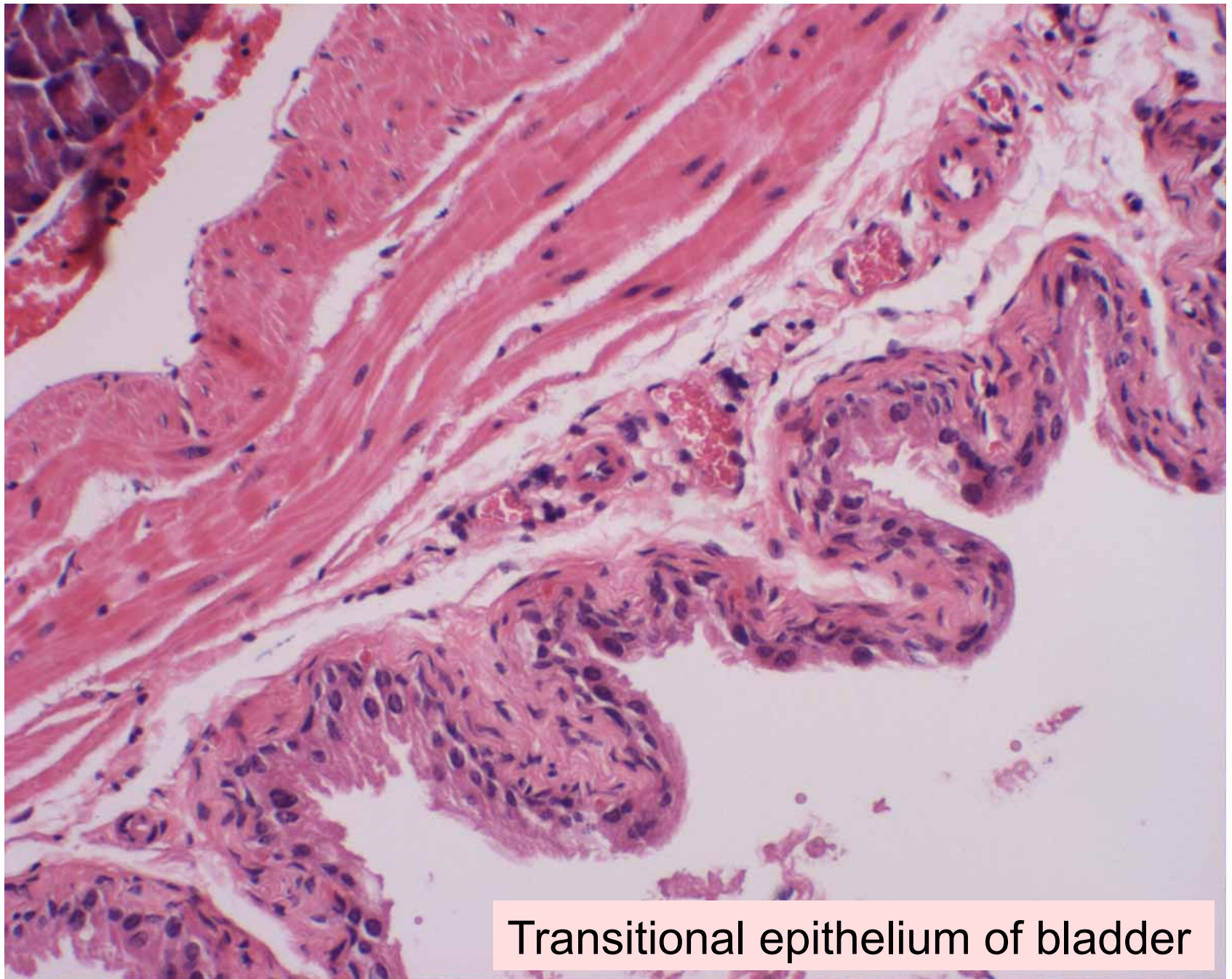
**H&E of mouse Kidney Pelvis x40 magnification**





Transitional epithelium of bladder and ureter





Transitional epithelium of bladder



# Histologic highlights within the Female and Male Reproductive systems

# Uterus, Fallopian Tubes, ovaries

Human

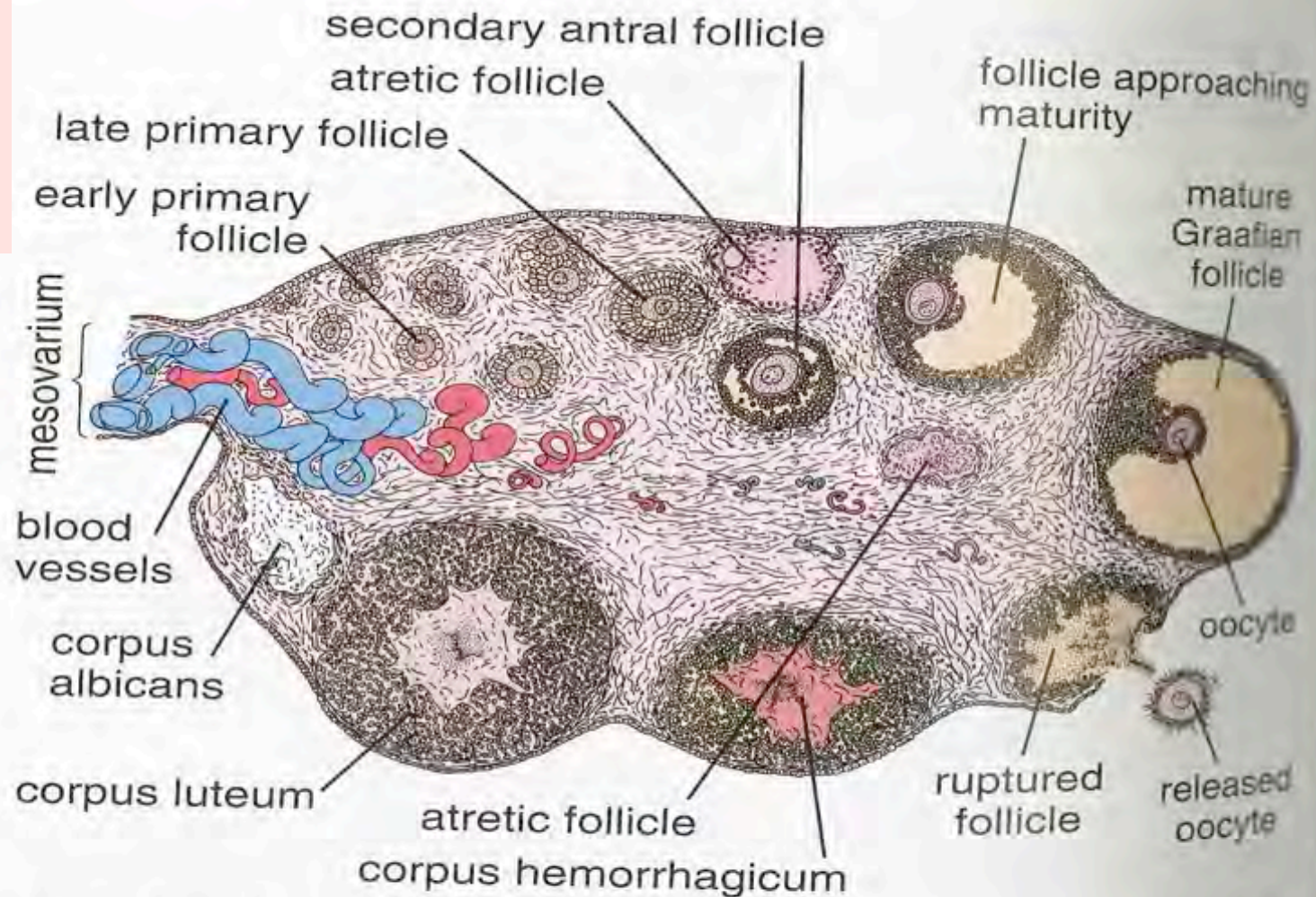


Mouse





## Stages of ovarian follicular development



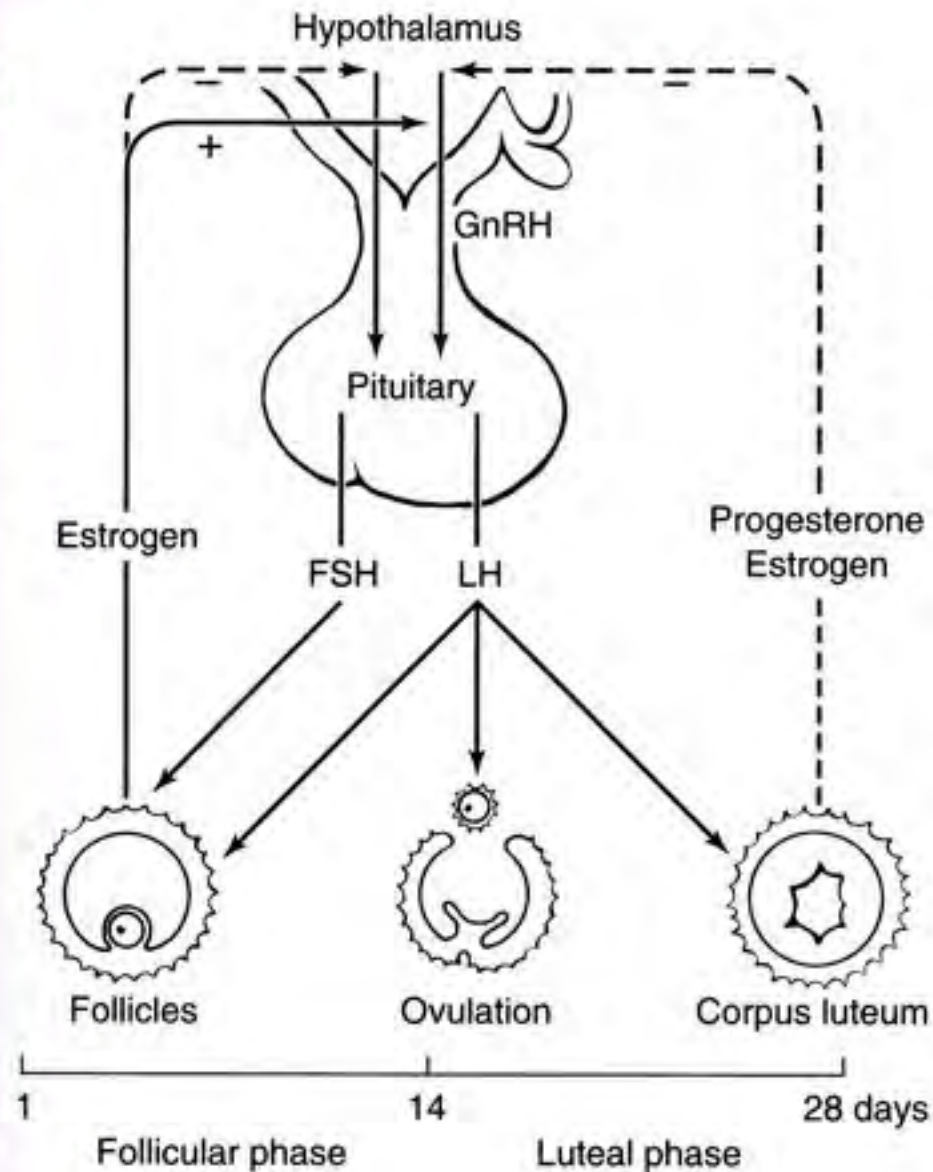
**FIGURE 23.2 Schematic drawing of a section through the ovary.** This section shows, in clockwise order, stages of follicular development from the early primary follicle to the mature (Graafian) follicle. Changes in the follicle after ovulation lead to development of the corpus luteum and eventually the corpus albicans. Note the highly coiled blood vessels in the hilum and medulla of the ovary.



# Hormonal control of the Ovary

At the time of menstruation, **Follicular Stimulating Hormone (FSH)** from the pituitary, initiates follicular growth.

At the time of the maturation of the follicle, the **estrogen** rise leads ( via the hypothalamic interface) to the “positive feed-back” effect, and a release of **Luteinizing Hormone (LH)** occurs over a 24-48 hour period. This 'LH surge' triggers ovulation.



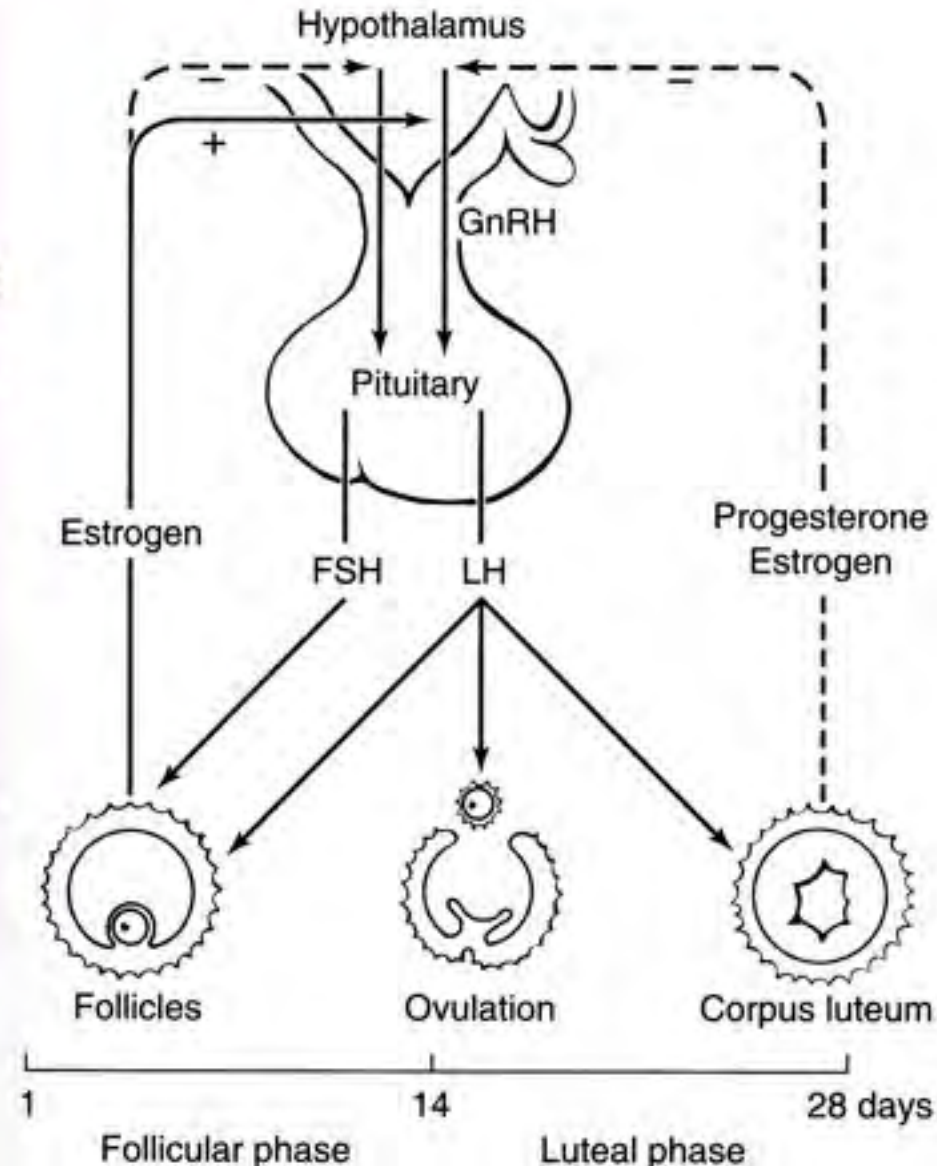


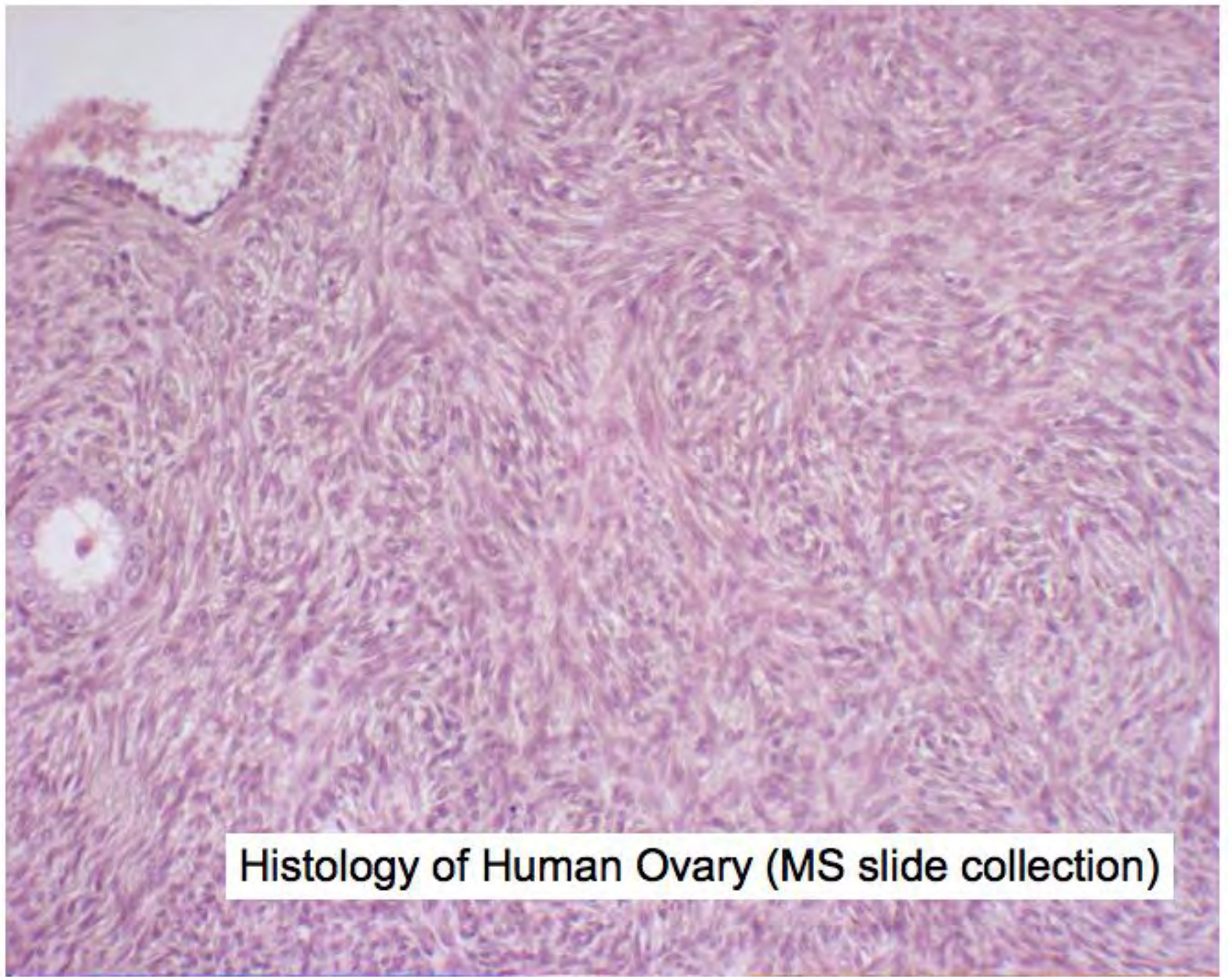
# Hormonal control of the Ovary

The LH surge not only releases the egg, but also initiates the conversion of the residual follicle into a corpus luteum.

The corpus luteum in turn, produces progesterone to prepare the endometrium for a possible implantation .

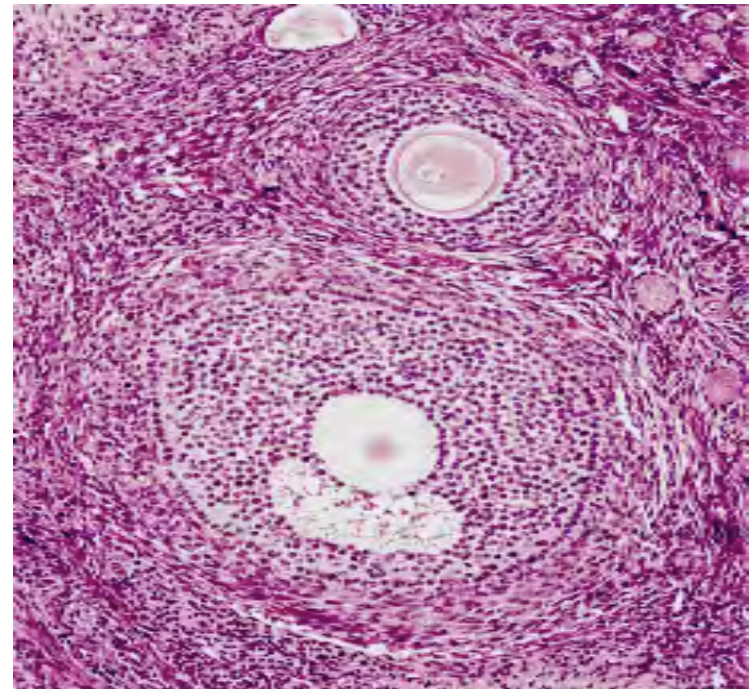
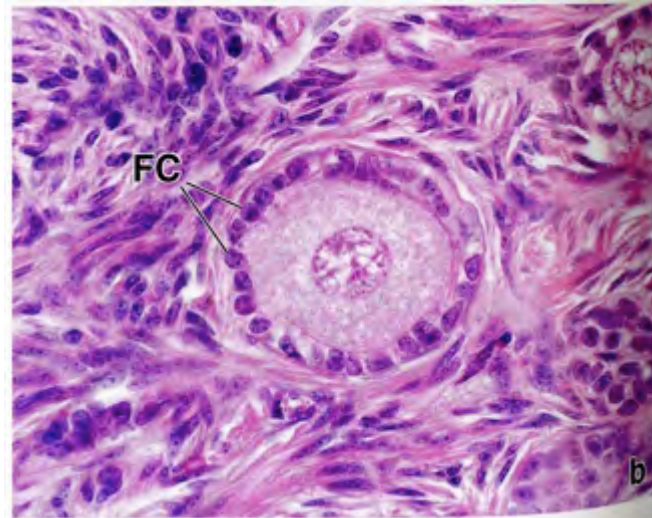
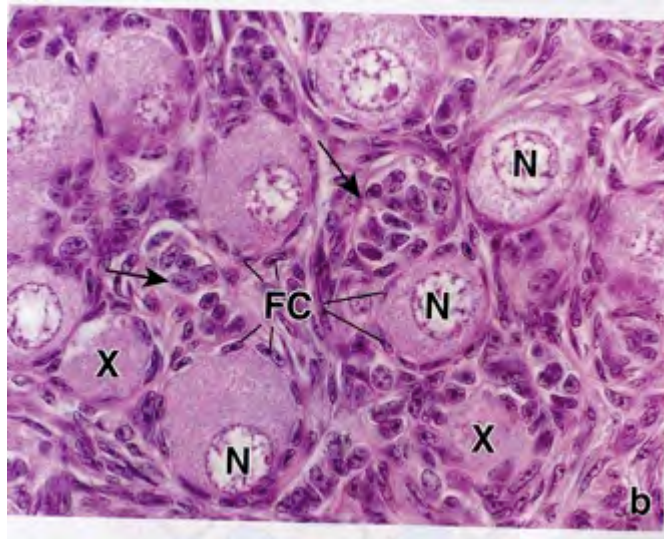
If there is no pregnancy, corpus luteum is cleared





Histology of Human Ovary (MS slide collection)





Stages of development  
of the ovarian follicle

Fluid in antrum of secondary follicle



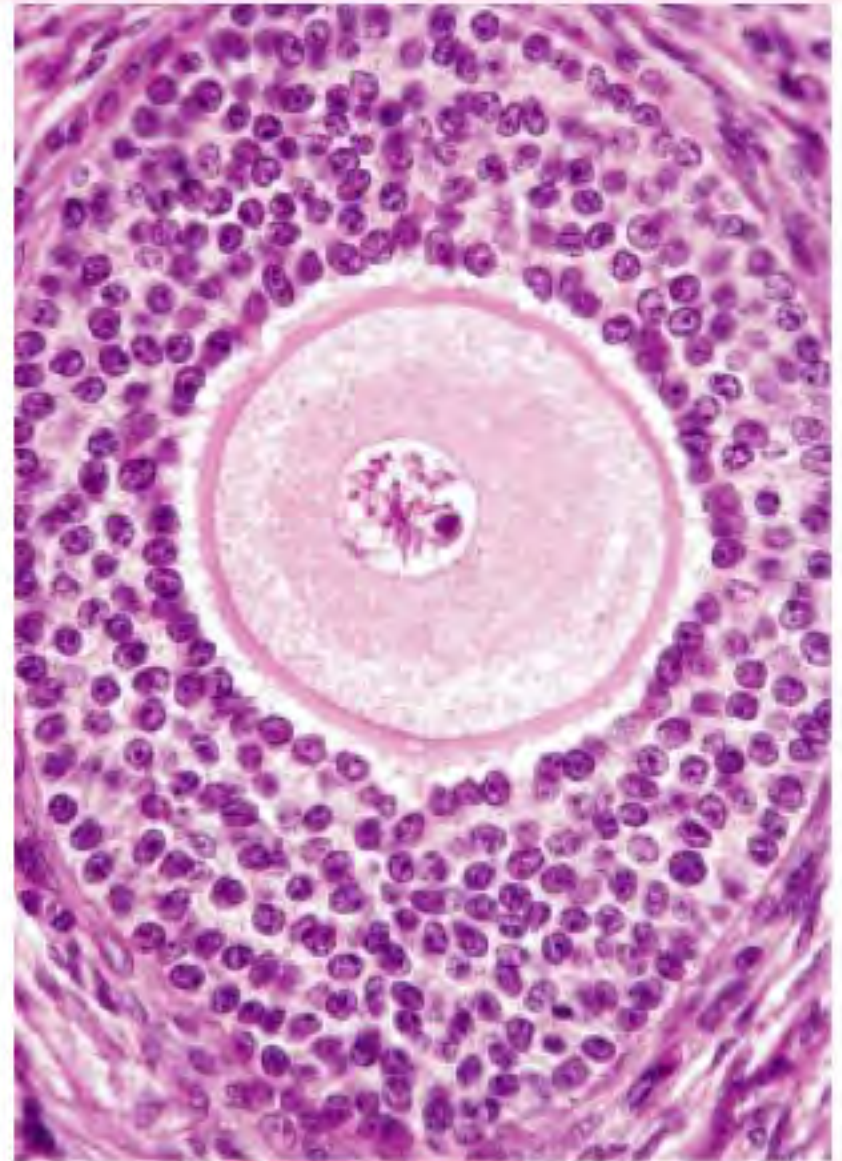
# Stages of Ovarian follicle maturation

## Primary Follicle

- oocyte
- zona pellucida
- granulosa cells
- theca folliculi

The **Zona Pellucida**: an acellular acidophilic glycoprotein ring that surrounds the oocyte

The primary follicle increases in size with the proliferation of the granulosa cells under the influence of FSH, Growth factors and calcium ions

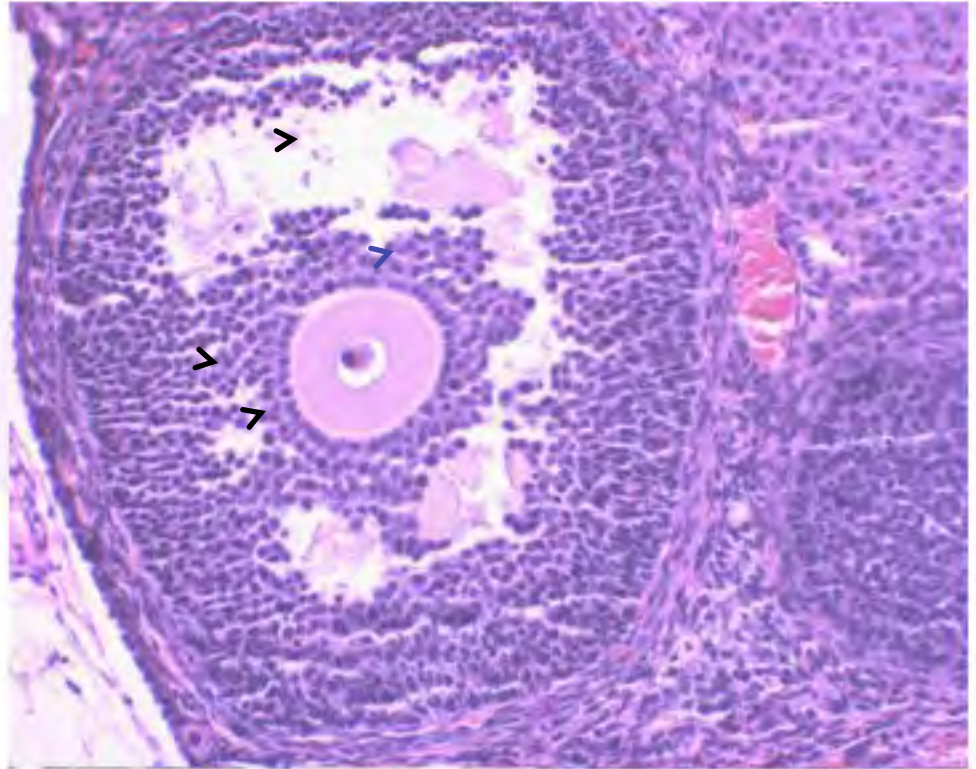




# Stages of Ovarian follicle maturation

**liquor folliculi**  
**cumulus oophorus**  
**corona radiata**

Zona pellucida



**Mature or Graafian follicle**  
**(~10 mm diameter)**

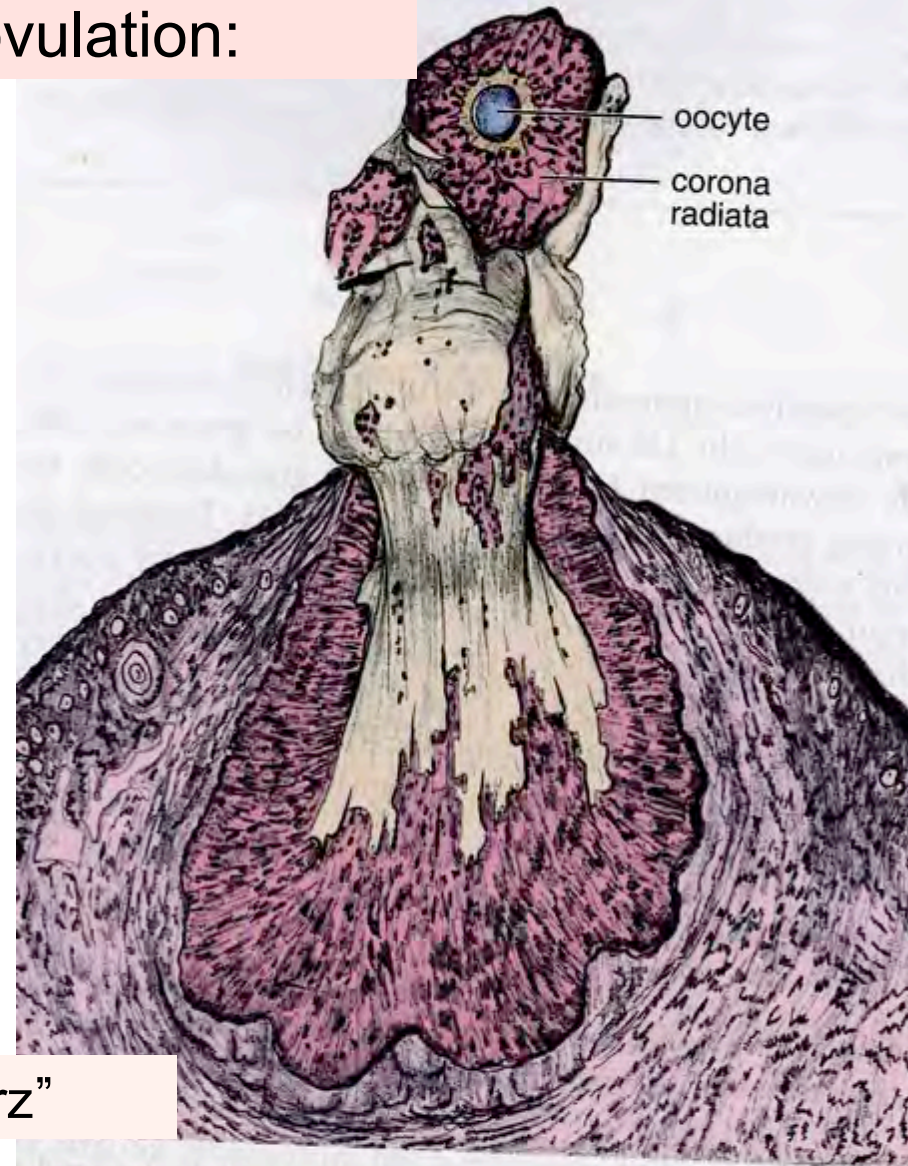
## Mature or Graafian follicle and ovulation:

Bulges from the ovarian surface

The Primary oocyte resumes the First Meiotic division, forming the Secondary oocyte and the first polar body

Just as the Secondary oocyte enters the Second meiotic division, Ovulation occurs

“mittelschmerz”



**FIGURE 23.8 Ovulation.** This drawing shows a rabbit oocyte, surrounded by the cumulus oophorus, being expelled from the ruptured ovarian follicle. (Based on Weiss L, Greep RO. Histology, 4th ed. New York: McGraw-Hill, 1977.)



# OVARIES



Corpus luteum (yellow body --after ovulation)

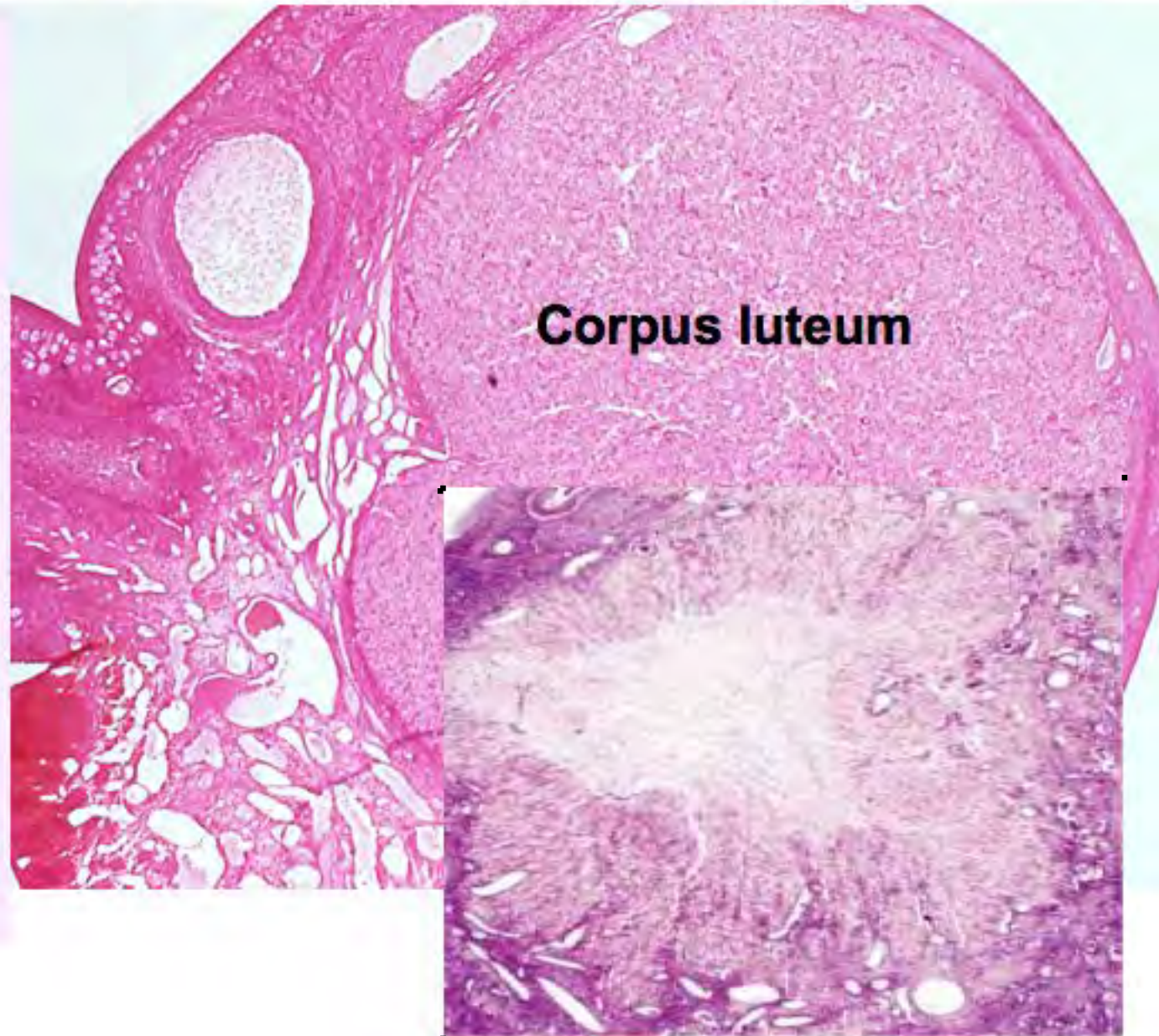
<http://library.med.utah.edu/WebPath/FEMHTML/FEM045.html>

# Stages of Ovarian follicle maturation

Following ovulation, the follicle the **corpus luteum** starts producing **progestogens**.

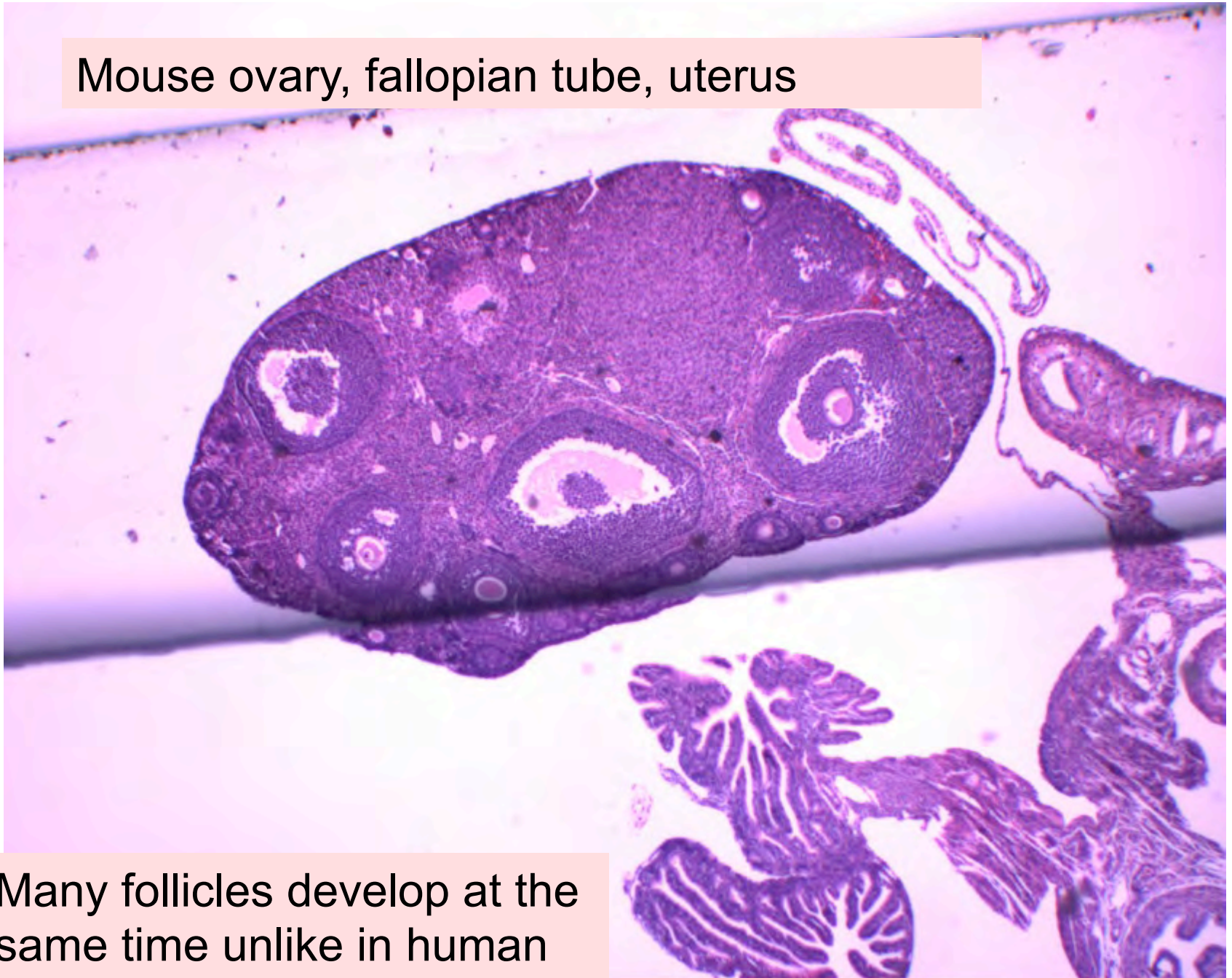
If fertilization does not occur, it undergoes atresia after 10-14 days

If fertilization does occur, it persists for about 6 months, stimulated by **placental human chorionic gonadotropin**.



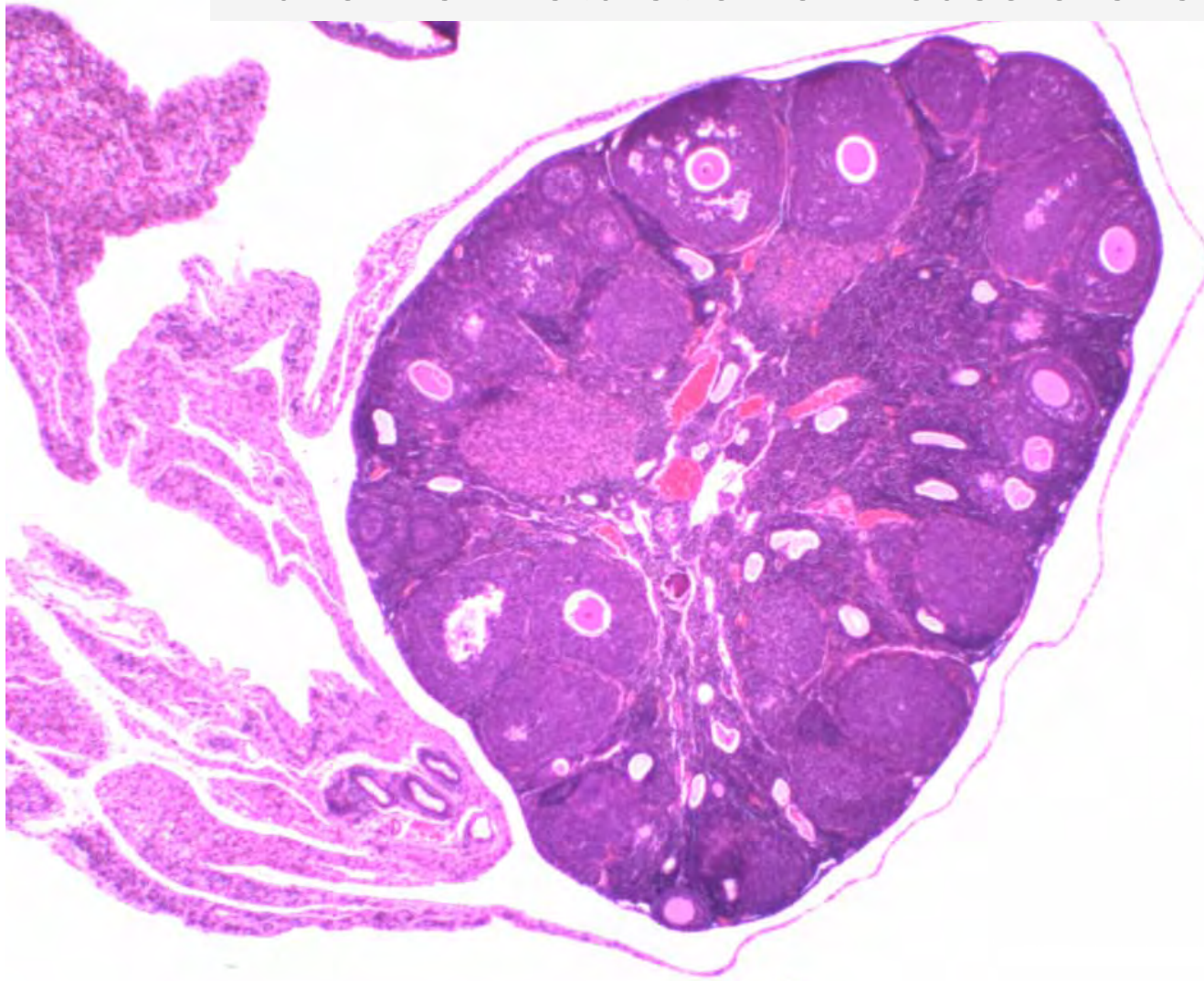


## Mouse ovary, fallopian tube, uterus

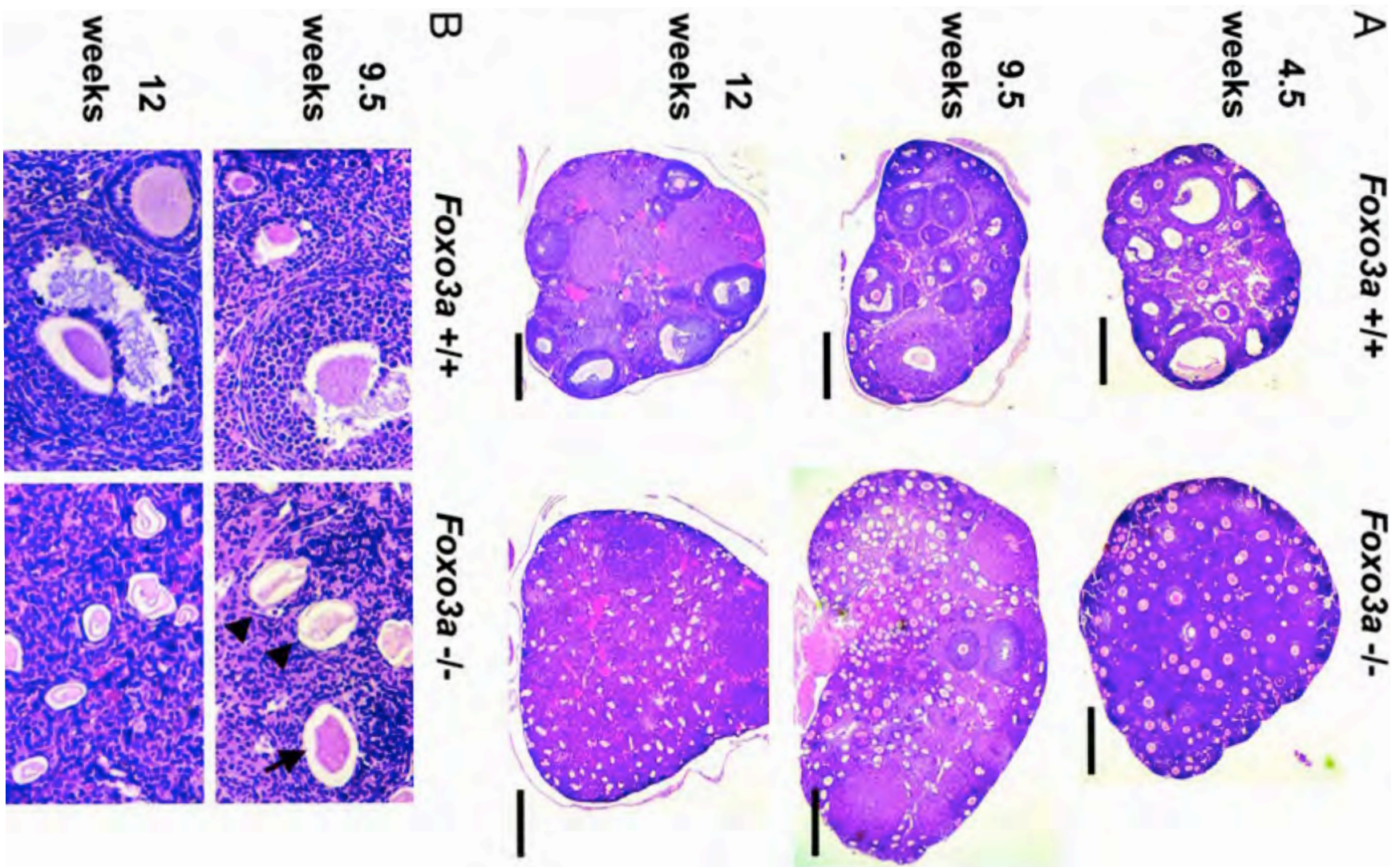


Many follicles develop at the same time unlike in human

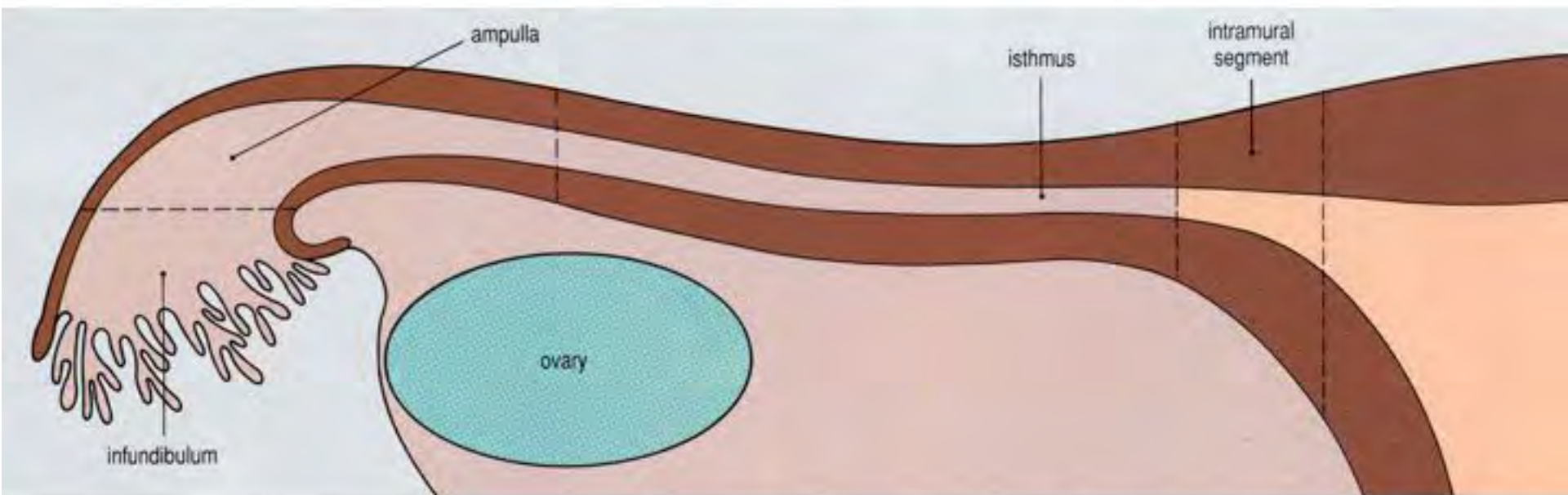
## Abnormal maturation of mouse ovarian follicles







# Fallopian Tube



The Fallopian tubes are paired structures (upto about 12 cm in length and 7 mm in diameter)

They are the site of fertilization and development of the early embryo to the blastocyst stage

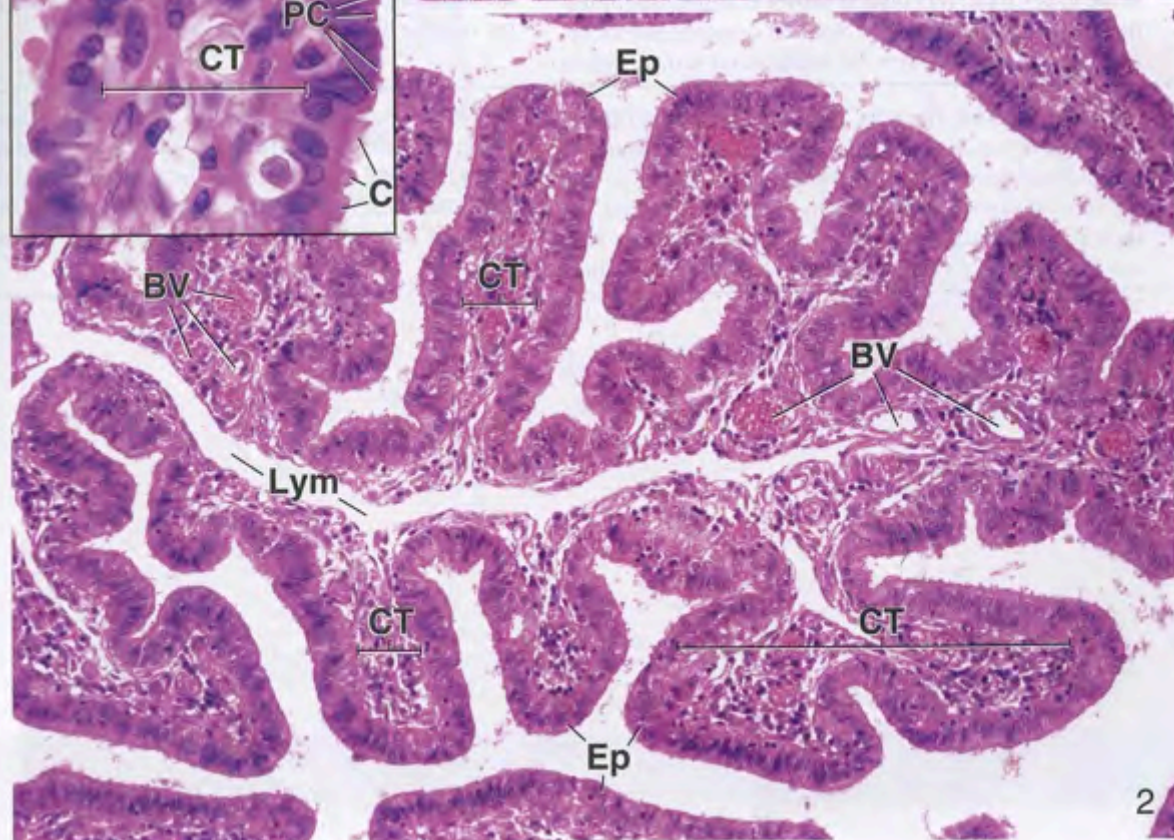
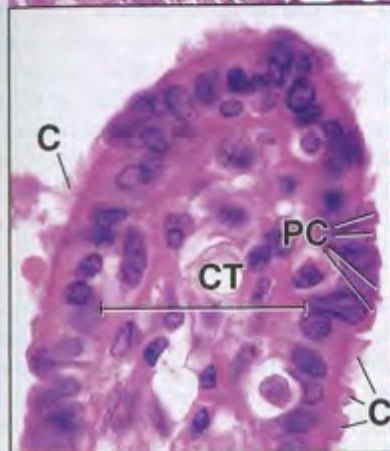
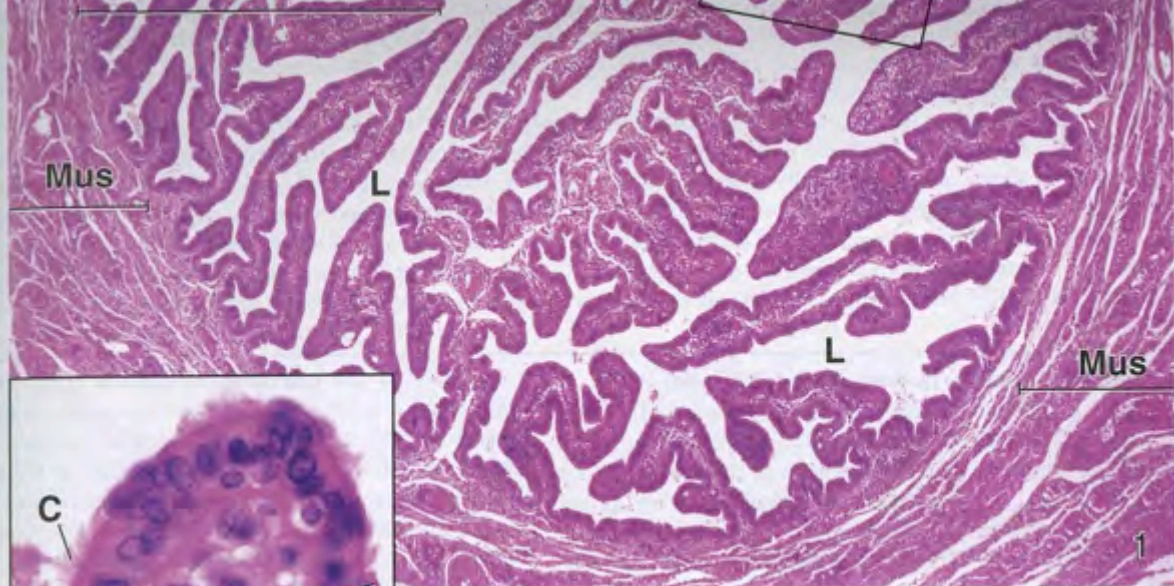
4 regions:

- infundibulum
- ampulla
- isthmus
- intramural region





Fallopian tube at  
different  
magnifications



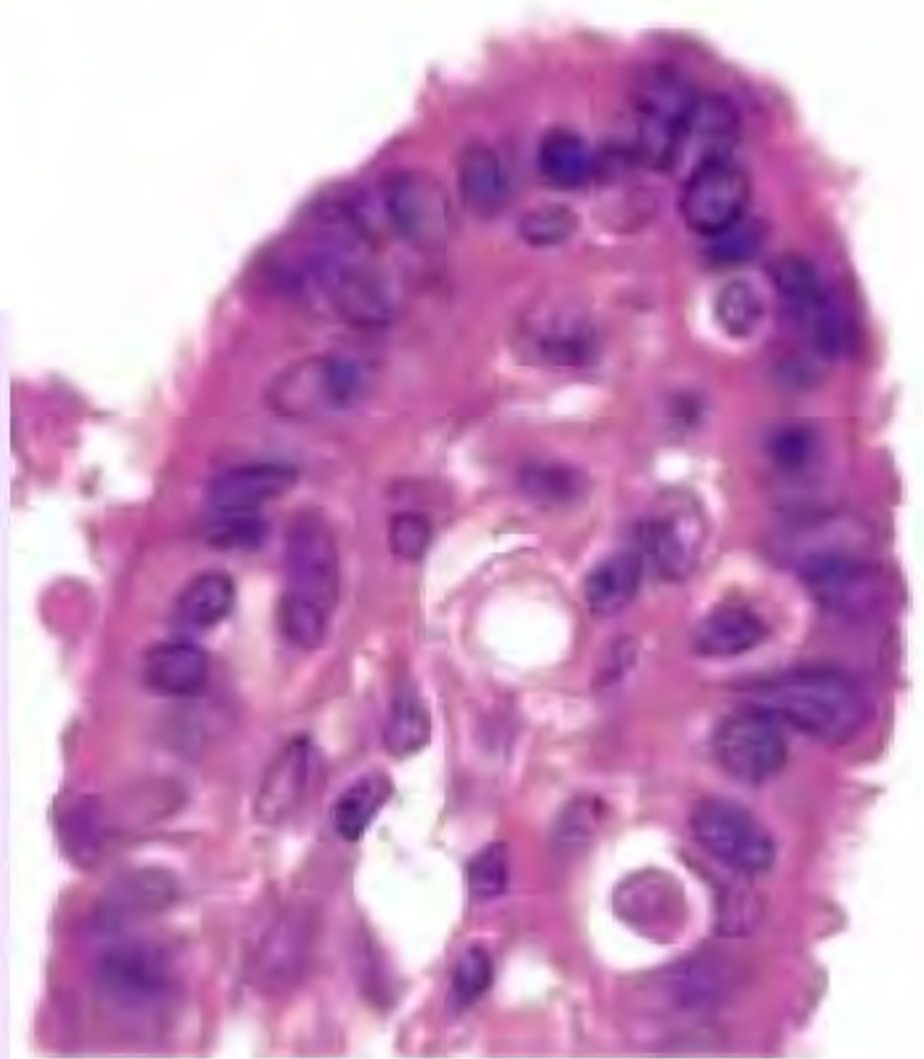


# Fallopian Tube, epithelium

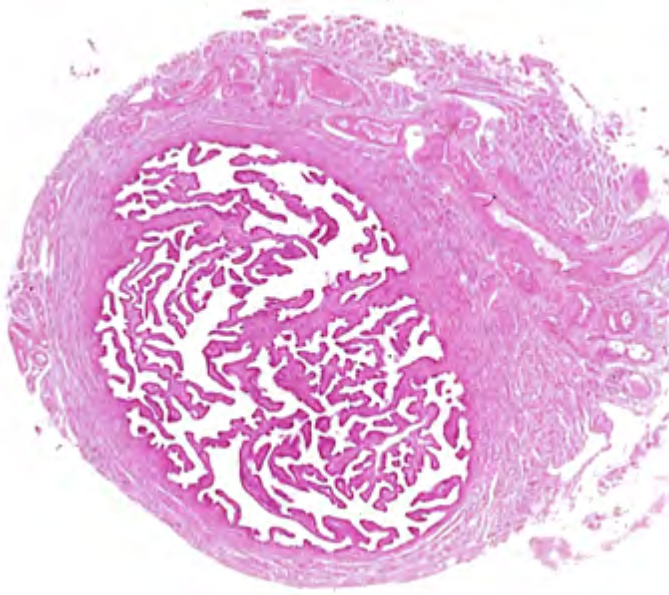
Mucosa consists of simple columnar epithelium

## 2 types of epithelial cells:

- **ciliated cells**, especially in the fimbrial portion to assist with *ovum transport* (and *sperm transport*)
- **secretory cells** (peg cells):
  - ovum nutrition*
  - sperm capacitation*





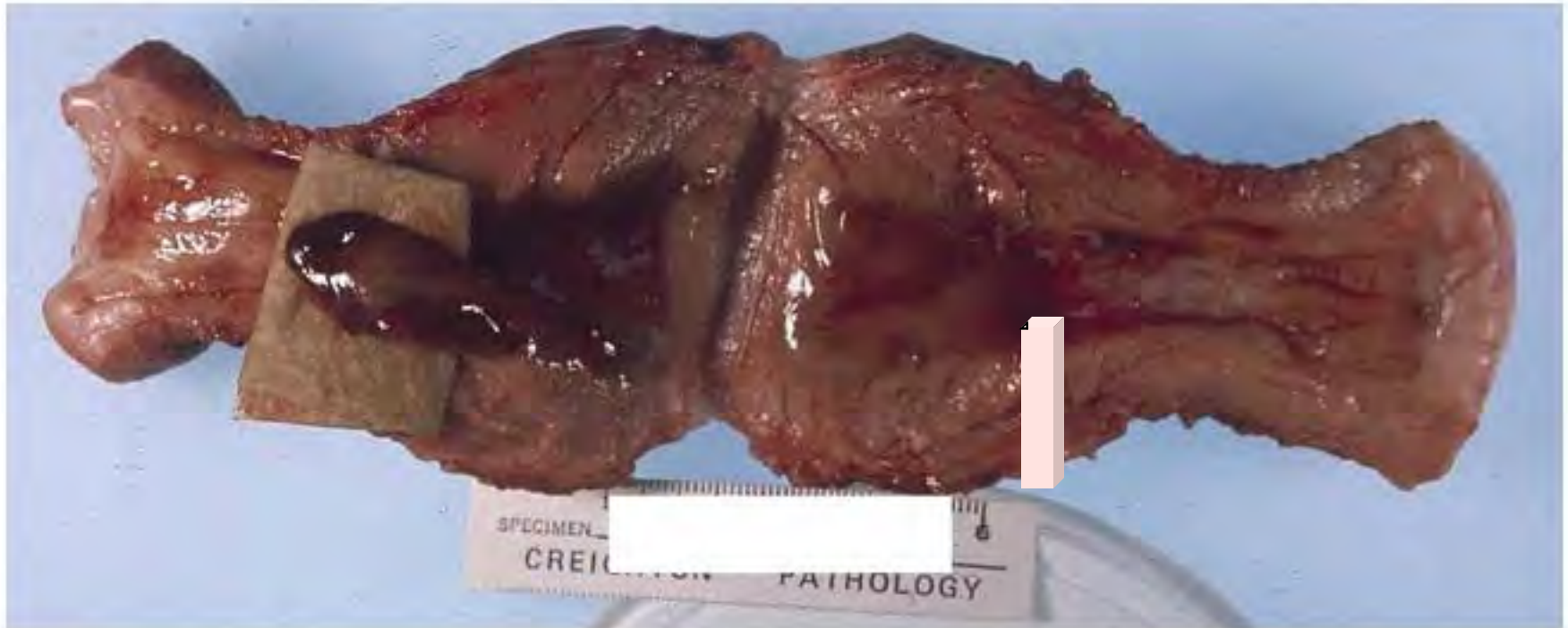


Normal human fallopian tube with highly folded mucosa at the ampulla which is the usual site of fertilization

Abnormal human fallopian tube which are both blocked and dilated after infectious events



# UTERUS



The Uterus is a hollow, muscular pear-shaped organ (~6.5 cm long, 3.5 cm wide and 2.5 cm thick)

It has a body with an upper fundus and a distal cervix

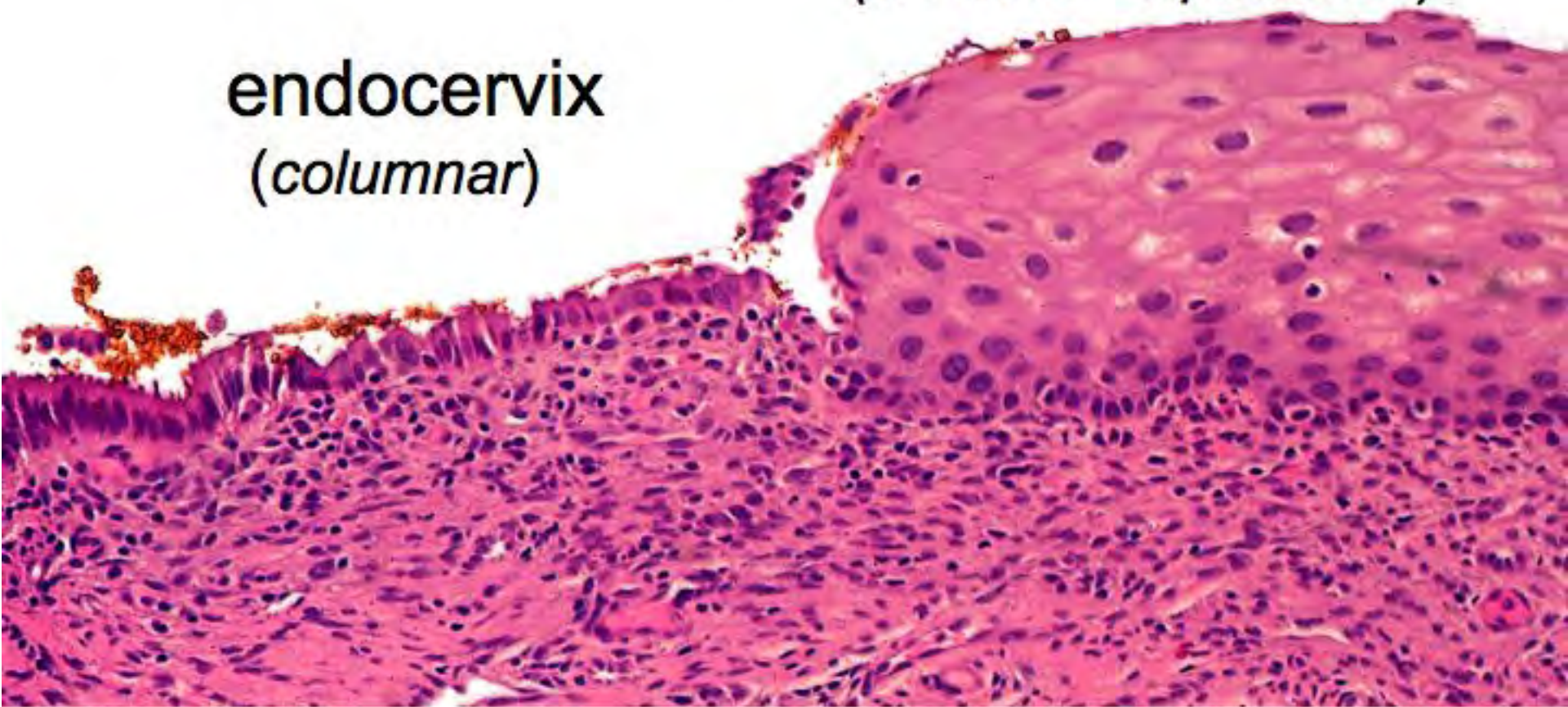
There are 3 layers: the endometrium, myometrium and serosa



# Abrupt squamo-columnar junction

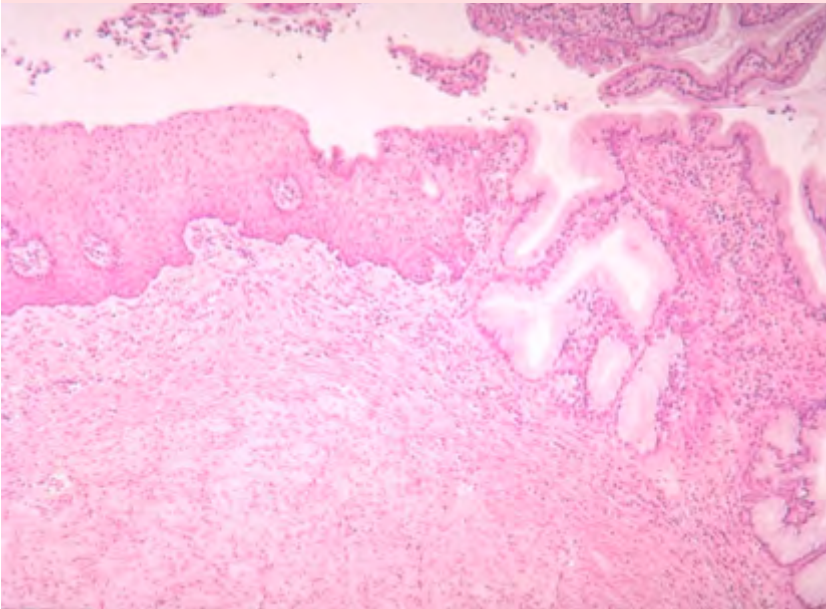
Exocervix:  
*(stratified squamous)*

endocervix  
*(columnar)*

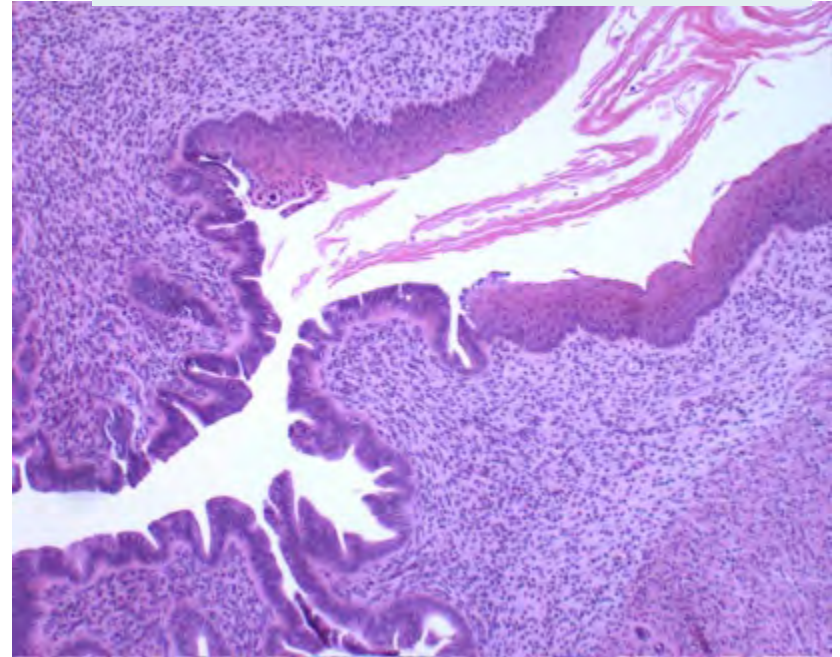


# Squamo-columnar junction of cervix

Human ecto and endo cervix



Mouse endo and ecto cervix





## Differences from Human Menstrual Cycle

One of the main differences between the rodent and the human cycle, other than the overall time it takes for a full cycle, is that the peaks of estrogens and progesterone are typically separated in humans, whereas these overlap in rodent.

In the mouse, the estrous cycle is divided into 4 stages (proestrus, estrus, metestrus, and diestrus) and repeats every 4 to 5 days unless interrupted by pregnancy, pseudopregnancy, or anestrus.

[http://www.biobserve.com/downloads/maria-gulinello/  
Rodent-Estrous-Cycle.pdf](http://www.biobserve.com/downloads/maria-gulinello/Rodent-Estrous-Cycle.pdf)

**Dr. Maria Gulinello**  
**Behavioral Core Facility**  
**Albert Einstein College of Medicine**

**Behavioral Core Protocols and Training**

**Estrous Cycle Staging**



**Albert Einstein College of Medicine**  
**Dominick P. Purpura Department of Neuroscience**  
1410 Pelham Pkwy S R912F Bronx, NY 10461  
718 - 430 4042  
mgulin@aecom.yu.edu











## **The Rodent Estrous Cycle**

Rats and mice are spontaneous ovulators – i.e. they do not need the presence of males to induce ovulation.

There is some evidence the stimulation from male rodents can induce and or/hasten ovulation, but it is not obligatory. Rats and mice typically have a 4-6 day cycle which consists of 4 stages – estrus, proestrus, diestrus I (a.k.a metestrus) and diestrus II.

**Proestrus** is defined by the presence of nucleated epithelial cells that are round. However, sometimes these cornify rapidly, especially in mice. If you are having trouble distinguishing between P and E, you should probably consider any slide in which nucleated cell predominate to be P, even if these cells are not the smooth round shape you are expecting. **Estrus** is characterized by non-nucleated, cornified epithelial cells. **Metestrus** (or Diestrus I) typically has a low cell number, often with a lot of cell debris. **Diestrus II** contains mostly lymphocytes.

Time of day	1400	1400	1400	1400-1800
Hours after ovulation	12	36	60	84-88
	Estrus	Diestrus I	Diestrus II	Proestrus
Vaginal smear				
	Primary corpus luteum	Primary follicle	Secondary follicle	Graafian follicle
Follicular development				
Sexual behavior	—	—	—	+

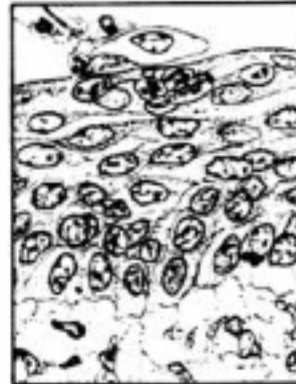




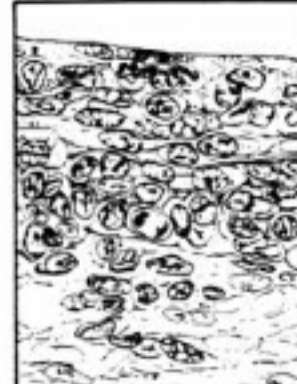
Proestrus



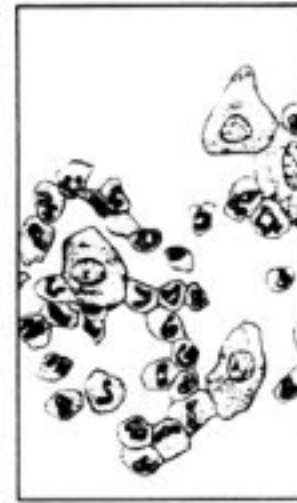
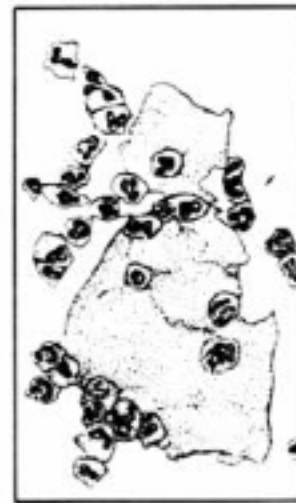
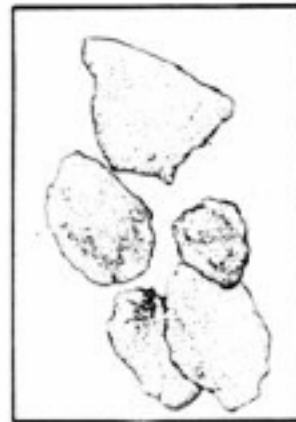
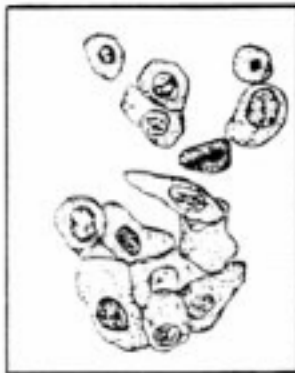
Estrus

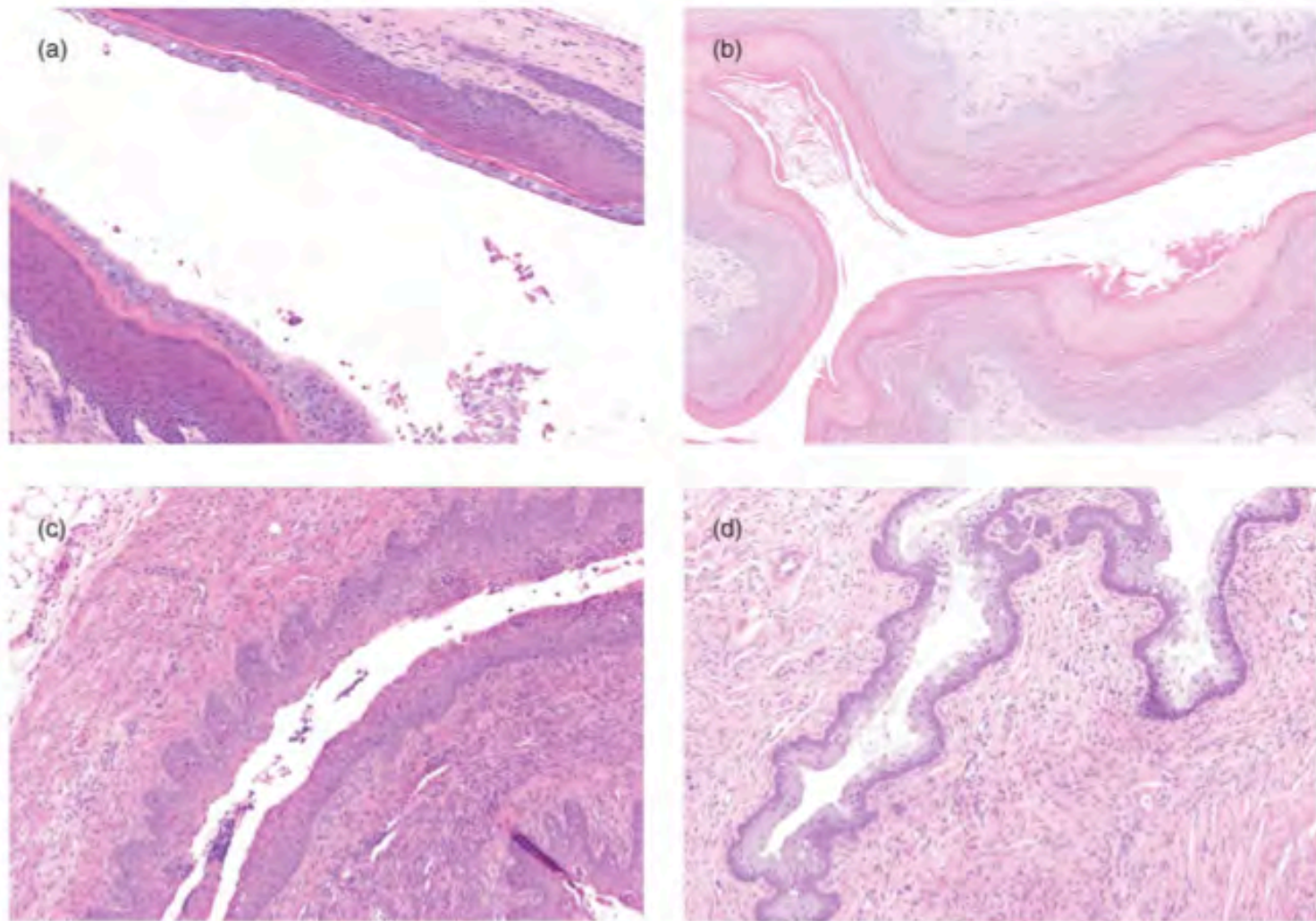


Metestrus



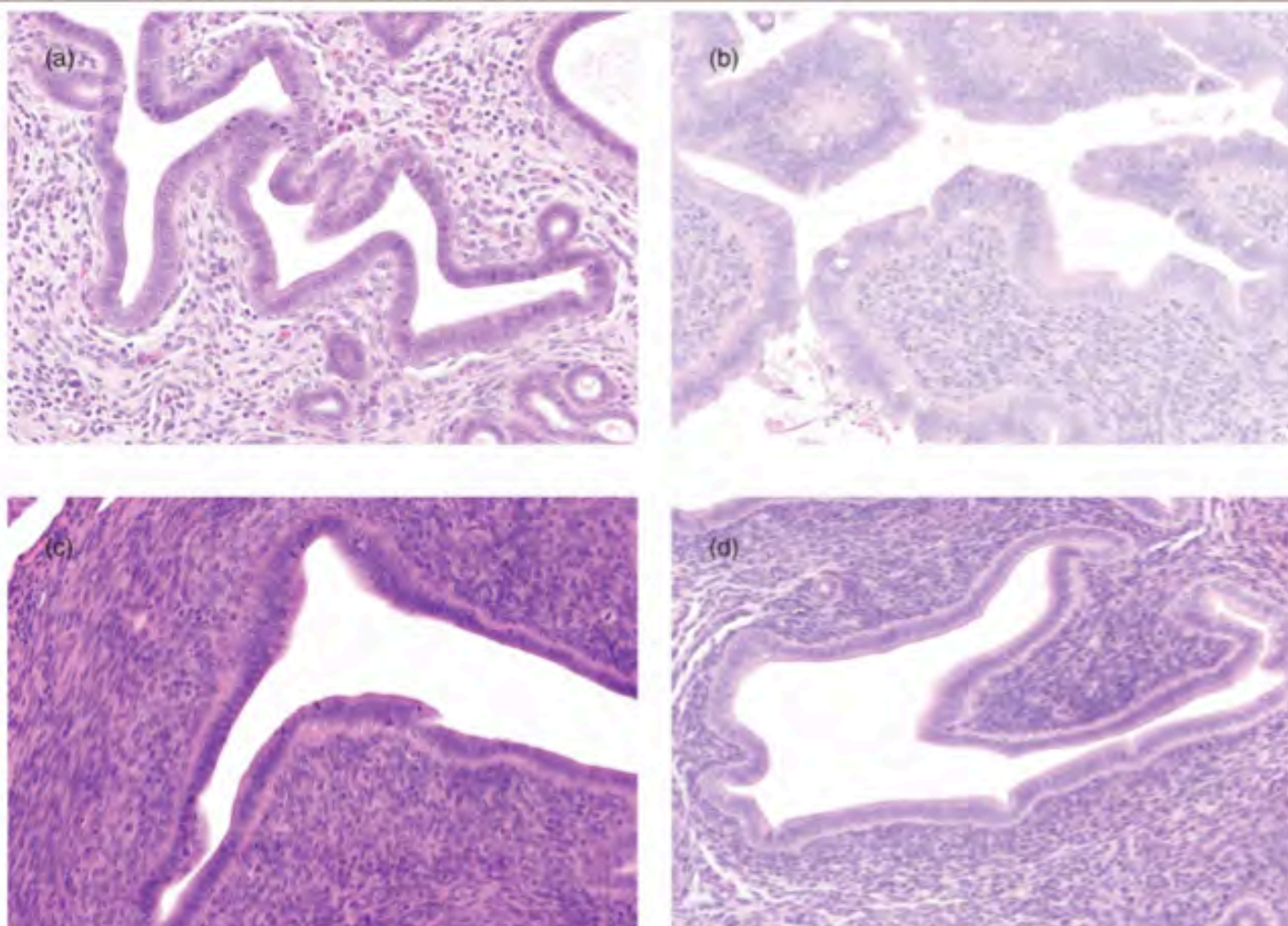
Diestrus





**Figure 6.12** Vaginal epithelium changes morphology with stage of oestrous cycle. (a) Proestrus, (b) oestrus, (c) metoestrus, (d) dioestrus.

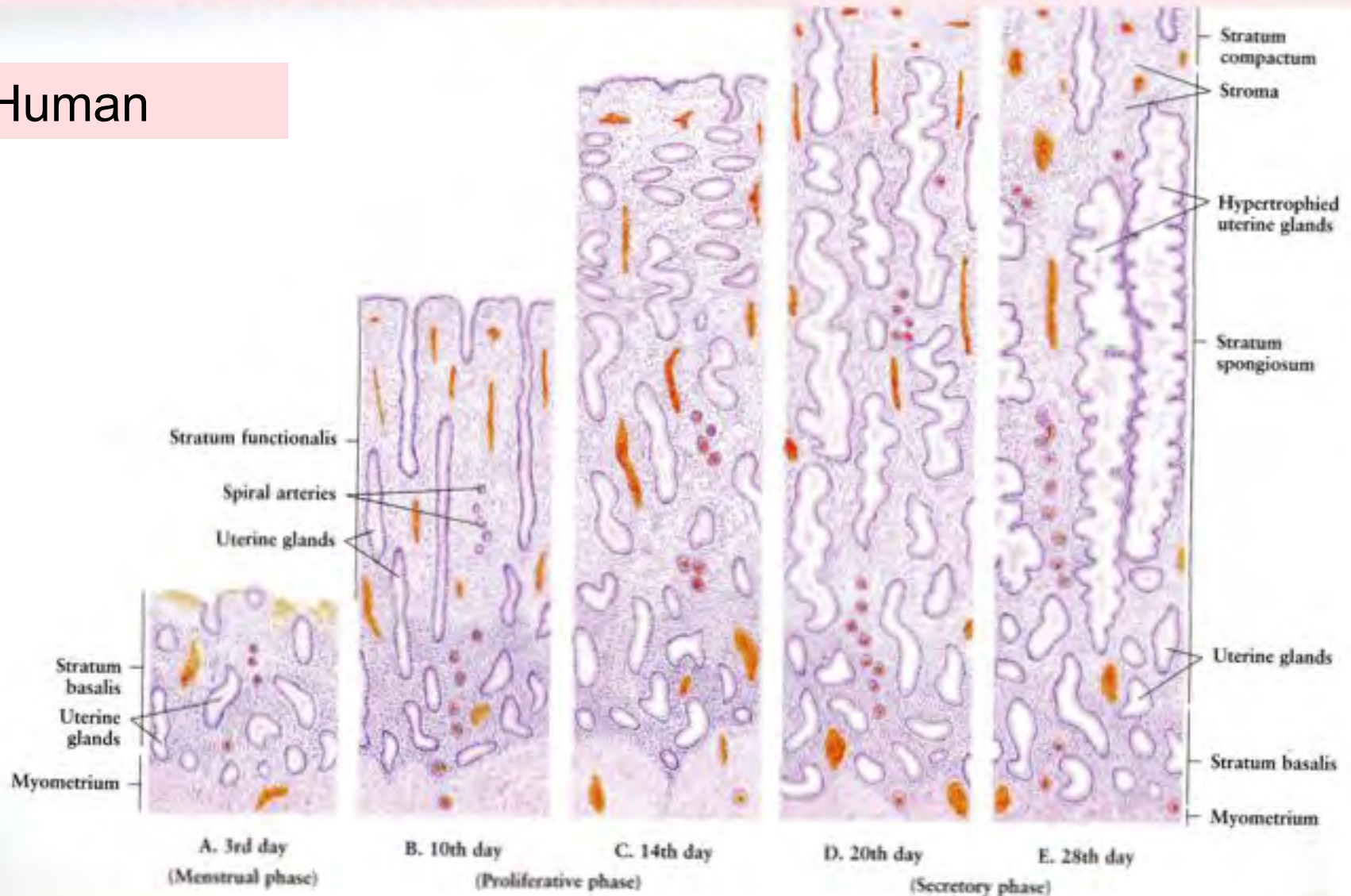




**Figure 6.10** Uterine endometrium changes morphology with stage of oestrous cycle. (a) During proestrus the epithelium may contain mitoses and the stroma may be oedematous, (b) epithelium starts to vacuolate and degenerate in oestrus, (c) mitoses are seen towards the end of metoestrus, (d) the epithelium is at its lowest in dioestrus.

# Uterine endometrium changes during the menstrual hormonal cycle

Human





Uterine  
endometrium  
changes during  
the human  
menstrual cycle

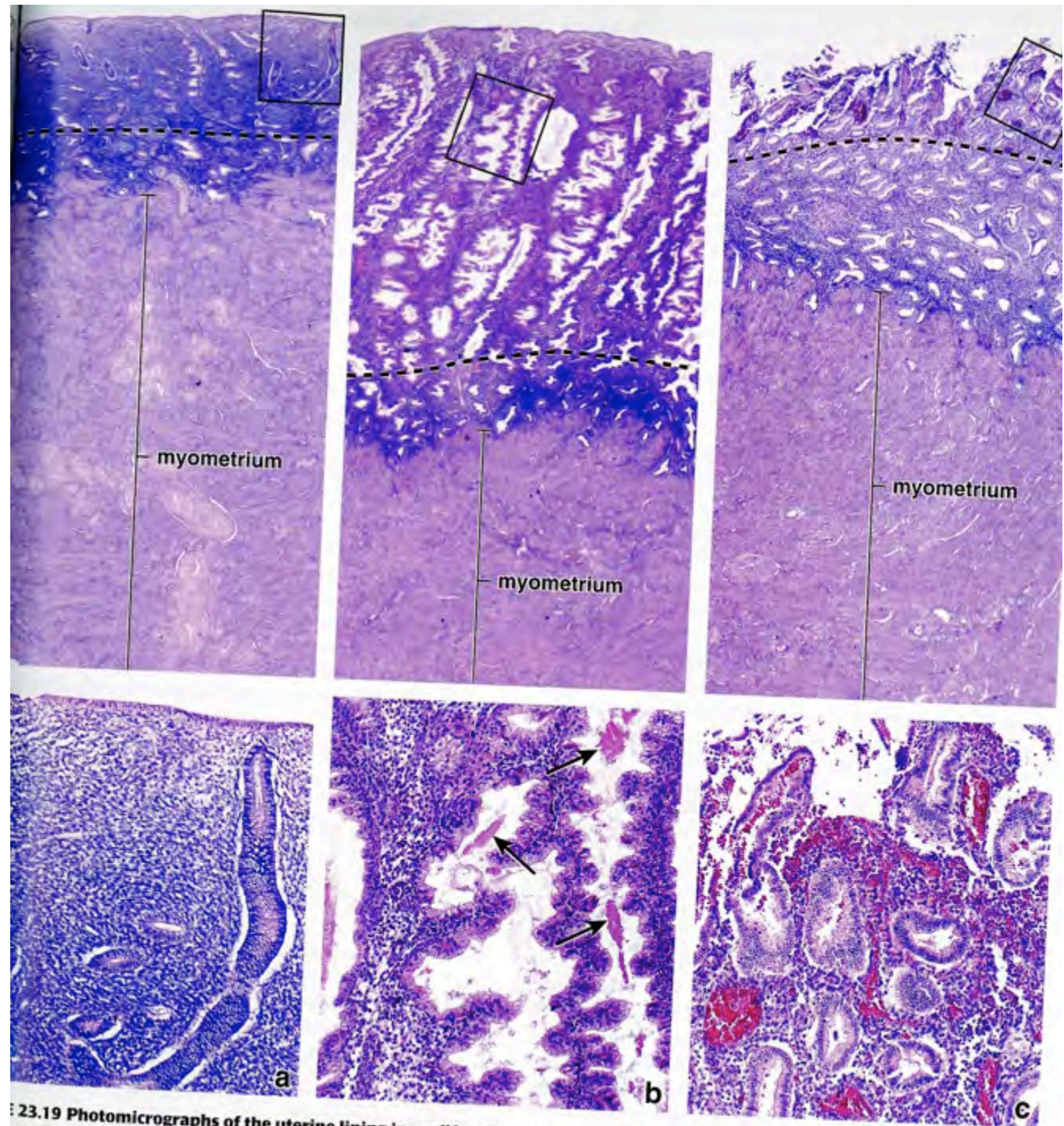
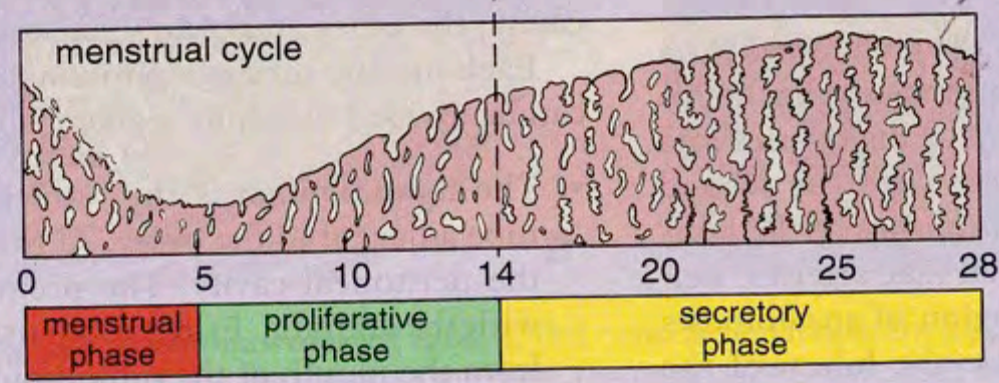
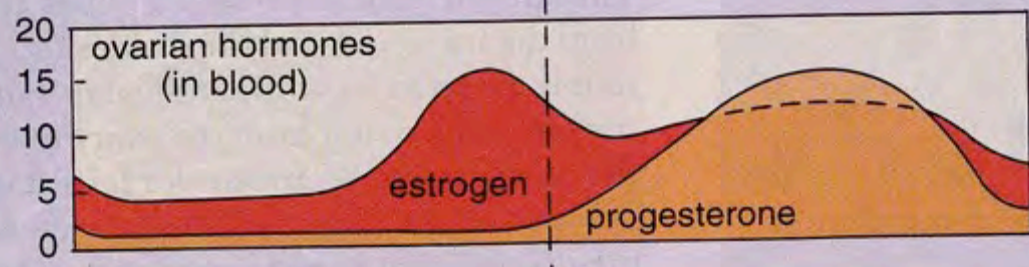
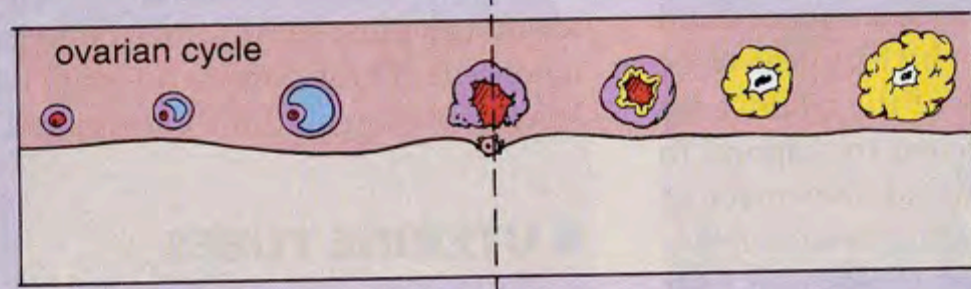
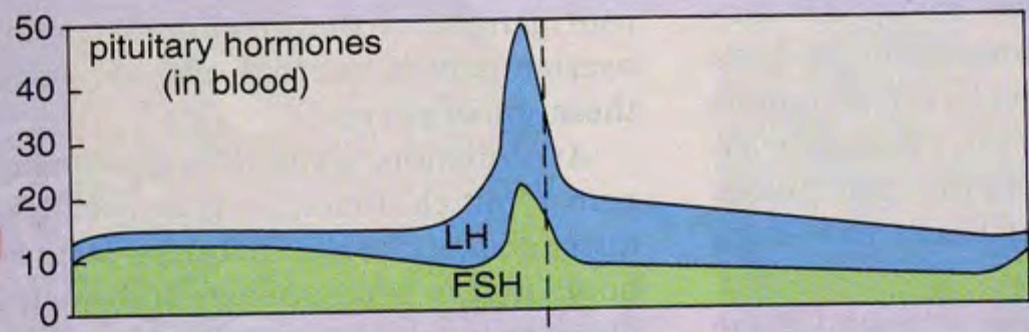


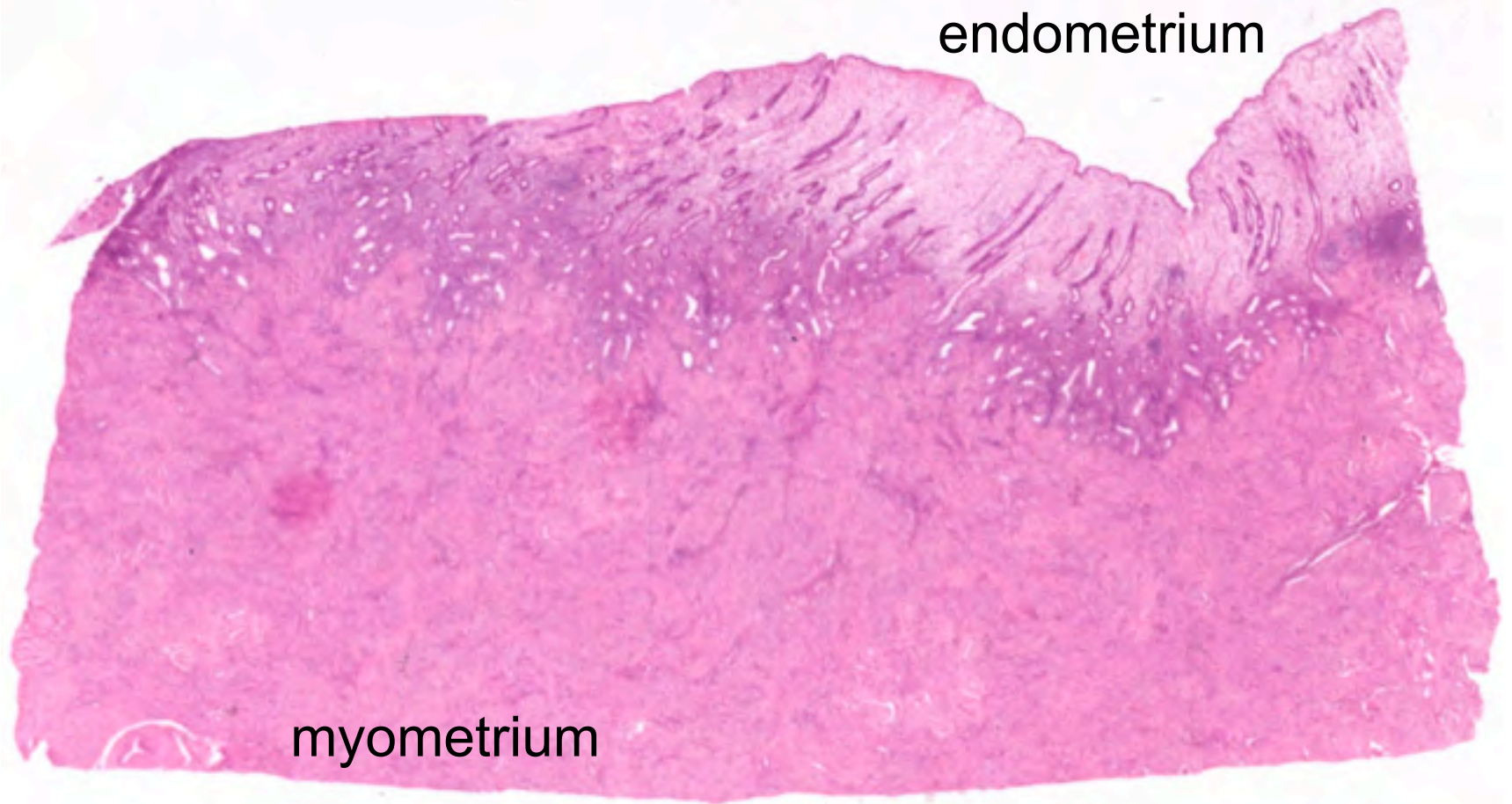
FIGURE 23.19 Photomicrographs of the uterine lining in proliferative, secretory, and menstrual phases of the human menstrual cycle.



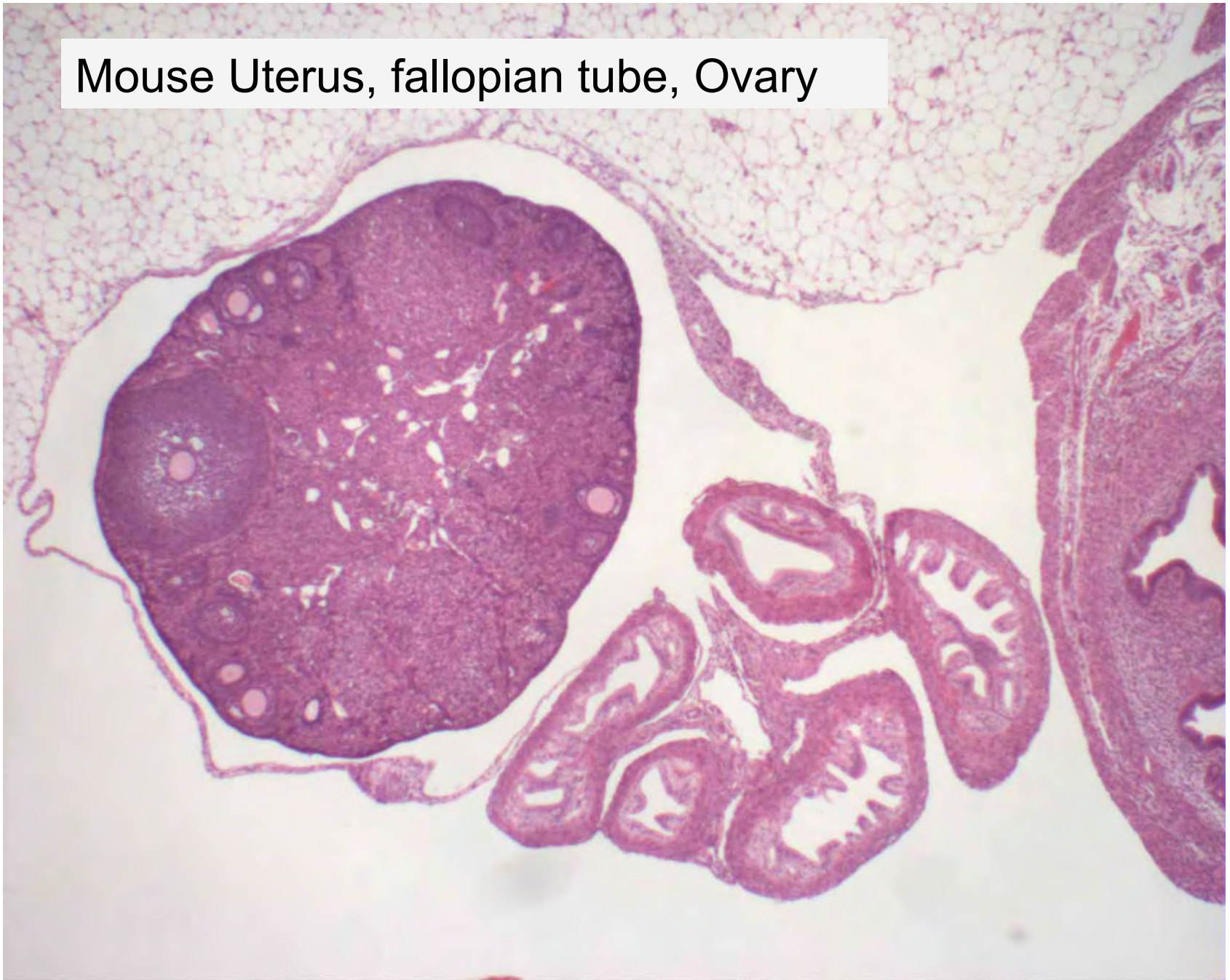




A section through a portion of the human uterus



Mouse Uterus, fallopian tube, Ovary

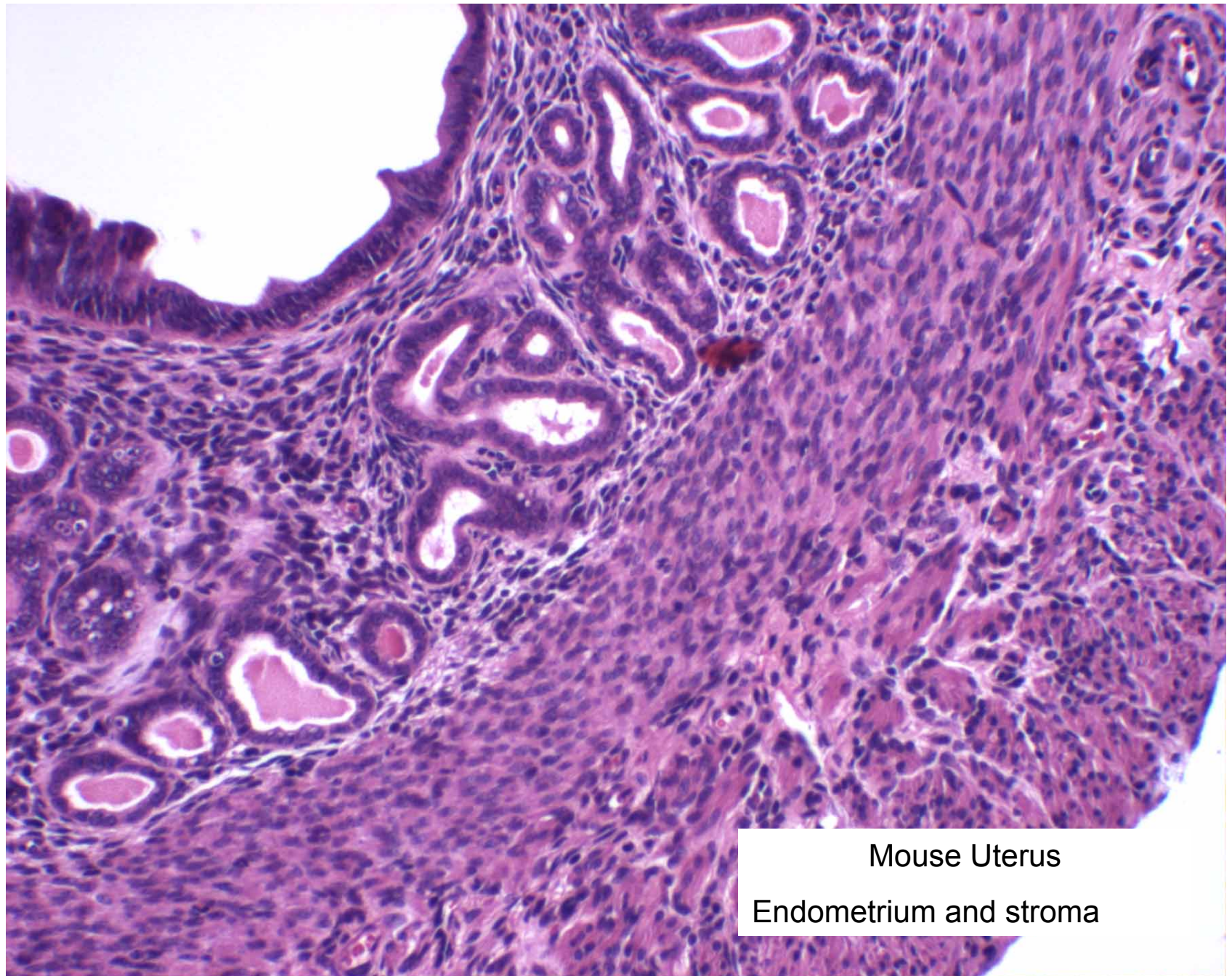






Mouse Uterus

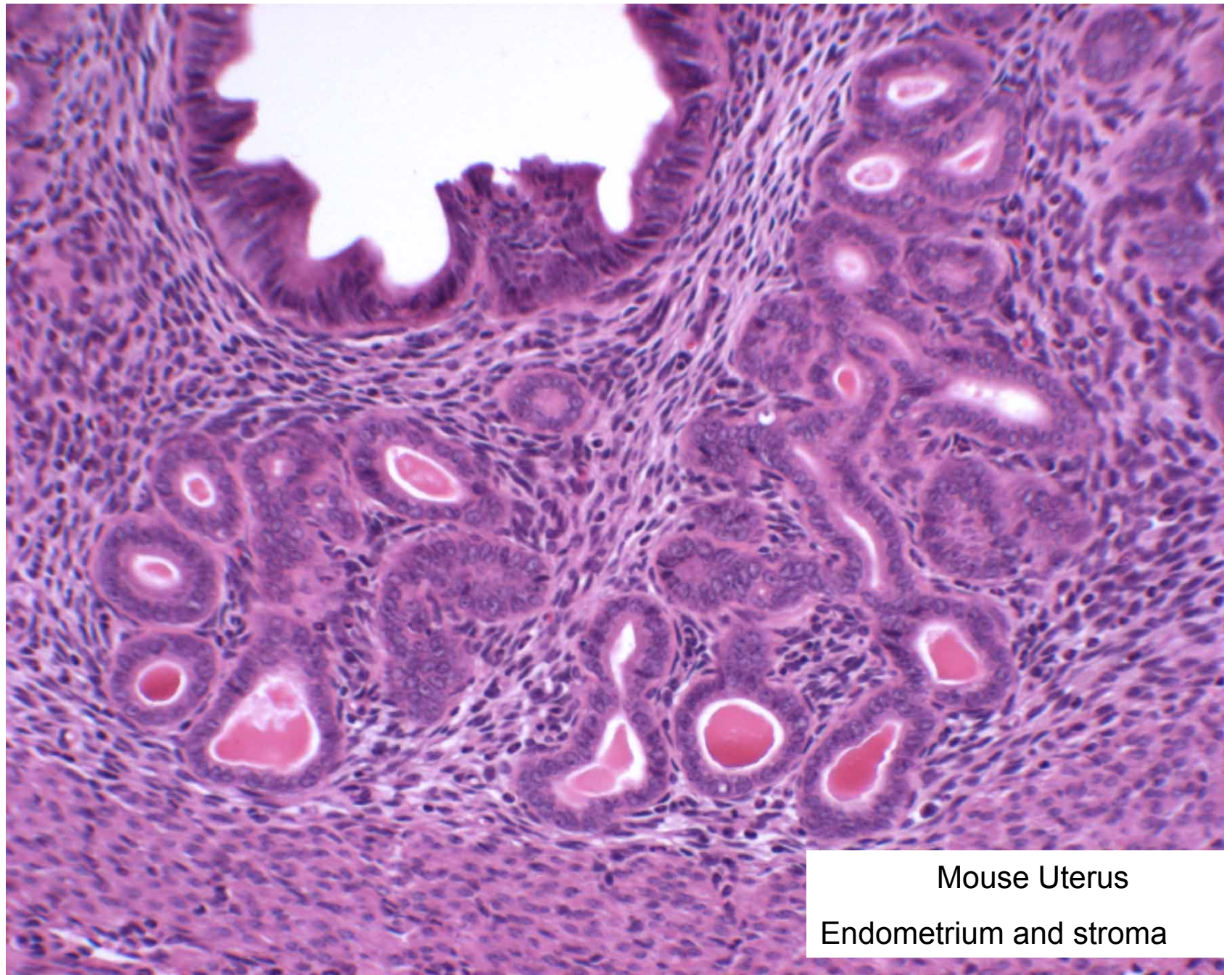




Mouse Uterus

Endometrium and stroma



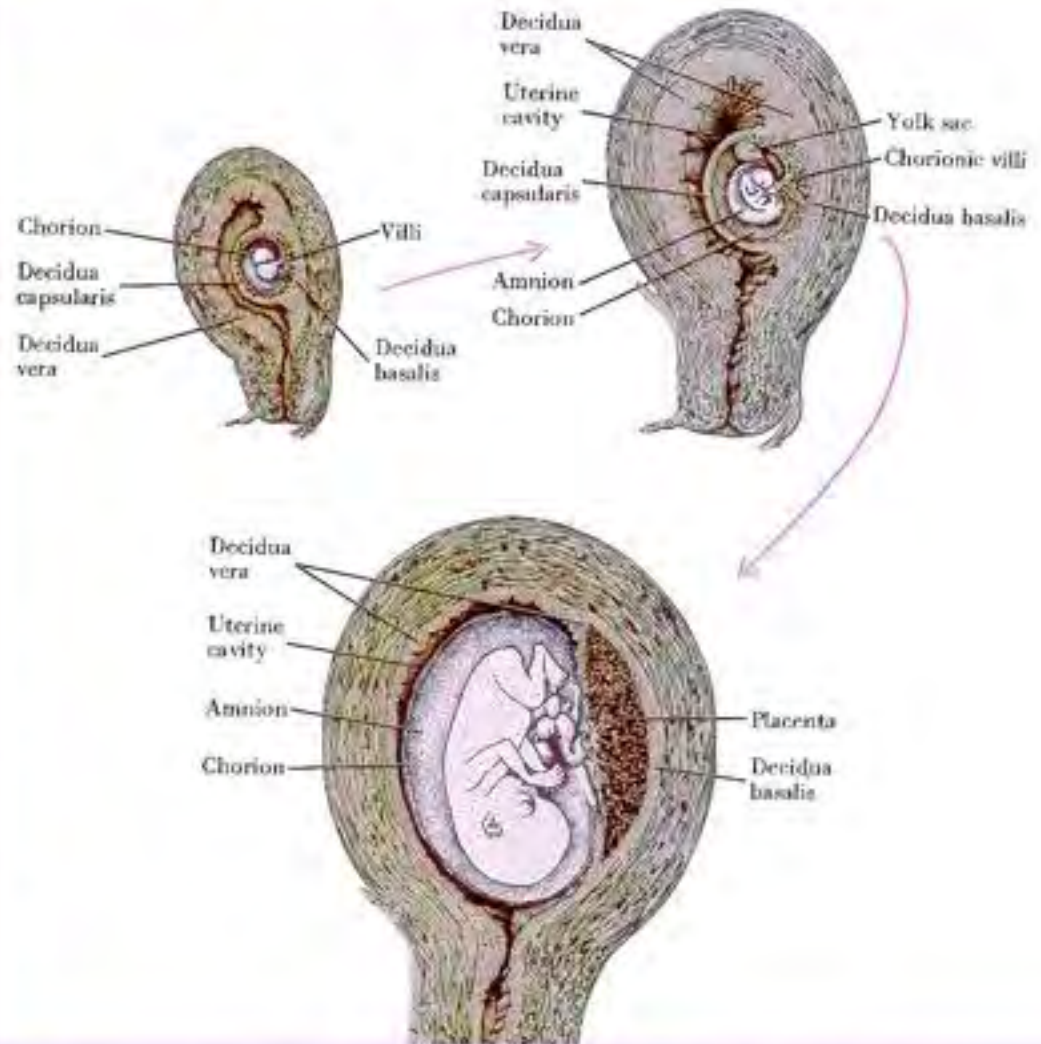


Mouse Uterus  
Endometrium and stroma



# Human Placenta

If fertilization and implantation occur, a gravid phase replaces the menstrual phase of the cycle and decline of the endometrium is delayed until after parturition. The endometrium is converted into decidual cells, which form the basal part of the placenta.



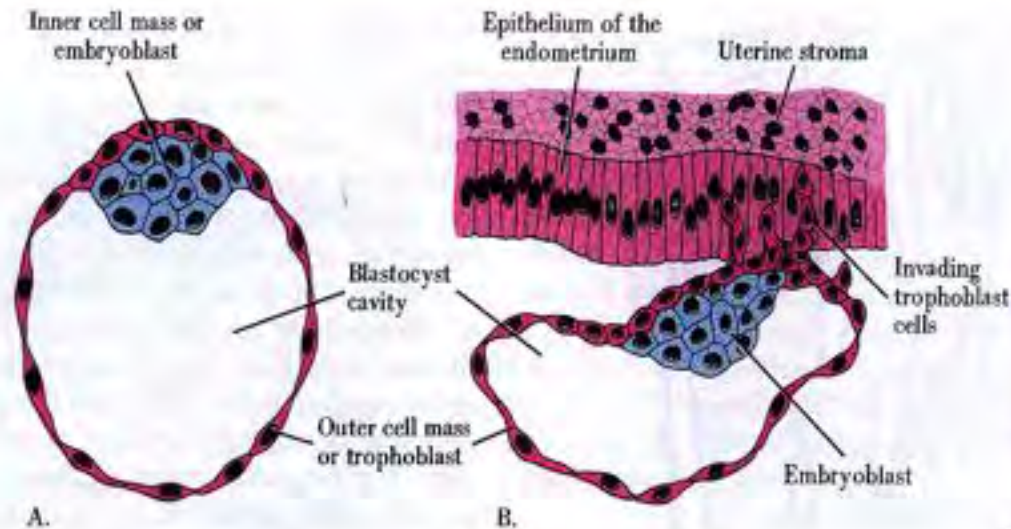
The placenta is a major endocrine organ producing steroid and protein hormones



# Human Placenta

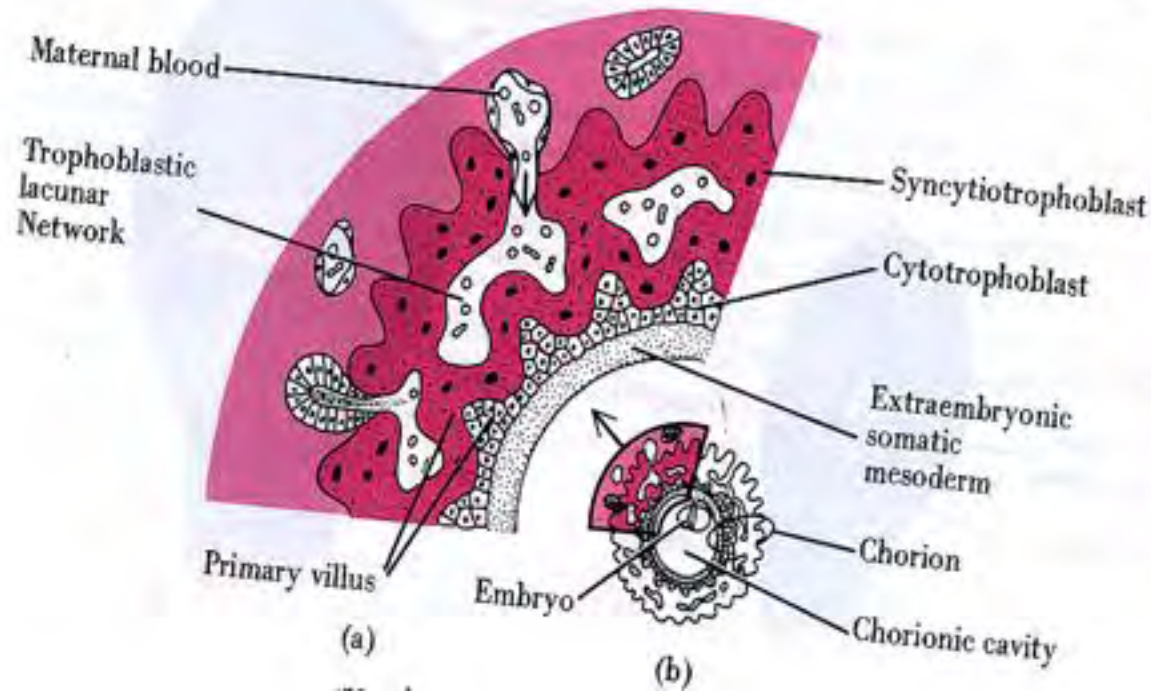
**Early stages of  
implantation**

**Human day 4, day 5**

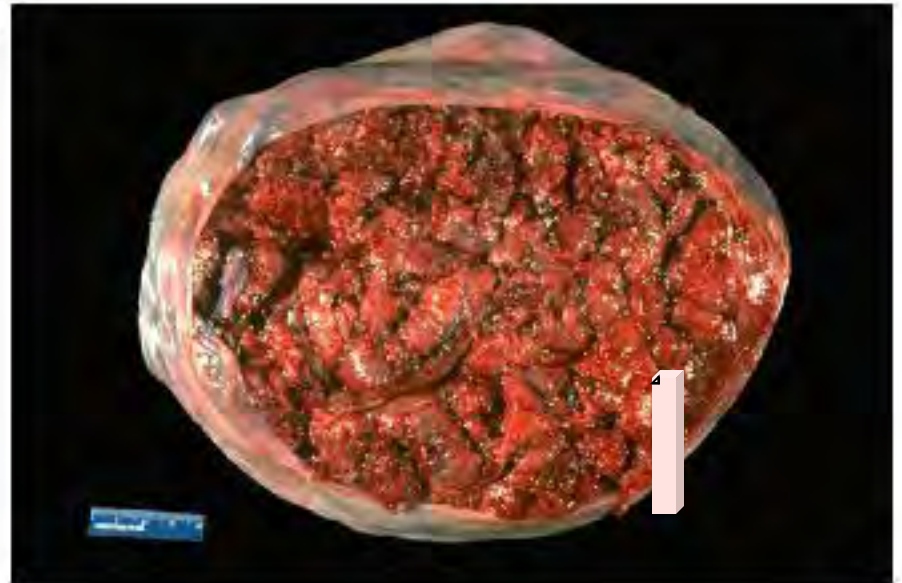


**Early stages of  
placental  
development**

**Human day 14**



# Human Placenta

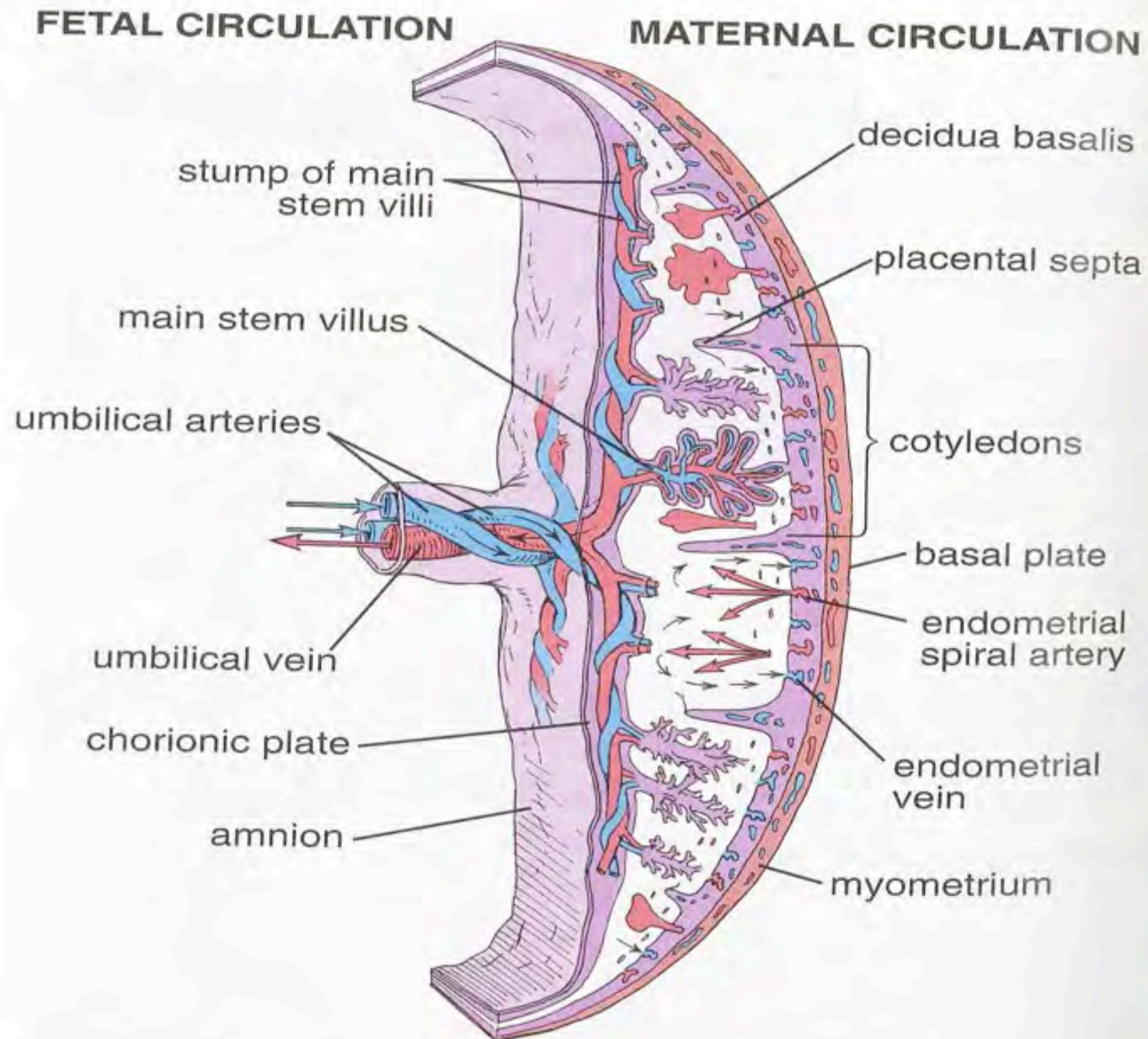


Fetal surface: Shiny, covered with amnion

Maternal surface: Uterine/decidual surface

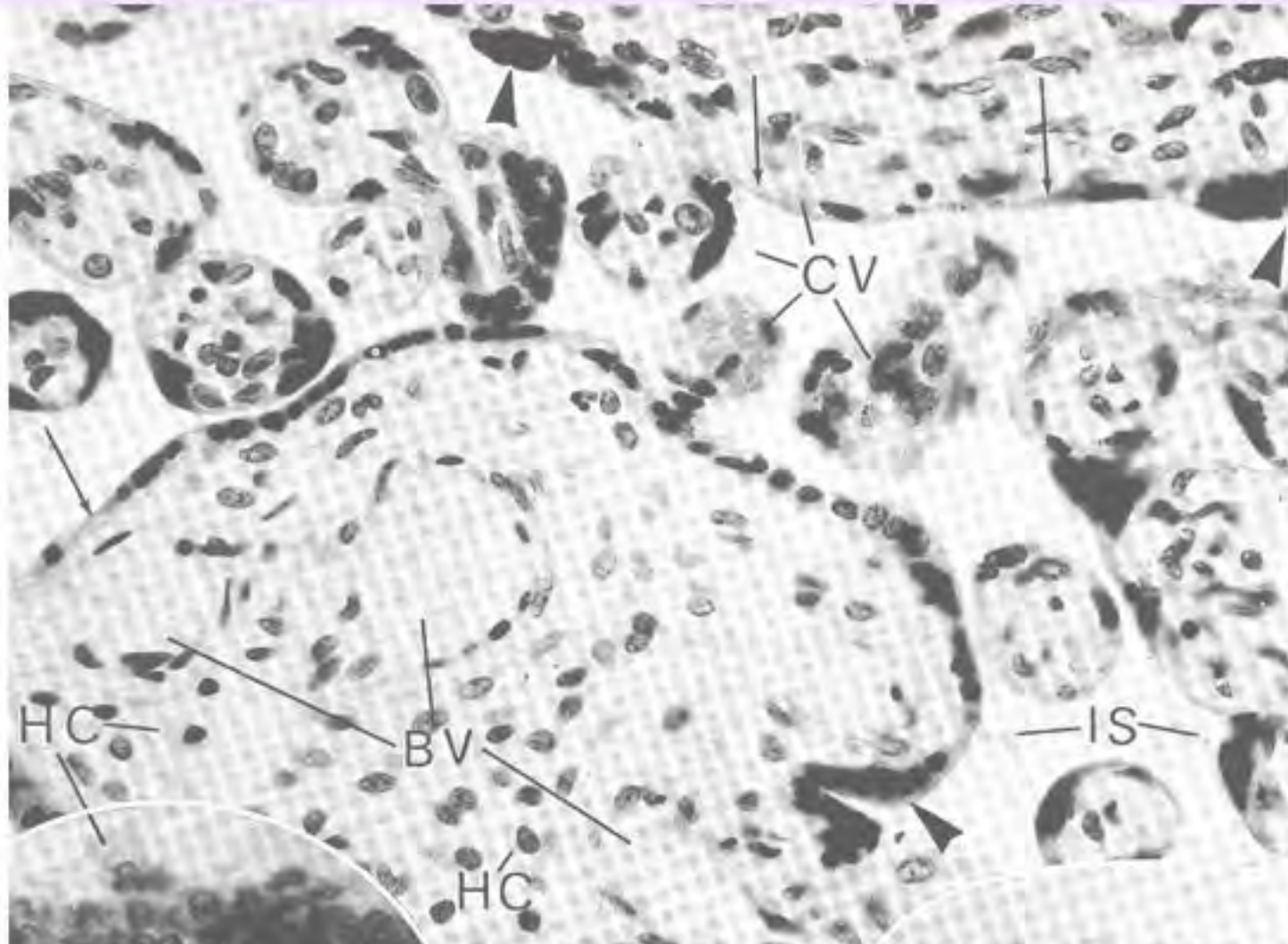


# Human Placenta : diagram



gittal section of the uterus (*left*) with the developing embryo shows the

# Histology of Human Placenta

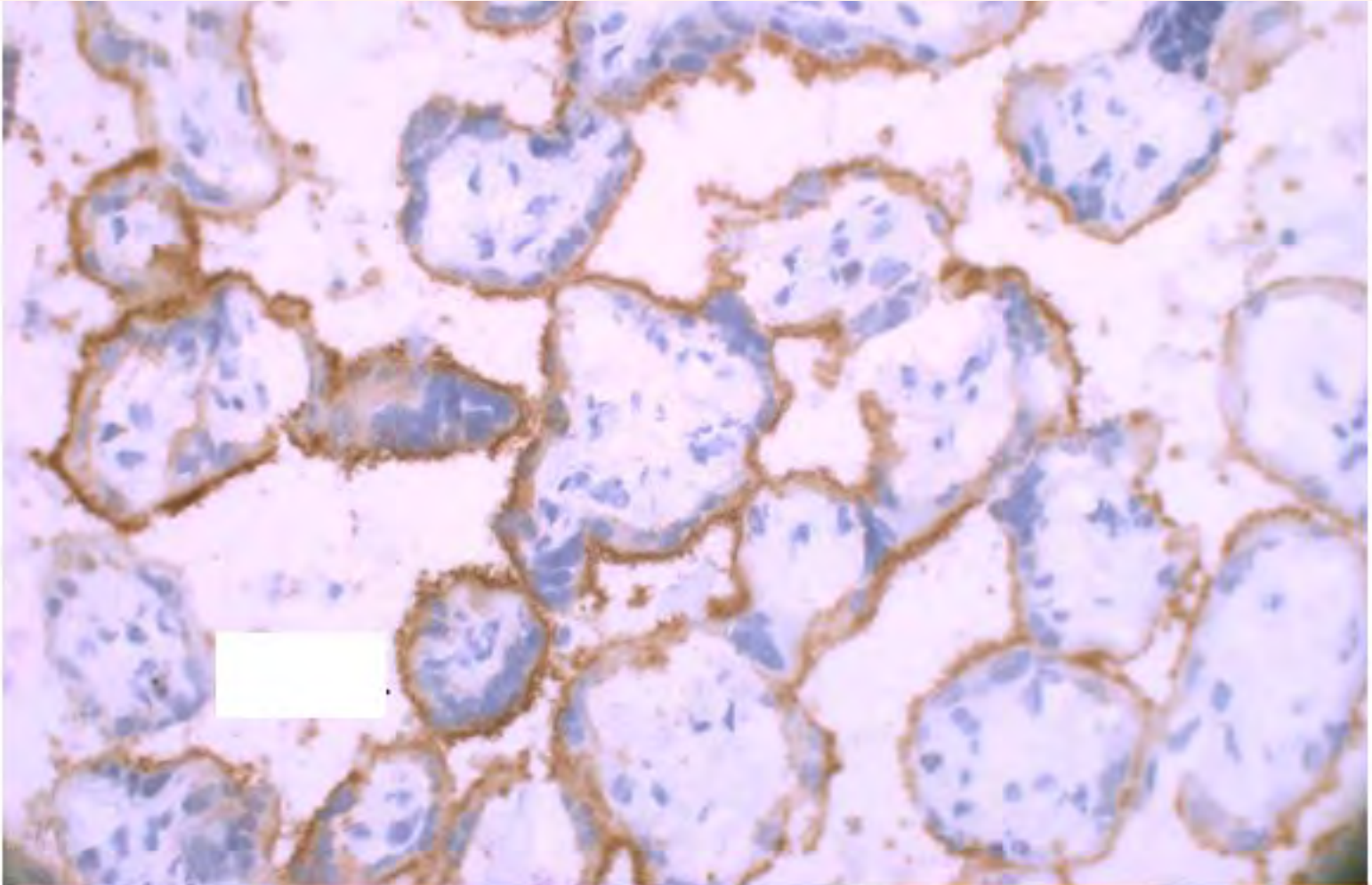


Chorionic Villi have abundant blood vessels and stroma contains macrophages (Hofbauer cells)

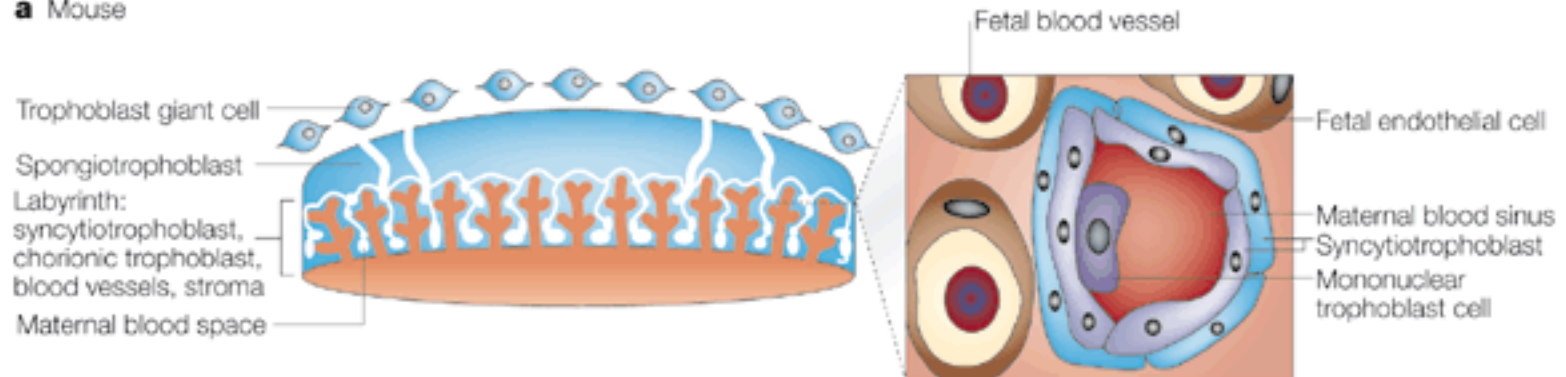


# Histology of Human Placenta, IHC

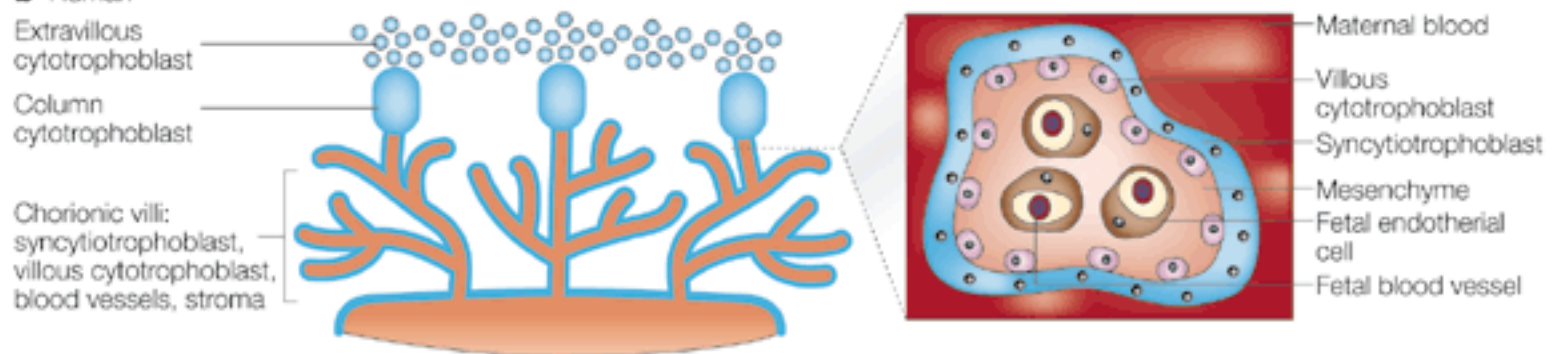
Chorionic Villi sections show trophoblast which is composed of an inner Cytotrophoblast and an outer multi-nucleated Syncytiotrophoblast.



**a** Mouse



**b** Human



Nature Reviews | **Genetics**

FIGURE 2 | Comparative anatomy of the mouse and human placenta.  
Placental development: Lessons from mouse mutants Janet Rossant & James C. Cross Nature Reviews Genetics 2, 538-548 July 2001



Placenta (2002), 23, 3–19

doi:10.1053/plac.2001.0738, available online at <http://www.idealibrary.com> on **IDEAL**<sup>®</sup>

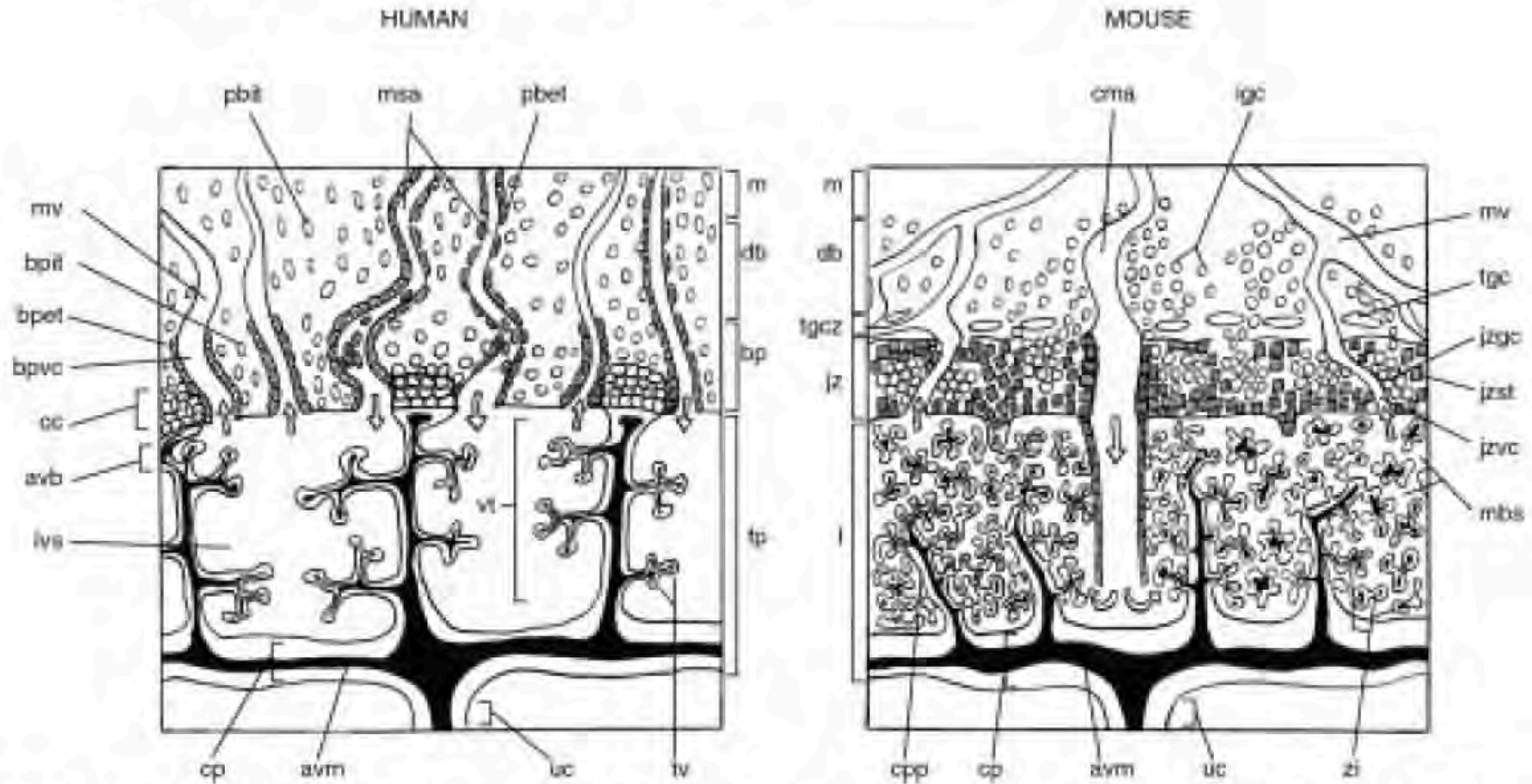
## **CURRENT TOPIC**

# **Comparative Developmental Anatomy of the Murine and Human Definitive Placentae**

**P. Georgiades<sup>a</sup>, A. C. Ferguson-Smith and G. J. Burton**

Department of Anatomy, University of Cambridge, Downing Street, Cambridge, CB2 3DY, UK

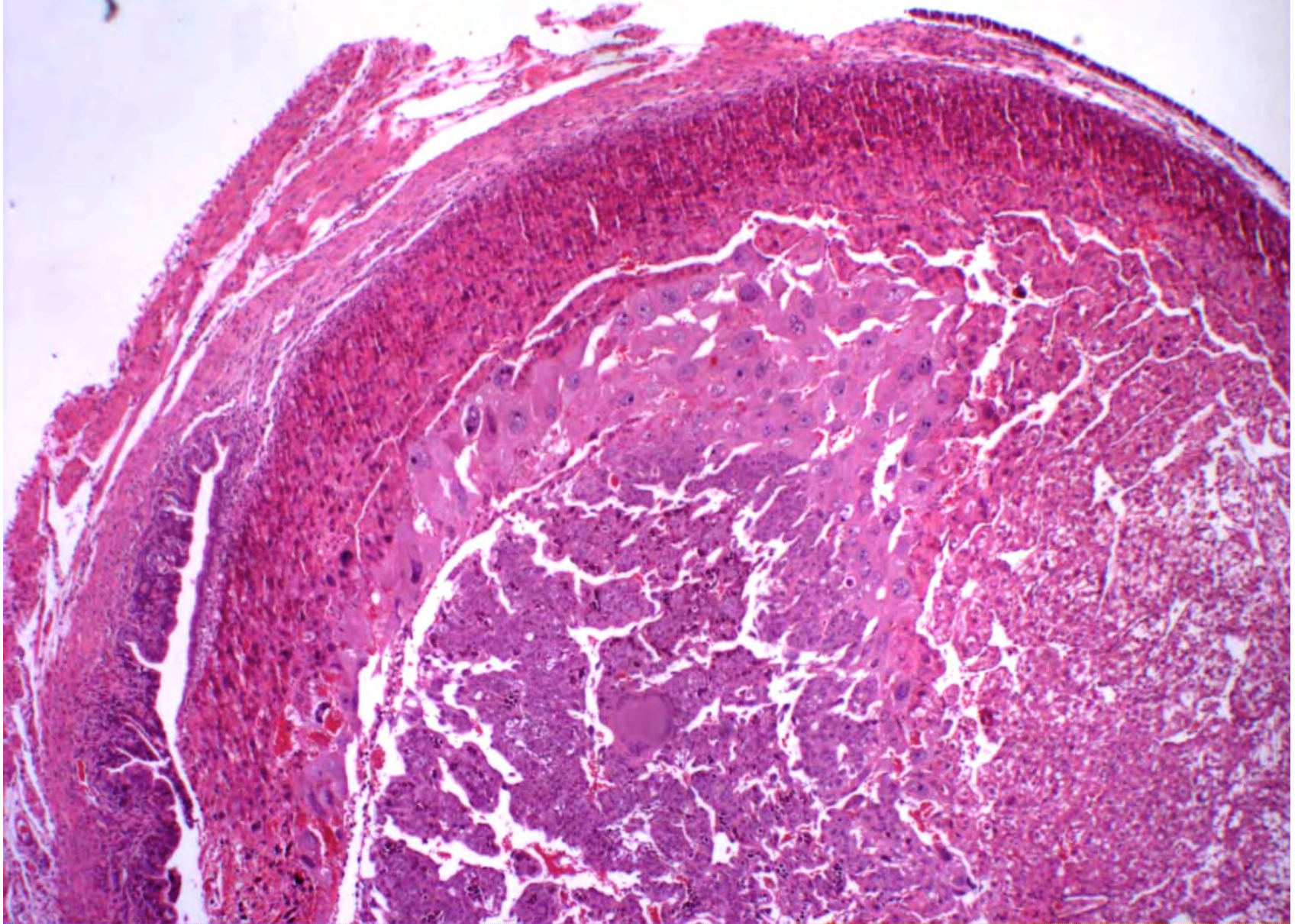
Paper accepted 28 August 2001



**Figure 1.** A simplified diagrammatic representation of the major regions and some cell types of human and murine placentae during the third trimester and the last fifth of gestation respectively. This is a cross-sectional view with the plane of sectioning being perpendicular to the flat (fetal) surface of the placenta. The side of the placenta facing the mother (maternal side) is at the top of the picture and that facing the fetus (fetal side) at the bottom. Note that one way of distinguishing the extent of the basal plate (*bp*) of the human placenta is the extent of endovascular trophoblast lining the lumen of maternal veins (*mv*). Although it is impossible to show in two dimensions, the zona intima (*zi*) regions of the murine labyrinth (*l*) are actually maze-like ramifications of the main chorionic plate projections (*cpp*). In both species, the venous and arterial channels of the junctional zone (*jz*) and basal plate (*bp*) are lined by endovascular trophoblast. The same applies to the labyrinthine part of the murine central maternal artery (*cma*). Arrows depict the direction of maternal blood flow within the utero-placental circulation. *avb*, anchoring villous branch; *am*, allantoic vasculature and mesoderm; *bp*, basal plate; *bp*, basal plate; *endovascular trophoblast*, *endovascular trophoblast*; *int*, basal plate, interstitial

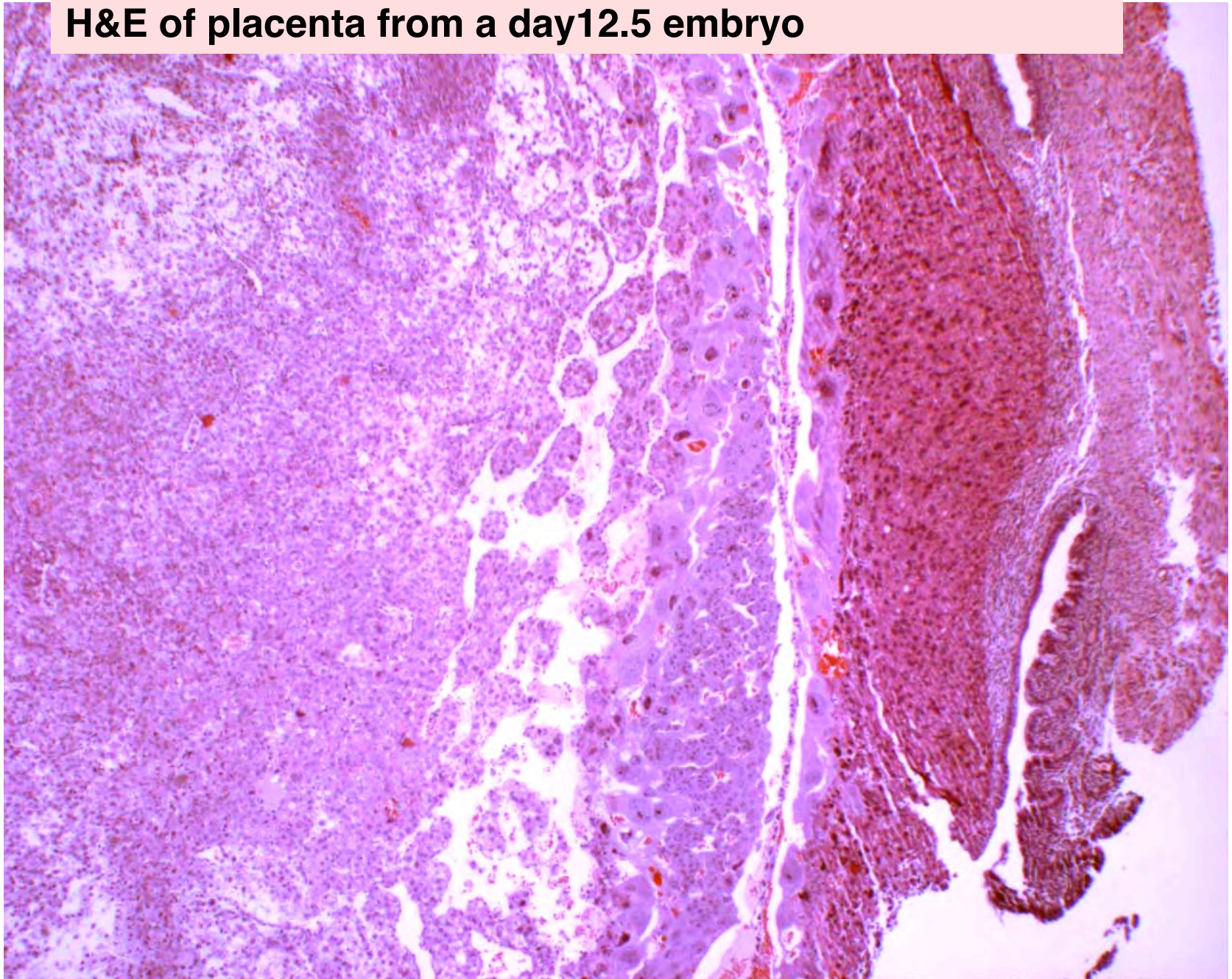


□ **H&E of placenta from a day12.5 embryo**

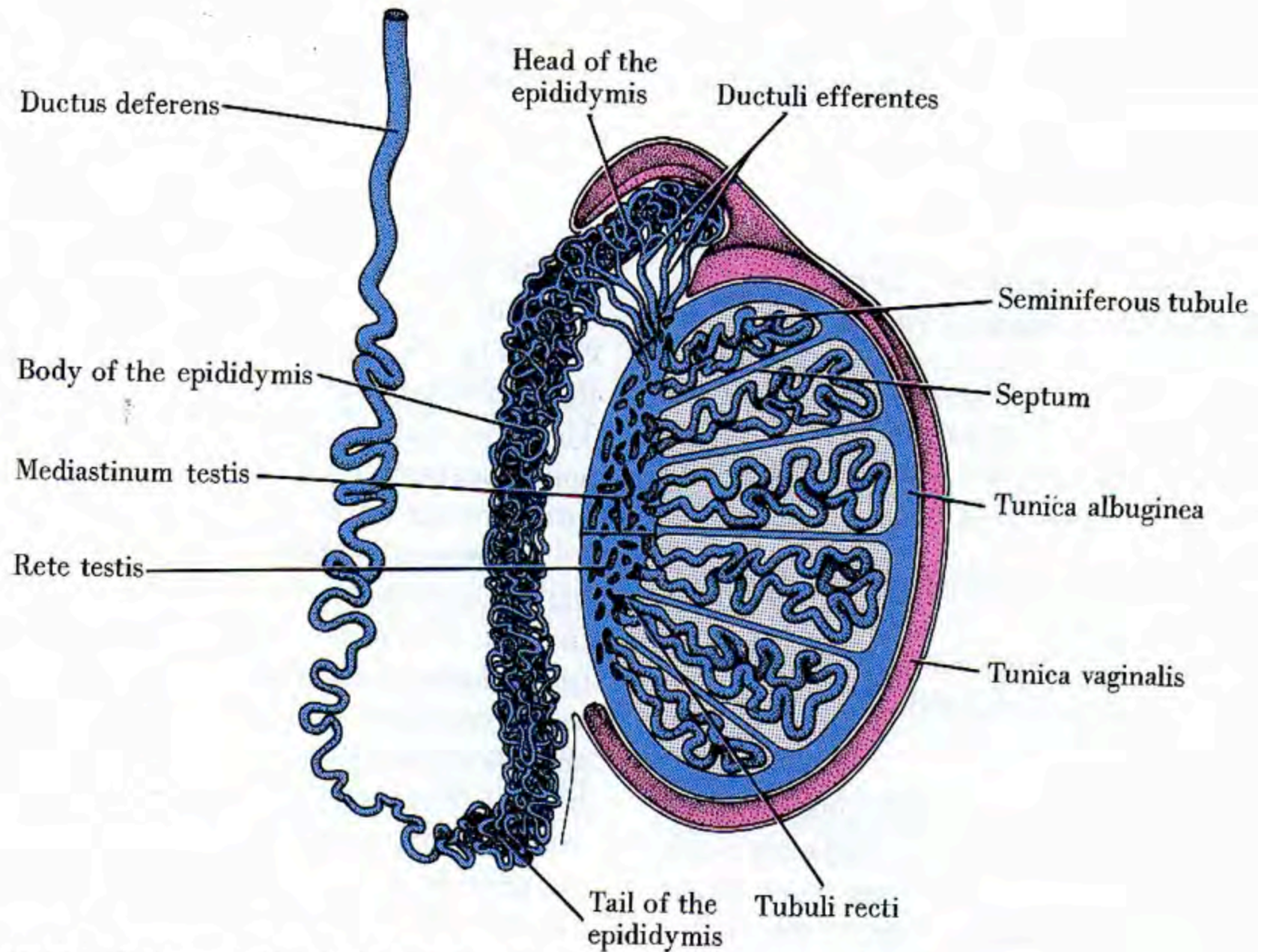




# H&E of placenta from a day12.5 embryo

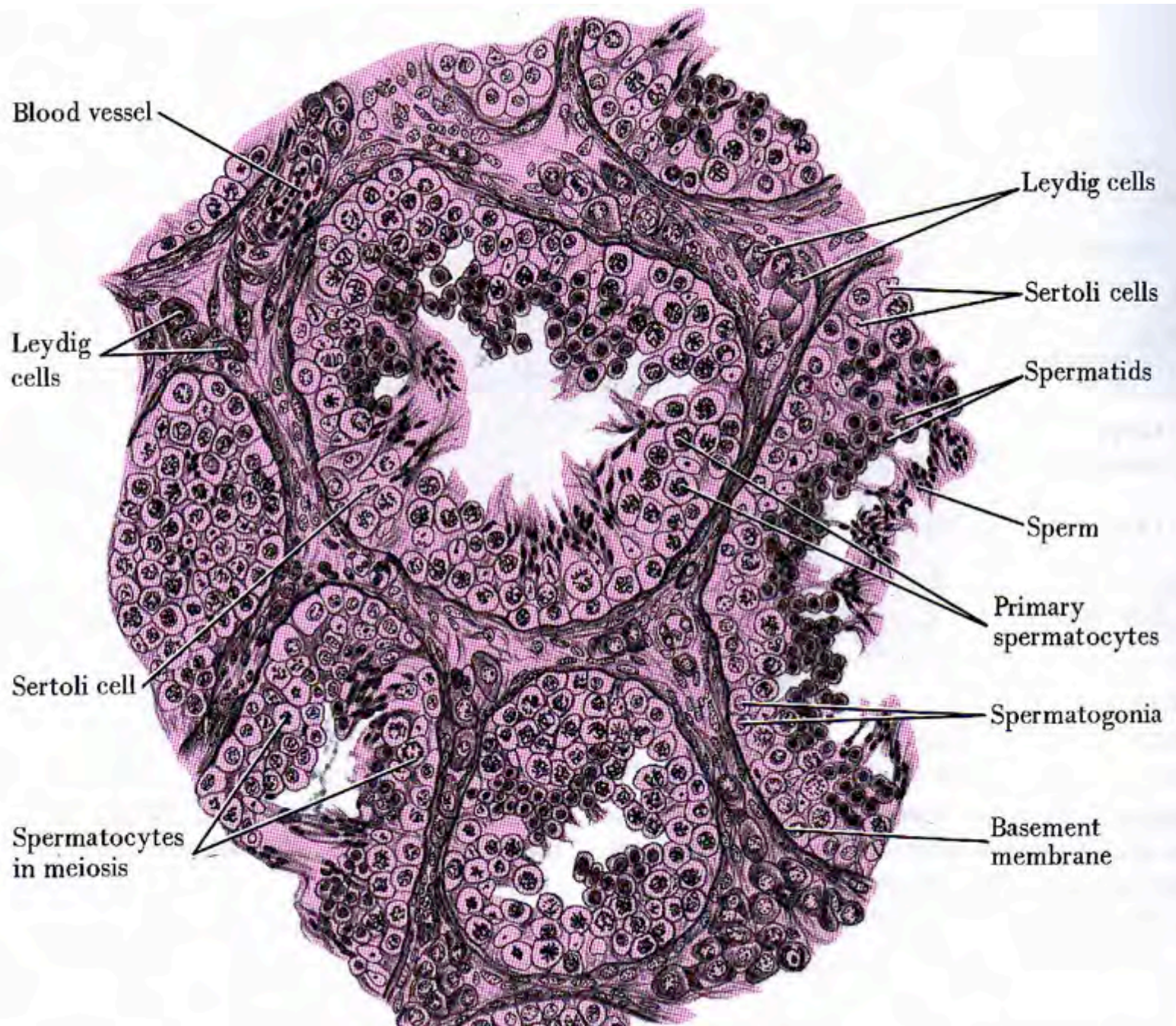






Schematic diagram of a longitudinal section







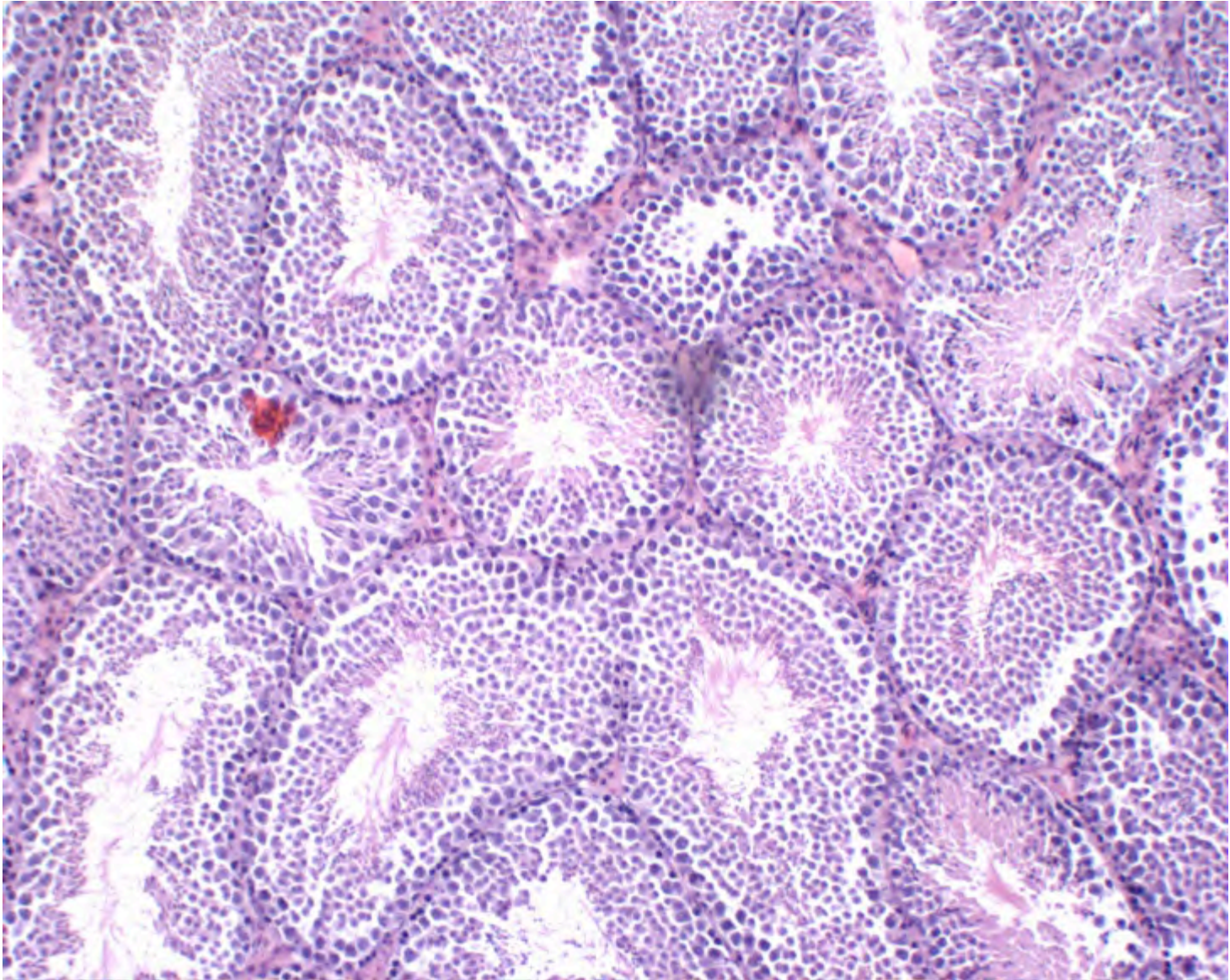
TESTIS: site of the development of spermatozoa and the production of testosterone. Spermatogenesis and testosterone production are controlled by FSH and LH (ICSH -- Interstitial Cell Stimulating Hormone).

**Seminiferous tubule** - site of spermatogenesis

**Interstitial cells (Leydig cells)** are in the stroma, between the seminiferous tubules, and are the site of testosterone production

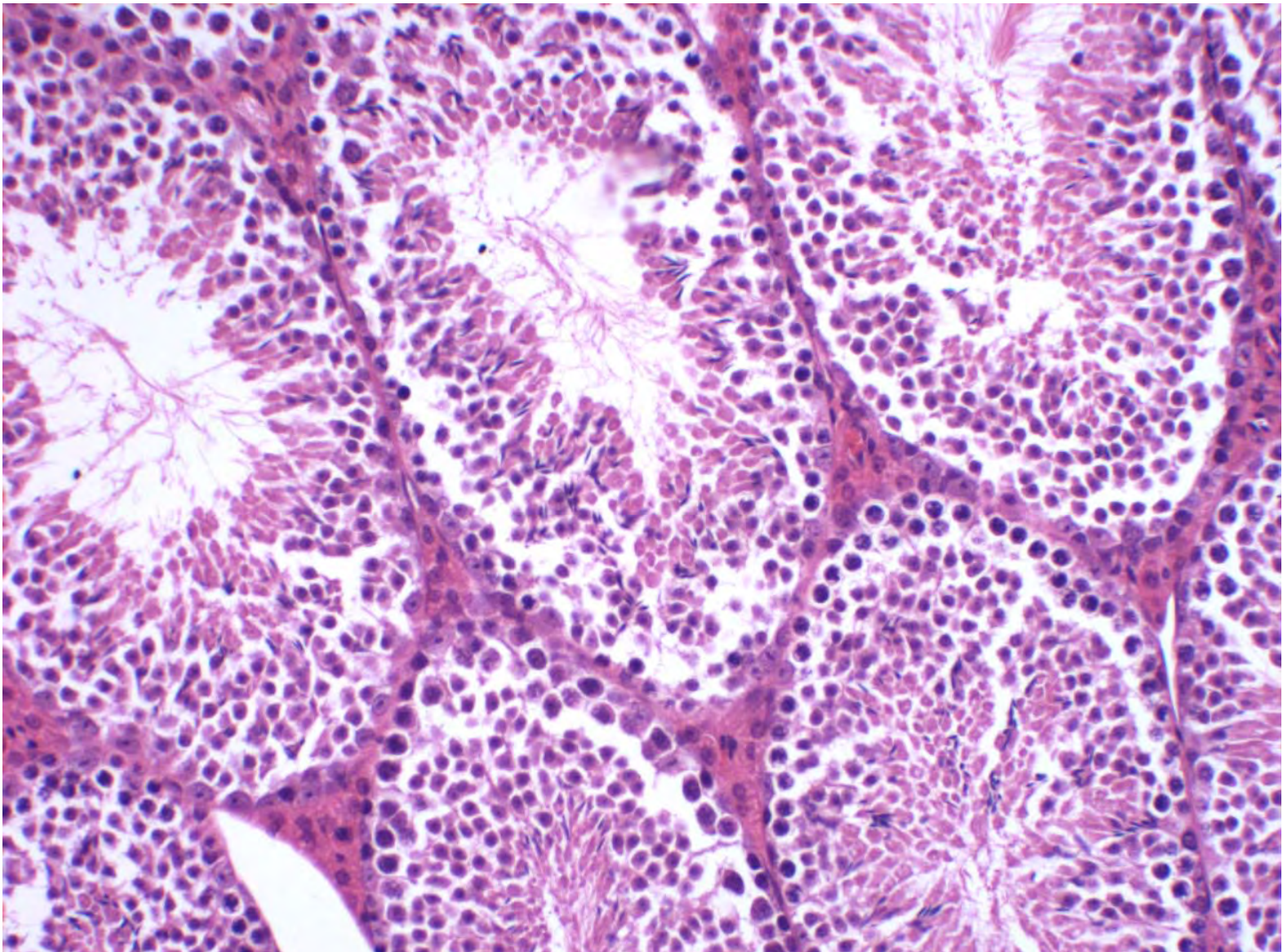
## ● Need-to-know

- The germ cell types, from basal location (immature) to luminal location (mature), are, respectively, spermatogonia, spermatocytes, spermatids, and spermatozoa in both the mouse and the human.
- In the mouse, it takes approximately 35 days for spermatogonia to develop into spermatozoa.
- In the human, it takes approximately 70 days for spermatogonia to develop into spermatozoa.



**H&E of a section of mouse testis**



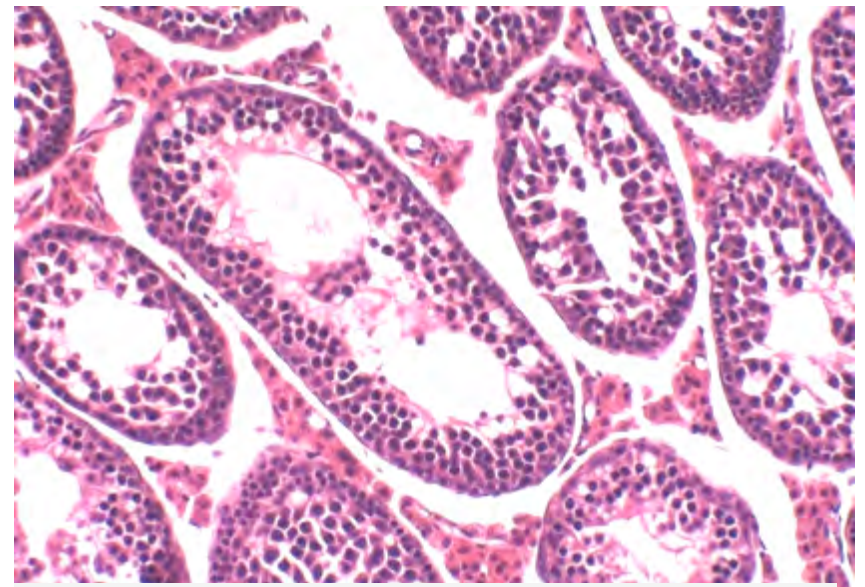
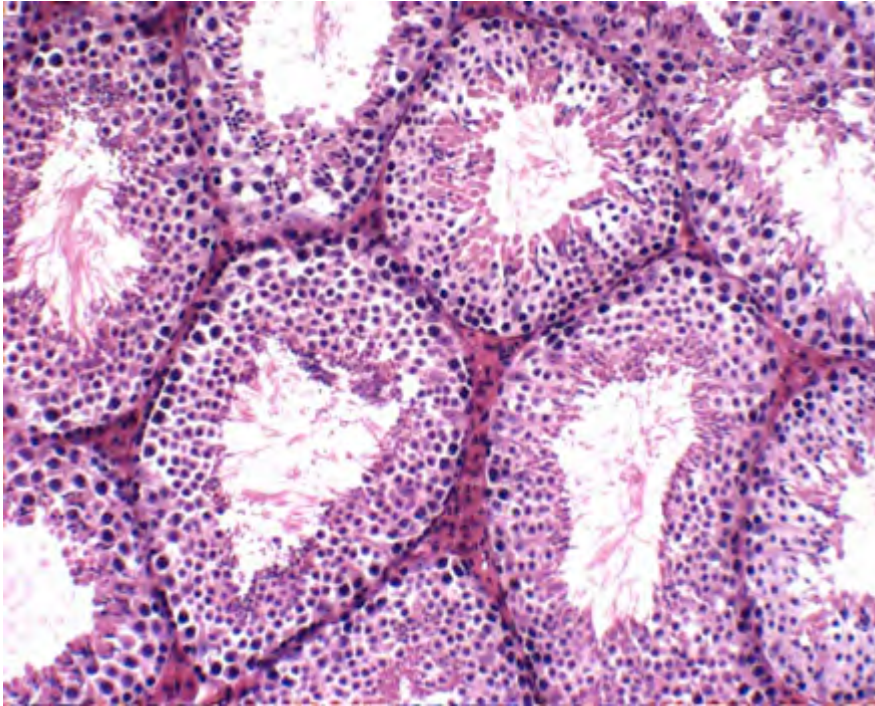


**H&E of a section of mouse testis (not fixed well)**

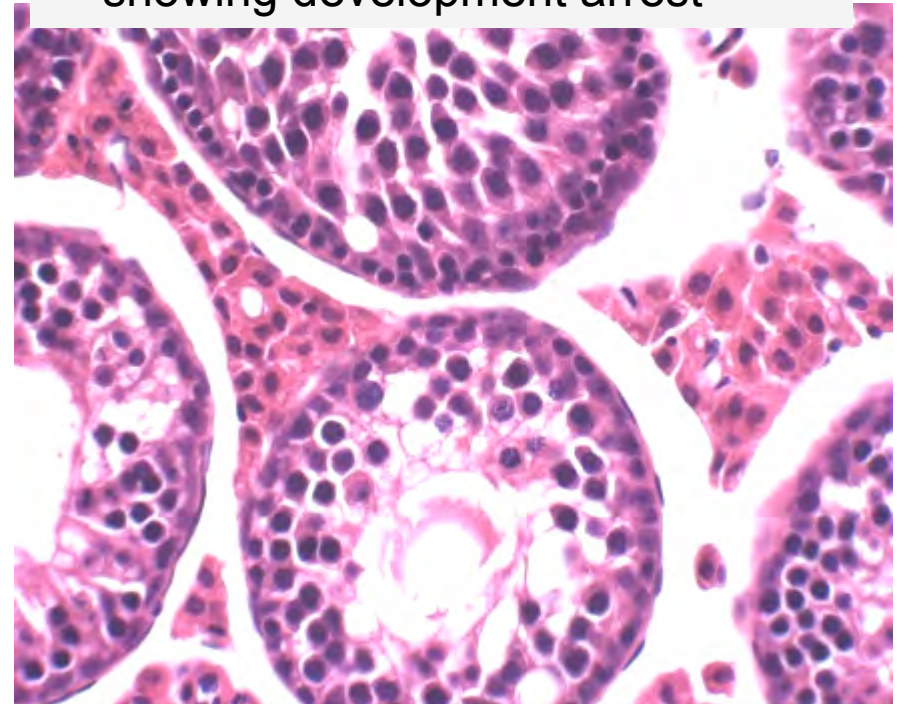
**If no IHC needed, fix in Bouin's for 6 hours and wash in 70% alcohol until clear and process into paraffin**



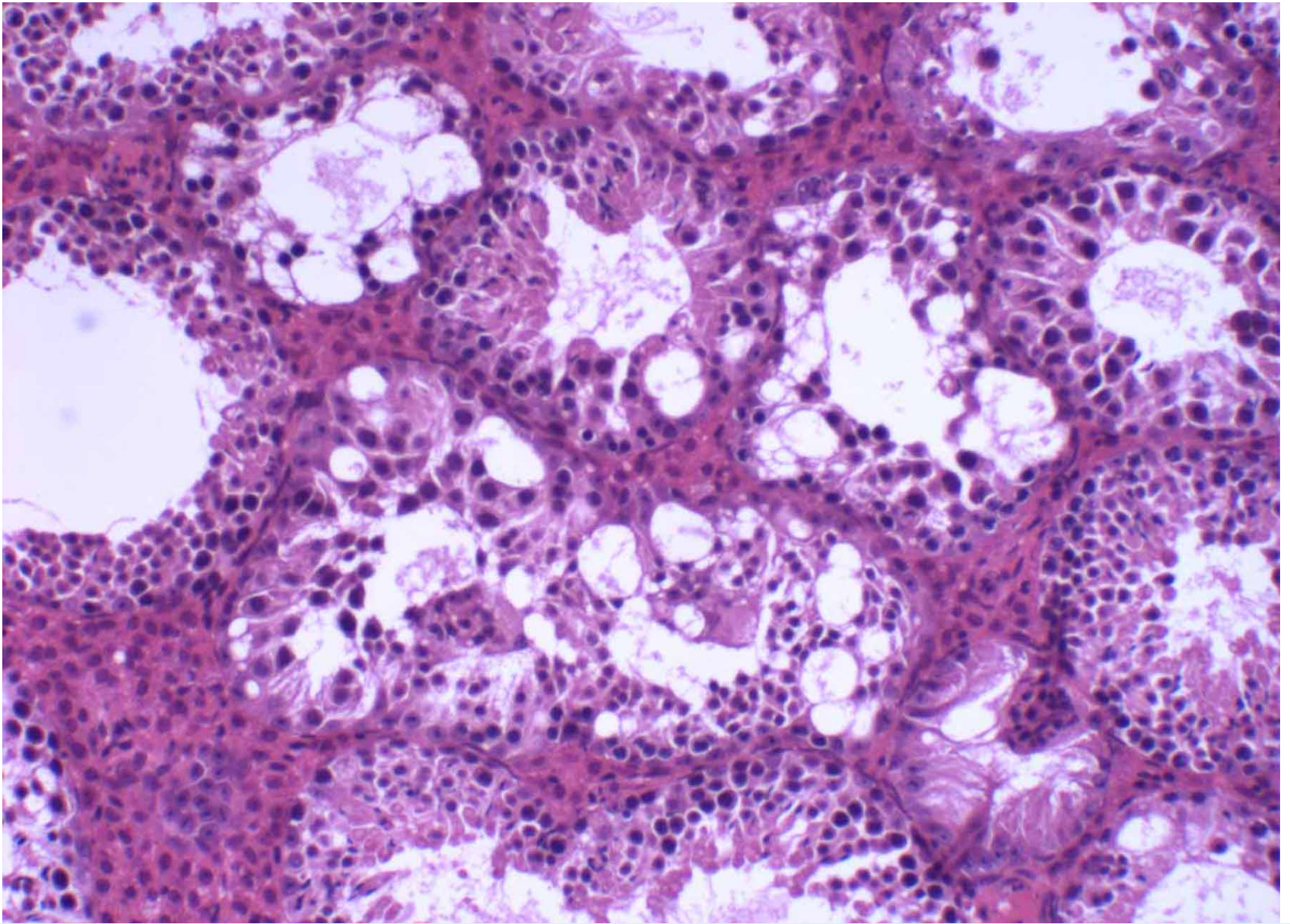
Seminiferous tubules with maturing sperm



Sections of testes from --null animal showing development arrest



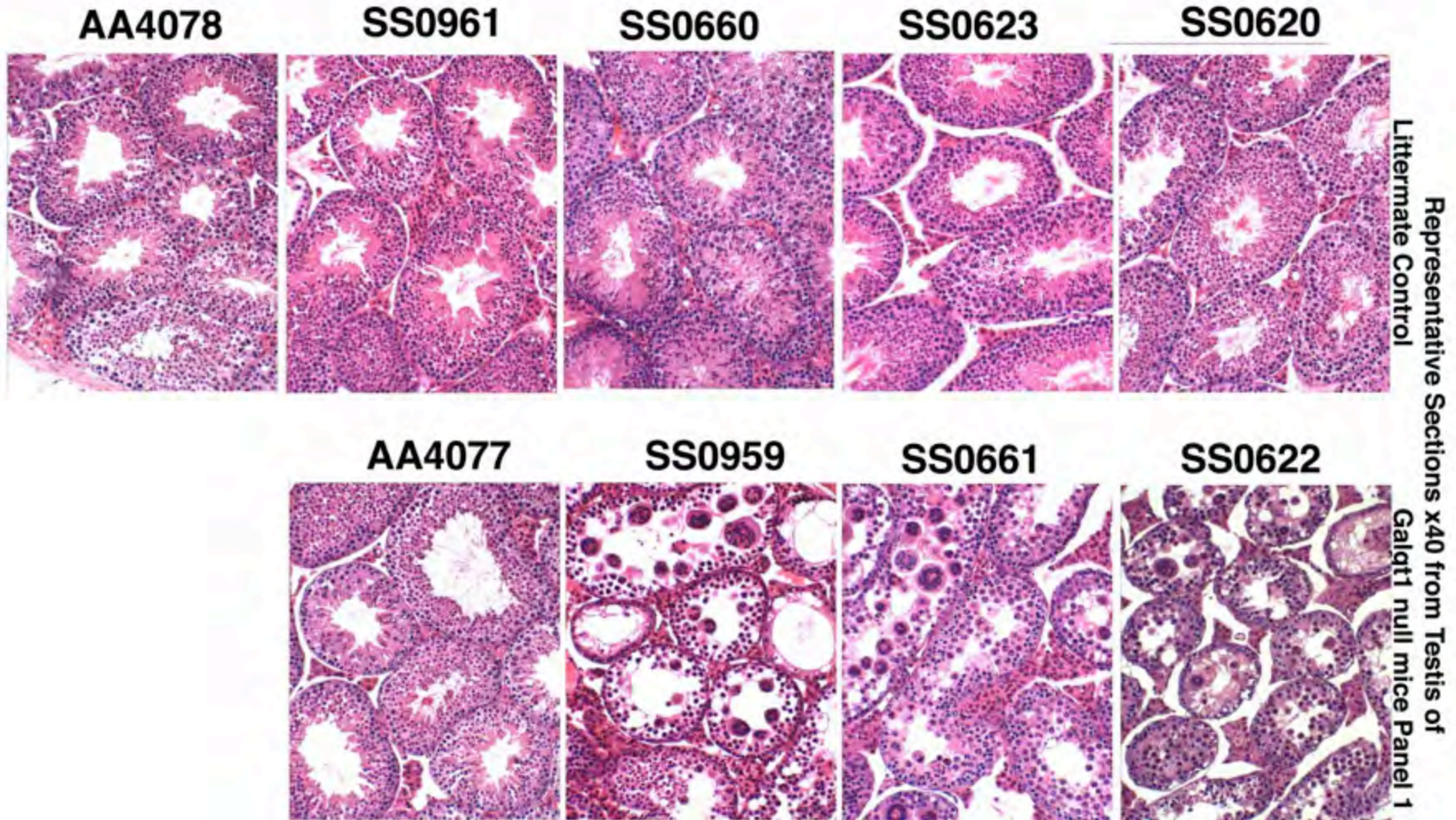




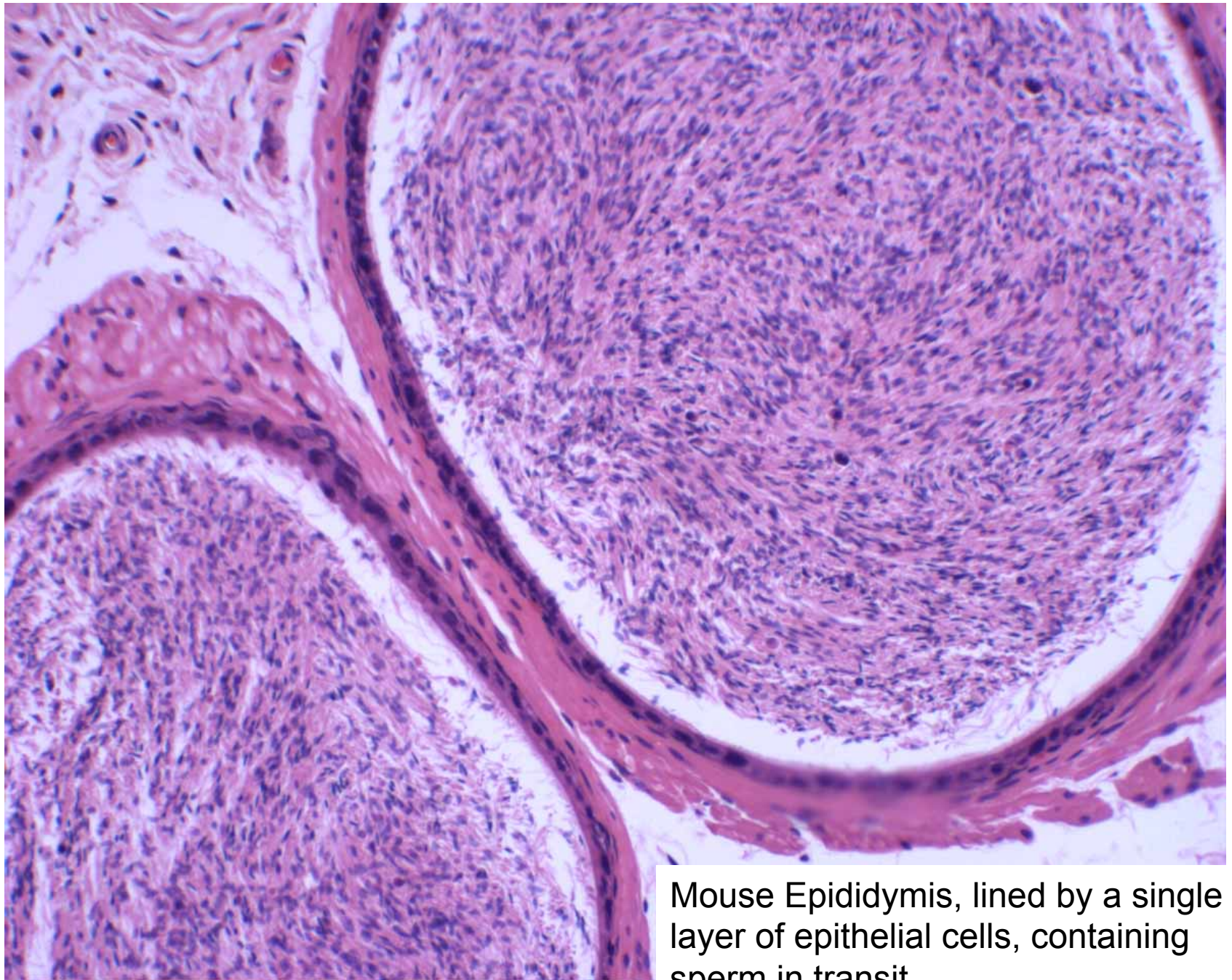
Abnormal degeneration of cells lining seminiferous tubules of mouse testis



**H&E of testes from littermate controls and from ---null mice, showing degenerated follicles**

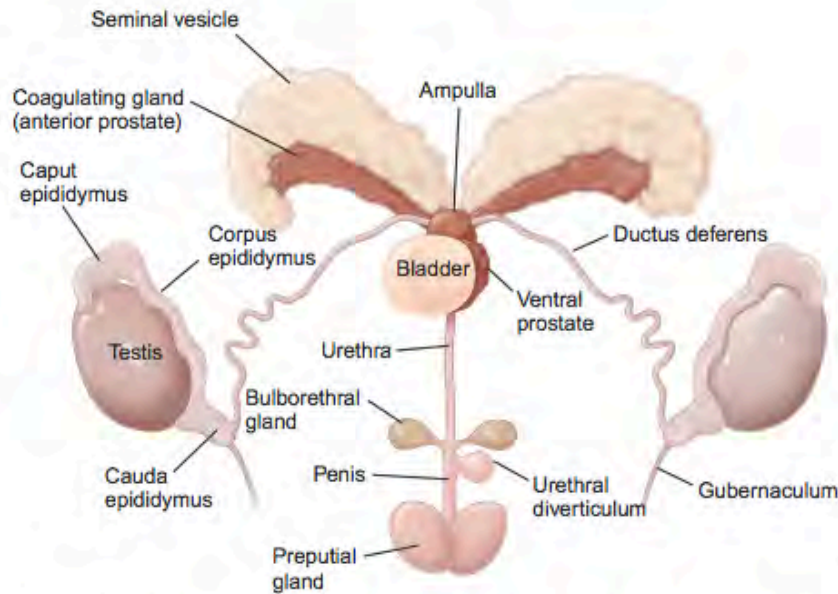






Mouse Epididymis, lined by a single layer of epithelial cells, containing sperm in transit

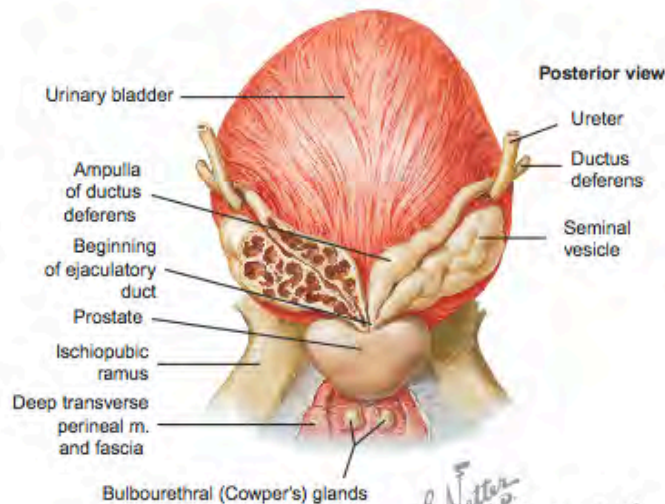




### ● Need-to-know

- Mice have ampullary and preputial glands.

**FIGURE 1 The male mouse reproductive system.** The reproductive tract consists of paired testes, epididymis, vas deferens, and the accessory sex glands which consist of paired seminal vesicles, prostate lobes, ampullary glands, bulbourethral glands, preputial glands, and the penis. Source: Adapted with permission from Anatomy of a Laboratory Mouse, Cook, M.J., 1965, with permission from Elsevier.

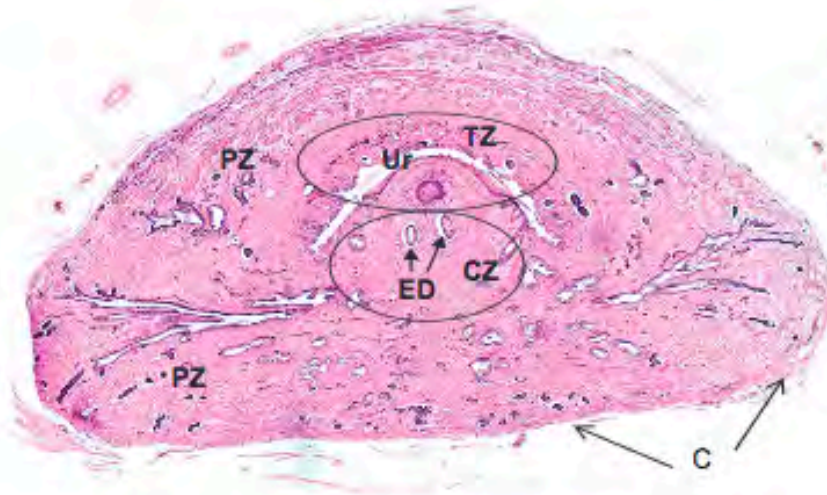


### ● Need-to-know

- Humans lack ampullary and preputial glands.

**FIGURE 2 The male human reproductive system.** The reproductive tract consists of paired testes, epididymis, vas deferens, seminal vesicles, bulbourethral glands, prostate gland, and the penis. Source: © Elsevier, Inc., www.netterimages.com.

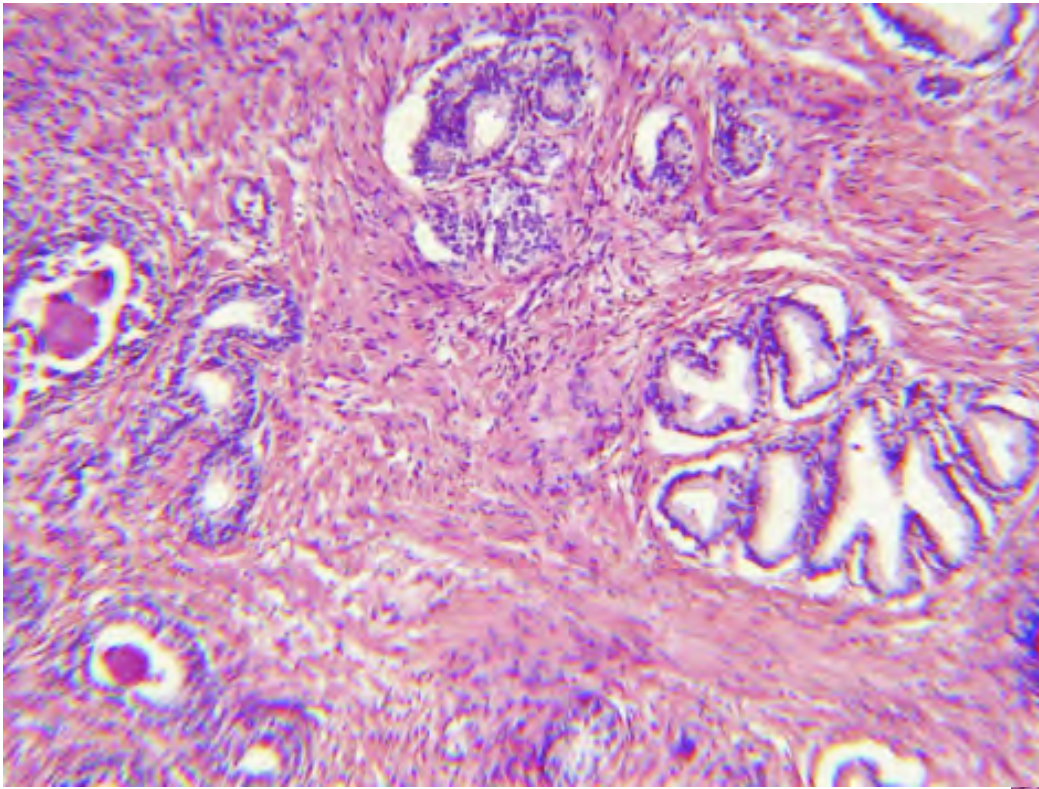




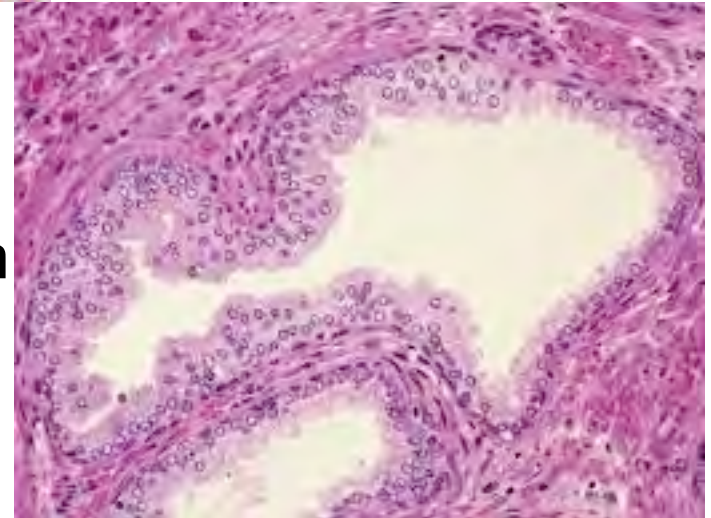
### ● Need-to-know

- The mouse prostate is divided into four histologically distinct lobes.
- The human prostate is divided into zones that are not clearly demarcated from one another but may be characterized by specific predisposition to disease.
- The human prostate is surrounded by a capsule, whereas the mouse prostate. Lobes are invested and separated by a thin mesothelial-lined capsule/membrane.

**FIGURE 27 Human prostate.** The prostate is alobular, but it is divided into three concentric zones. The central zone (CZ) encircles the ejaculatory ducts (ED), the transition zone (TZ) surrounds the prostatic part of the urethra (Ur), and the peripheral zone (PZ) makes up the bulk of the gland. The human prostate is surrounded by a capsule (C).

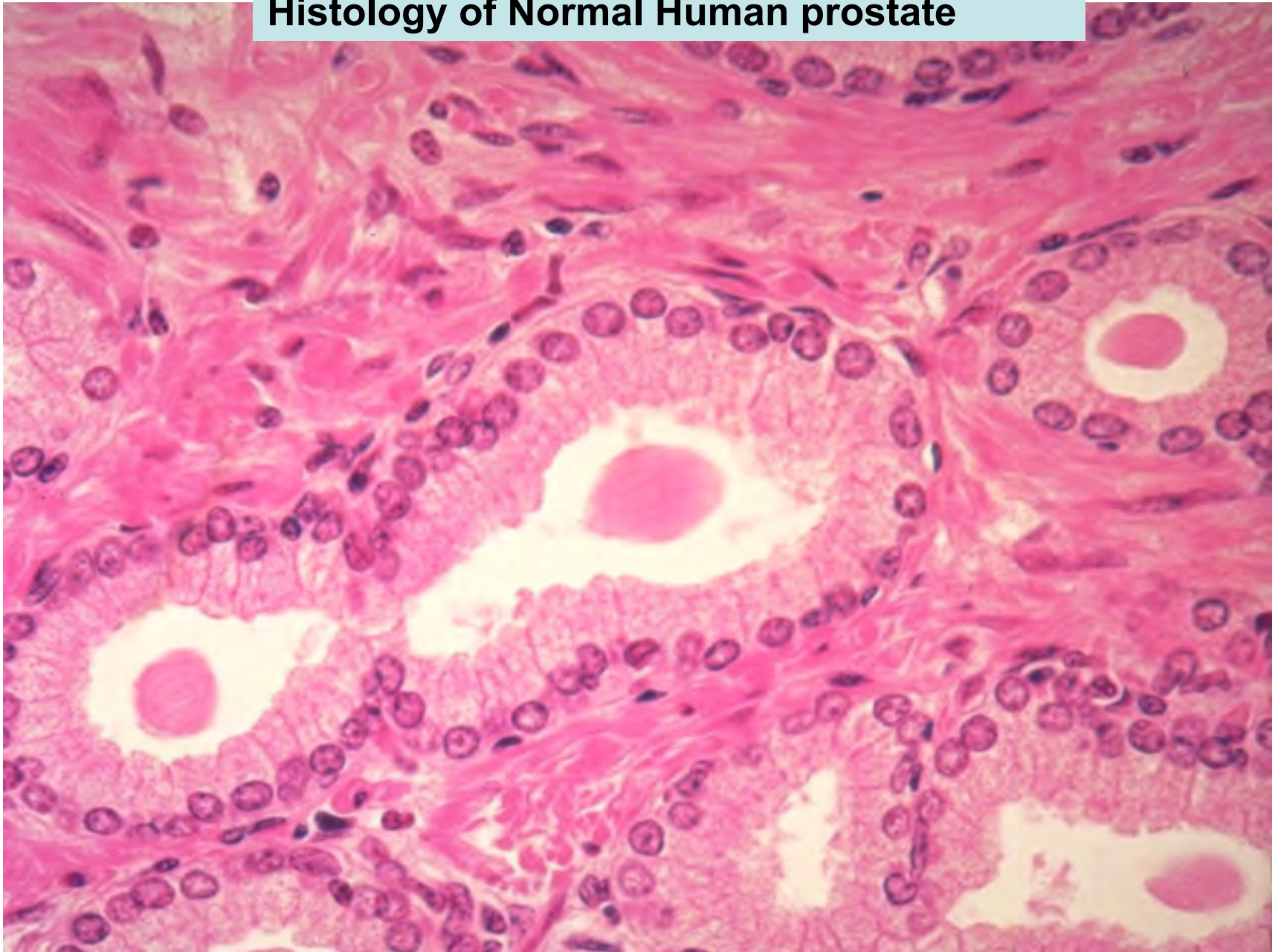


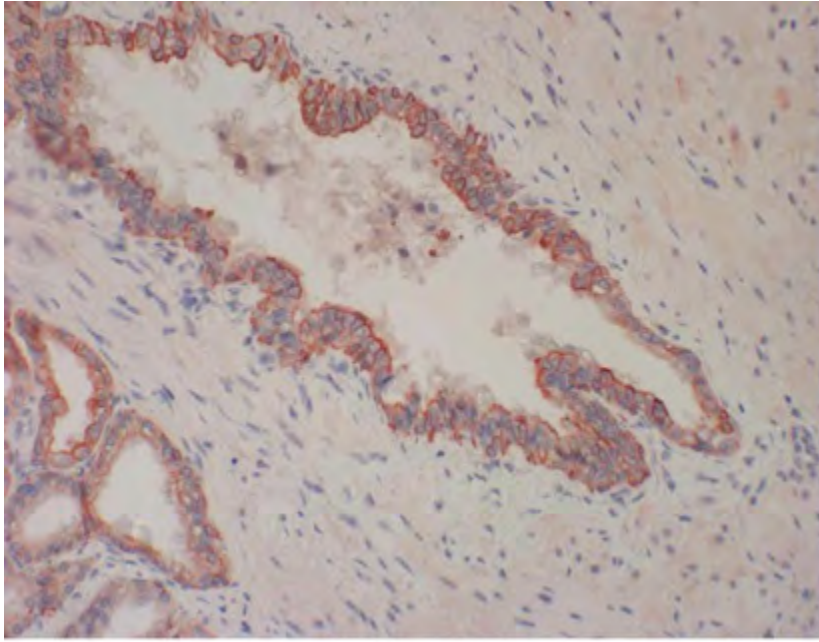
Human prostate:  
Tubulo-alveolar glands embedded in  
fibro-muscular stroma



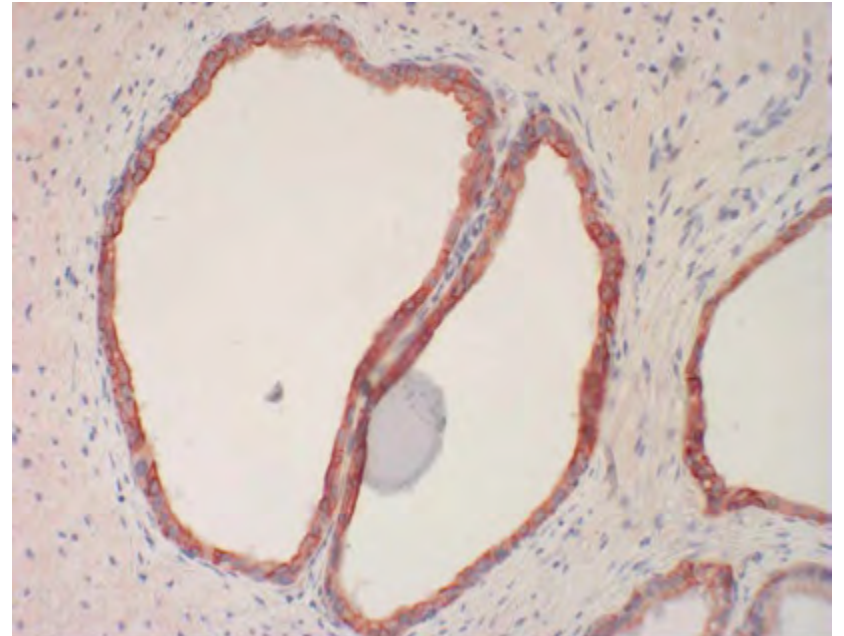


## Histology of Normal Human prostate





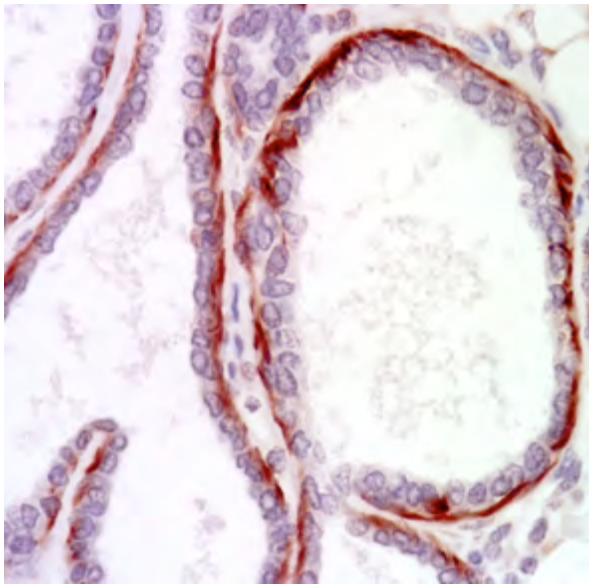
Normal Human Prostate  
Single layer of epithelial cells  
marked with anti-keratin



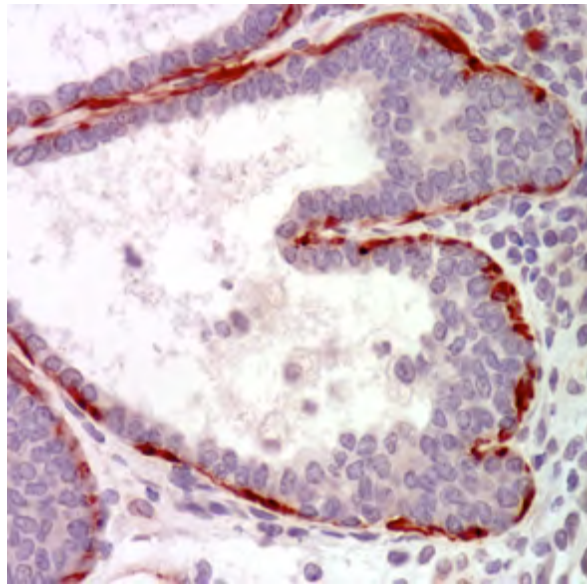


# Markers of Differentiation

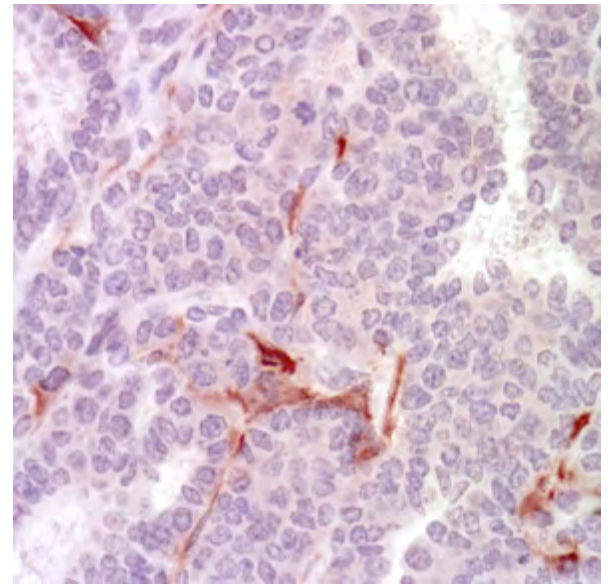
Smooth Muscle Actin (SMA)



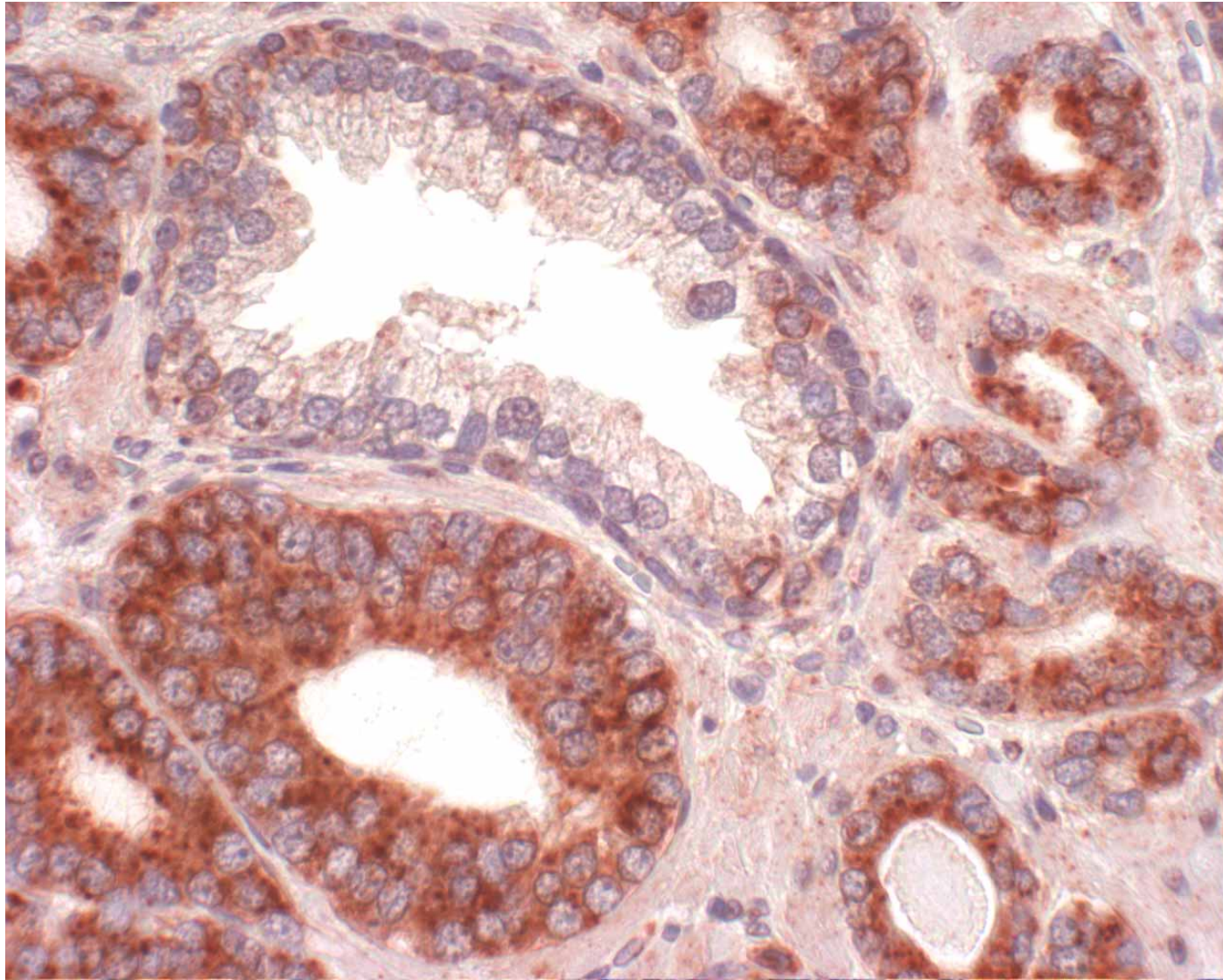
Normal



Hyperplasia



Tumor



Prostate carcinoma around a normal prostate duct, with IHC (?prostate specific acid phosphatase PSA) marker



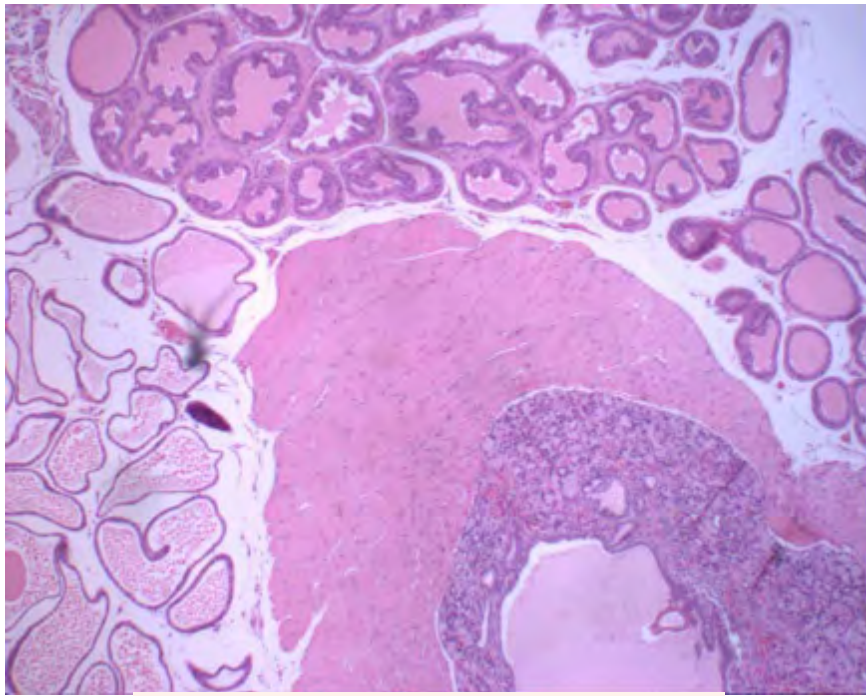
[http://  
tvmouse.ucdavis.edu/  
prostate/mouse/  
prostateindex2.htm](http://tvmouse.ucdavis.edu/prostate/mouse/prostateindex2.htm)



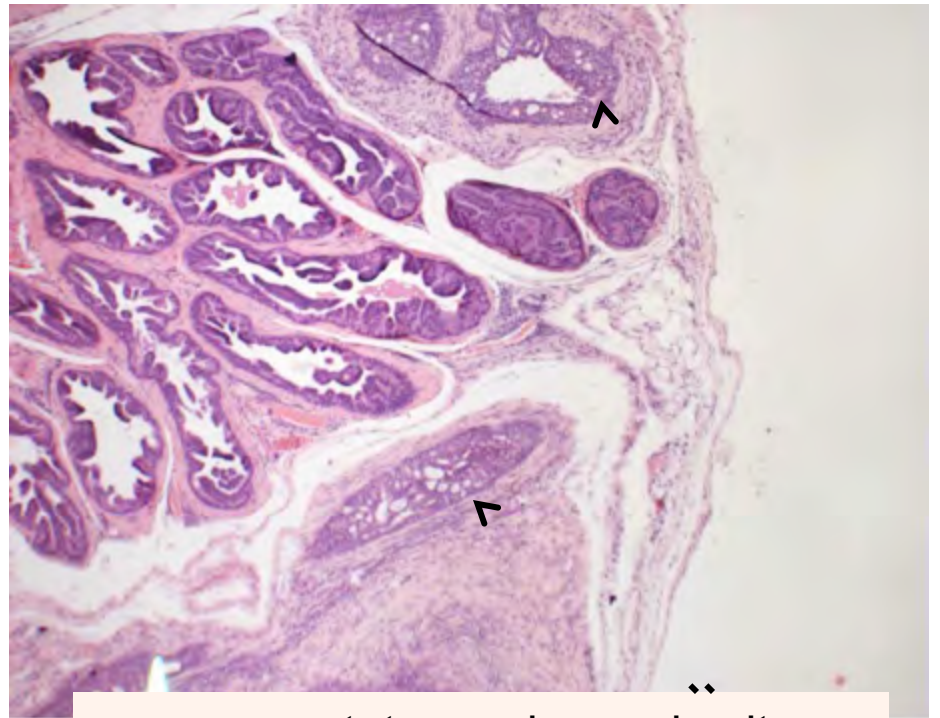


Mouse prostate





Normal mouse prostate

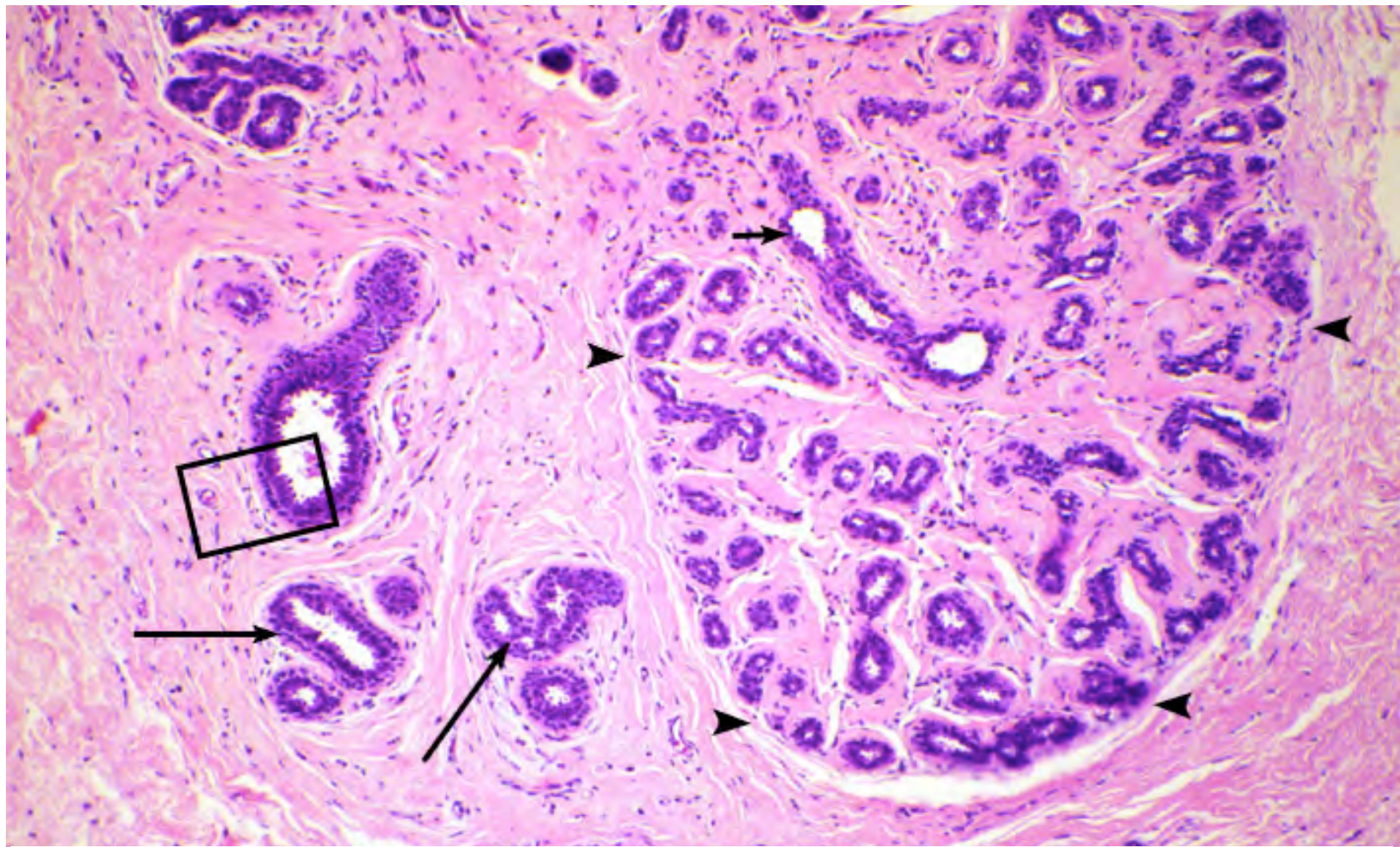


mouse prostate carcinoma in situ



Mouse  
prostate  
early  
carcinoma





Human breast lobules and ducts

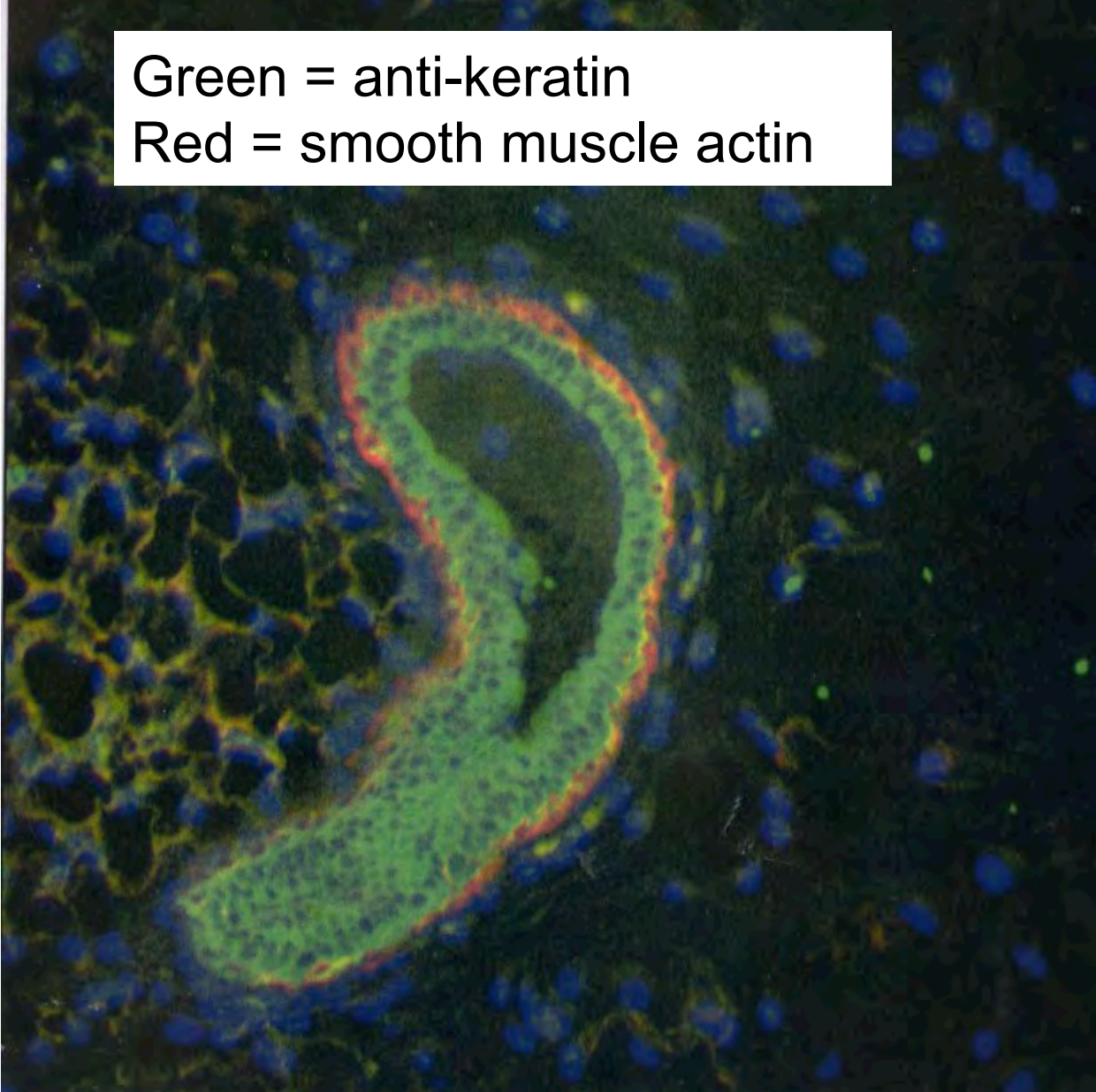
Normal with one layer of lining epithelial cells and an outer layer of myoepithelial cells



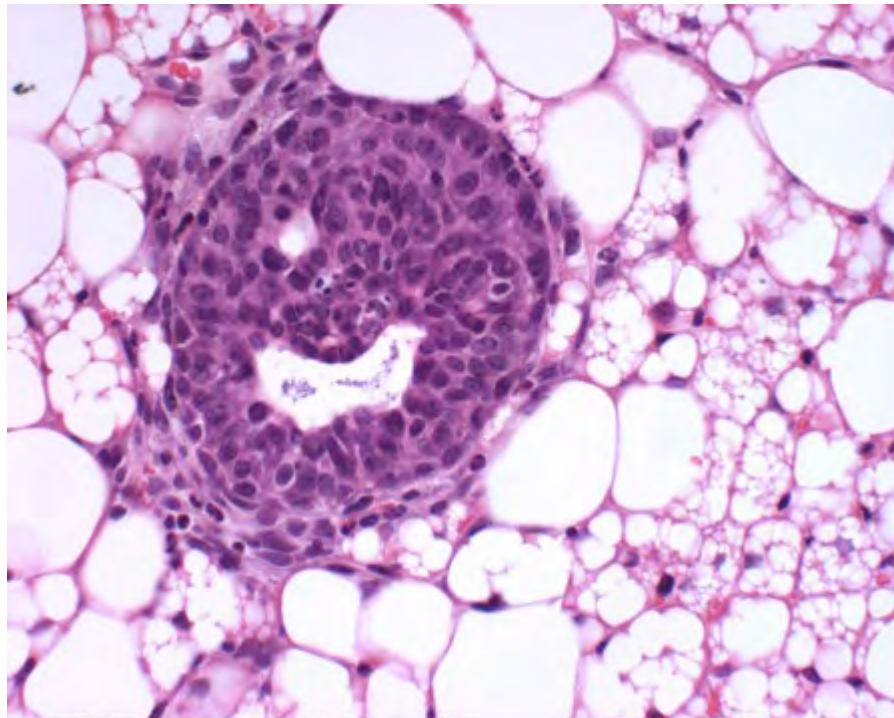
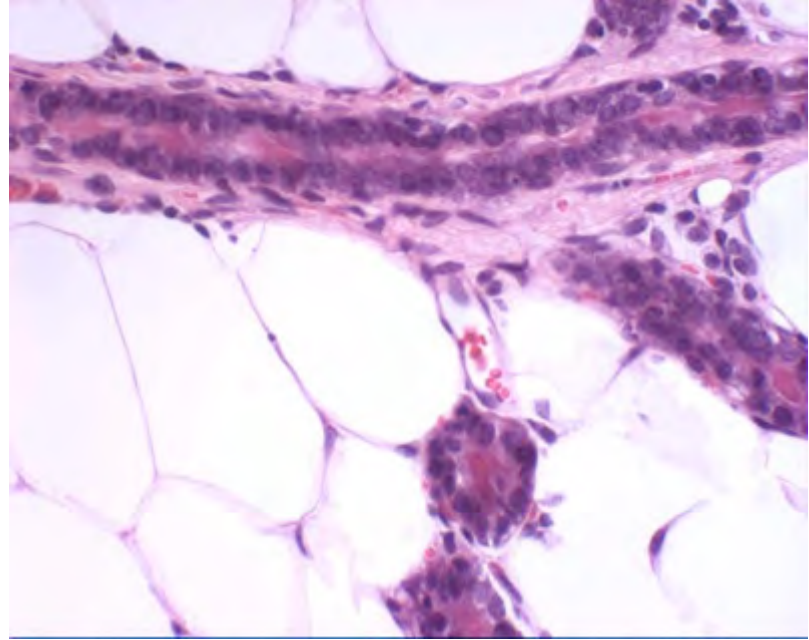
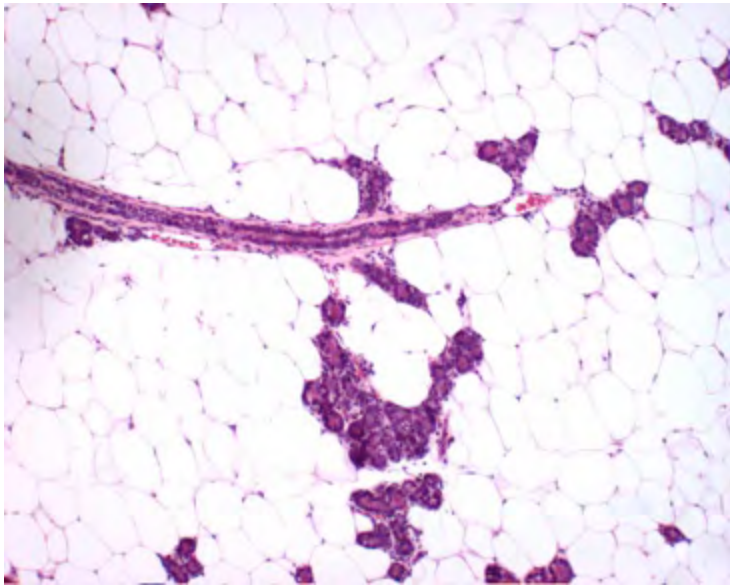
Normal breast ducts and alveoli have an Inner layer of cuboidal epithelial cells and an Outer layer of myoepithelial cells

Green = anti-keratin

Red = smooth muscle actin







Normal  
mammary  
gland and  
carcinoma  
in-situ

